Screening of Tomato Genotypes for Resistance to Early Blight (*Alternaria Solani*)

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Abstract: Early blight disease caused by Alternaria solani has been a serious problem in tomato growing areas particularly at humid tropical, subtropical and temperate regions of the world. An investigation was carried out at Horticulture Experimental farm, AAU Jorhat with forty-five genotypes of tomato. The experiment was conducted consecutively for two years during Rabi seasons of 2012-13 and 2013-14 in replicated trails. The site of the experiment was a sick plot where tomato was grown continuously for three years before testing the genotypes. Recommended package of practices were followed to raise the crop. Blight incidence was evaluated from the disease symptoms using Percent Disease Index (PDI) after the genotypes were artificially inoculated with isolates of A. Solani. The genotypes Sel-35 (TLBRH-6 X Konbilahi) and Sel-19 (TLBRH-6 X Konbilahi) were highly resistant, 7 were resistant, 14 were moderately resistant, 16 were susceptible and 6 were highly susceptible. The result was found to be similar for both the years. The loss in yield due to the disease ranged from 2.15% in highly resistant genotype to 42.75% for highly susceptible genotypes to high yielding genotypes.

Keywords: Tomato genotypes, screening, Alternaria solani, resistance.

1. Introduction

Tomato (Solanum lycopersicum L.) belongs to the family Solanaceae and is nutritionally a good crop and a source of vitamin A, vitamin C and minerals. Tomato is affected by various diseases caused by fungi, bacteria, viruses, nematodes and also abiotic factors (Balachand, 1992). Early blight caused by the pathogen Alternaria solani is an economically important disease of tomato worldwide including India. The causal organism responsible for early blight is air borne and soil inhabiting (Datar and Mayee, 1981). Warm temperature and extended periods of leaf wetness from dew, rain fall and crowded plantation is favourable for development of this disease. Disease symptoms appeared on all above ground parts of plants particularly leaves, stem, petiole, flowers and fruits (Pandey et al., 2002). The yield loss due to early blight has been increasing with the incidence of this disease becoming more prominent with the change in environmental conditions. Early blight can cause loss to the extent of 78% in fruit yield (Singh, 1985; Datar and Mayee, 1981). Under severe epiphytotics loss of fruit may be as high as 95% (Sridhara and Naik, 1983). Disease severity of 90 per cent was recorded in Indo-Gangetic region of the country (Pandey et al., 2002). The use of chemicals and fungicides could effectively control the disease but repeated used of this chemicals pollutes the environment and caused health hazards to human beings. Thus, screening of genotypes resistant against early blight would be an effective measure to reduce the dependency on fungicides and chemicals. Thus, identification and utilization of genetic resources resistant to A. solani in tomato is the only way to develop early blight-resistant tomato cultivars following appropriate breeding methods. Field evaluation after inoculation of the pathogen isolates is most utilized method for screening of tomato genotypes for early blight resistance. In the present study, the objective was to identify and screen sources of resistance against early blight in tomato genotypes.

2. Materials And Methods

The experiment was conducted Horticultural at Experimental Farm, AAU, Jorhat-13 to screen out the genotypes for resistance to early blight during Rabi season 2012-13 and 2014-15 consecutively for two years in a Randomized Block Design. Forty five genotypes of tomato were used in this experiment which was sown in three rows with 2 replications and maintaining planting distance of 60cm between rows and 50cm between plants. Pathogen causing the disease early blight of tomato, Alternaria solani was isolated from diseased tomato leaves and was grown on Potato Dextrose Agar (PDA) medium. inoculums suspension pathogen was grown on PDA plates. The plates were incubated on 25°±1°C under a cool white fluorescent diurnal light with 12hr photoperiod for 10-15 days. Ten days old mycelia culture with thickening of conidiogenous hyphae and chlamydospore like structure were used for inoculation. The plants were inoculated 45 days after transplanting with a concentration of 157 cfu/ml during the screening process with a manual backpack sprayer. The disease severity was assessed on all leaves and scored on 0-5 points scale as suggested by Pandey et al. 2003 and percent disease incidence (PDI) was calculated following Mckinney (1923) formula. Later, the disease reaction based on PDI was recorded according to the scale given by Peteira et al. 2002. After 7 days of incubation, plants were individually evaluated for disease scoring in each genotypes using 0-5 disease scale as given by Pandey, 2003 which is described as 0=Free from infection, 1=One or two necrotic spots on a few lower leaves of plants, 2=A few isolated spots on leaves, covering nearly 5-10% of the surface area of the plant, 3=Many spots coalesced on the leaves, covering 25% of the surface area of the plant, 4=Irregular, blighted leaves and

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sunken lesions with prominent concentric rings on the stem, petiole, and fruit, covering 40-50% of the surface area, 5=Whole plant blighted, leaves and fruits starting to fall; foliar part free of disease. The Percent Disease Index (PDI) was calculated by the formula given by McKinney, 1923; Pandey *et al.*, 2003.

PDI = Sum of all ratings X 100

Total no. of observations X maximum rating grade

Disease reaction classes for early blight infection based on percent disease severity in tomato as given by Peteira *et al.* 2002.

Disease reaction	PDI range
Highly resistant	0-12.5
Resistant	12.6-25.0
Moderately resistant	25.1-37.5
Susceptible	37.6-50.0
Highly susceptible	50.1 and above

The yield per plant was recorded from disease free condition and disease infested condition differently. The value was converted to yield per ha. Later, the loss in yield due to disease incidence was calculated and converted into percentage.

3. Results and Discussion

The 45 genotypes were screened for resistance to early blight by inoculating the plant isolates with *A. solani* and the PDI values were recorded for both the years taken at the last reading as given in table-1. The reaction of the genotypes against early blight is shown in table-2 after their score obtained from the PDI values as in table-1.

The PDI ranged from 12.00% to 79.43% during 2012-13 and the value ranged from 11.68% to 77.53% during 2013-14. The PDI of highly resistant genotypes were 12.50% for Sel-35 and 12.00% for Sel-19 during 2012-13. Similar PDI values were obtained during 2013-14 with 12.30% for genotype Sel-35 (TLBRH-6 X Konbilahi) and 11.68% for Sel-19 (TLBRH-6 X Konbilahi). Resistant reaction was recorded in 2012/SPT/TOINDVAR-4, Sel-16, Sel-46, 2012/SPT/TODVAR-5, 2012/SPT/TODVAR-6, 2012/TOLCVRES-3 and Sel-9 from the readings of both the years. The genotypes showing highly susceptible reaction 2012/TOLCVRES-5 (79.43%) and 77.53%). were (75.32%) 2012/TOLCVRES-8 and 54.45%), (73.56%) 72.25%), 2012/TOLCVRES-1 and 2012/SPT/TOINDVAR-3 (72.56%) and 53.78%), 2012/SPT/TODVAR-10 (64.34% and 60.45%) and 2012/SPT/TOINDVAR-9 (54.56% and 45.44%) during both the years of observation. Out of the total materials screened, 14 were moderately resistant and 16 were susceptible under field condition after inoculation during both years.

The high resistance reaction in genotypes Sel-35 and Sel-19 could be due to the resistant gene derived from Konbilahi (Solanum pimpinellifolium) which is a wild species of tomato as reported by Kim et al. (2006). The available sources of resistance were mostly confined to the weedy relatives which have less in use and demand, like L. pimpinellifolium (Kalloo and Banerjee, 1993) and L. esculentum var. cerasiforme (Fageria, 1997). The response of the plant against A. solani was measured by observing the symptoms developed after inoculation. This result was similar to the findings of Poysa and Tu (1997) that attained resistance by incorporating resistance from other tomato species. Some experimental findings similar to the present study were also reported by Kamble et al. (2007) where they found five lines moderately resistant; Upadhay et al. (2009); Singh et al. (2011) revealed that accessions of wild relatives of tomato were highly resistant when screening was done under in vitro and Mahantesha et al. (2012) also reported resistant tomato genotypes against Alternaria solani under field conditions.

It was also evident from the study conducted that the reduction in yield due to early blight for highly resistant genotypes varies from 2.15% (Sel-19) to 3.05% (Sel-35). The percentage loss ranged from 5.79% to 9.86% for resistant, 10.61% to 14.50% for moderately resistant, 16.76% to 19.86% for susceptible and 30.15% to 42.75% for highly susceptible genotypes. 78% loss in fruit yield was also reported by Datar and Mayee, 1981; 95% fruit loss under severe epiphytotic condition as reported by Sridhara and Naik, 1983. The yield loss was worked out for the year 2012-13 and is given in table-3.

	Percent Disease Index (PDI)					Saara 27 Davis After		
Genotypes	7 Days After		22Days After		37 Days After		Score 37 Days After Inoculation	
	Inoculation		Inoculation		Inoculation			
	2012-13	2013-14	2012-13	2013-14	2012-13	2013-14	2012-13	2013-14
2012/TOLCVRES-1	39.25	42.34	51.08	54.45	73.56	72.25	5	5
2012/TOLCVRES-2	27.04	27.45	36.20	37.35	48.32	49.33	4	4
2012/TOLCVRES-3	11.98	10.24	12.66	14.35	14.34	20.34	2	2
2012/TOLCVRES-4	21.53	20.12	29.28	27.45	35.55	32.12	3	3
2012/TOLCVRES-5	41.23	39.09	55.33	53.69	79.43	77.53	5	5
2012/TOLCVRES-6	22.35	23.00	31.24	29.76	36.44	36.99	3	3
2012/TOLCVRES-7	23.02	21.99	33.22	35.24	42.54	41.21	4	4
2012/TOLCVRES-8	40.01	33.45	53.65	42.32	75.32	54.45	5	5
2012/TOLCVRES-9	20.16	19.31	25.03	24.31	30.43	31.21	3	3
2012/SPT/TOINDVAR-1	19.35	18.33	28.25	27.35	36.64	34.11	3	3
2012/SPT/TOINDVAR-2	20.25	18.73	29.14	27.35	35.35	36.47	4	4
2012/SPT/TOINDVAR-3	35.36	24.31	49.71	36.35	72.56	58.78	5	5
2012/SPT/TOINDVAR-4	12.47	13.45	17.25	16.34	24.34	20.24	2	2
2012/SPT/TOINDVAR-5	28.35	25.33	39.60	35.24	50.00	42.33	5	5

Table 1: Percent disease incidence of early blight in the tomato genotypes

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2012/SPT/TOINDVAR-6	27.16	21.35	38.71	33.25	49.54	40.33	4	4
2012/SPT/TOINDVAR-7	30.22	29.45	41.66	37.55	49.50	47.64	5	5
2012/SPT/TOINDVAR-8	13.10	12.20	15.24	17.34	23.44	24.10	3	3
2012/SPT/TOINDVAR-9	34.25	25.25	42.34	36.66	54.56	45.44	4	4
2012/SPT/TOINDVAR-10	18.90	21.01	29.00	27.45	36.46	35.44	3	3
2012/SPT/TODVAR-1	17.34	18.33	26.75	23.13	33.67	32.22	3	3
2012/SPT/TODVAR-2	22.44	20.73	32.14	31.35	37.50	37.00	4	4
2012/SPT/TODVAR-3	21.45	21.75	35.08	32.66	44.66	42.24	4	4
2012/SPT/TODVAR-4	23.46	22.44	30.14	32.33	38.64	40.44	4	4
2012/SPT/TODVAR-5	13.45	15.21	14.35	16.25	18.65	20.14	2	2
2012/SPT/TODVAR-6	17.35	15.45	19.25	19.85	23.34	24.00	2	2
2012/SPT/TODVAR-7	21.34	20.01	26.54	24.31	30.21	30.99	3	3
2012/SPT/TODVAR-8	19.08	19.09	27.33	24.13	32.35	30.12	3	3
2012/SPT/TODVAR-9	28.21	33.13	37.10	42.43	46.23	50.00	4	4
2012/SPT/TODVAR10	38.25	39.12	49.21	47.27	64.34	60.45	5	5
10/TOLCVRES-1	17.67	16.24	28.00	26.45	32.91	30.44	3	3
10/TOLCVRES-2	25.80	24.31	33.41	35.13	41.58	43.00	4	4
10/TOLCVRES-3	24.35	23.12	34.12	30.24	45.66	43.56	4	4
10/TOLCVRES-5	21.00	19.13	28.00	27.54	37.68	37.68	3	3
10/TOLCVRES-6	22.54	21.15	28.71	27.75	36.66	37.00	3	3
Sel-35	10.23	7.75	12.25	11.46	12.50	12.30	2	2
Sel-19	10.50	7.56	11.20	10.00	12.00	11.68	2	2
Sel-46	9.78	12.45	15.23	21.70	20.19	24.34	2	2
Sel-16	15.20	14.32	17.34	19.74	25.34	24.45	2	2
Sel-9	10.28	10.25	12.13	11.45	13.00	13.00	2	2
Arka vikas	29.73	28.42	30.08	32.95	38.12	40.99	4	4
Hisar Arun	21.44	14.05	25.77	22.02	33.00	35.24	3	3
H-86	28.29	17.72	34.48	22.45	43.24	38.54	4	4
Punjab Chhuhara(C)	25.34	24.34	29.12	28.75	37.00	37.45	3	3
H-24(C)	16.89	24.75	29.45	32.45	40.35	39.46	3	3
NDT-3 (C)	28.37	23.37	36.00	36.54	45.45	42.34	4	4
C.D. (5%)	3.26	5.37	2.79	3.20	4.40	3.84		
S.E. (m)	1.14	1.8	0.98	1.12	1.54	1.34		

Table 2: Reaction of tomato genotypes against early blight on the basis Percent Disease Index (PDI)

Score	Reaction	PDI value range (%)	Genotypes				
1	Highly resistant	0-12.5	Sel-35 and Sel-19				
2	Resistant 12.6–25.0		2012/SPT/TOINDVAR-4, 2012/SPT/TODVAR-5, 2012/SPT/TODVAR-6, 2012/TOLCVRES-3, Sel-46, Sel-16 and Sel-9				
3	Moderately resistant	Dederately resistant 25.1–37.5 2012/TOLCVRES-4, 2012/TOLCVRES-6, 2012/TOLCVRES-9, 2012/SPT/TOD 2012/SPT/TOINDVAR-8, 2012/SPT/TOINDVAR-10, 2012/SPT/TODVAR-8, 10/TO 2012/SPT/TODVAR-2, 2012/SPT/TODVAR-7, 2012/SPT/TODVAR-8, 10/TO 10/TOLCVRES-6, Punjab Chhuhara and Hisar Arun					
4	Susceptible	37.6–50.0	2012/TOLCVRES-2, 10/TOLCVRES-3, 2012/TOLCVRES-7, 2012/SPT/TOINDVAR-2, 2012/SPT/TOINDVAR-5, 2012/SPT/TOINDVAR-6, 2012/SPT/TOINDVAR-7, 2012/SPT/TODVAR-3, 2012/SPT/TODVAR-4, 2012/SPT/TODVAR-9, 10/TOLCVRES-2, 10/TOLCVRES-5, H-24, Arka Vikas, NDT-3 and H-86				
5	Highly susceptible	50.1 and above	2012/TOLCVRES-1, 2012/TOLCVRES-5, 2012/TOLCVRES-8, 2012/SPT/TOINDVAR-3, 2012/SPT/TOINDVAR-9 and 2012/SPT/TODVAR-10				

Table 3: Comparison of yield and its loss percentage between disease free condition and disease infested condition for the
year 2012-13

			year 2012-15			
Sl. No.	Genotypes	Resistant reaction from PDI reading	Yield per ha (q) (disease free condition)	Yield per ha (q) (disease infested condition)	Loss in yield per ha (q)(due to disease incidence)	% loss in yield (due to disease incidence)
1	2012/TOLCVRES-1	HS	246.53	172.21	74.32	30.15
2	2012/TOLCVRES-2	S	192.4	154.21	38.19	19.85
3	2012/TOLCVRES-3	S	240.00	220.21	45.59	17.15
4	2012/TOLCVRES-4	MR	237.9	203.43	34.47	14.49
5	2012/TOLCVRES-5	HS	221.83	151.34	70.49	31.78
6	2012/TOLCVRES-6	MR	222.47	190.21	32.26	14.50
7	2012/TOLCVRES-7	S	181.6	146.42	35.18	19.37
8	2012/TOLCVRES-8	HS	230.2	160.24	69.96	30.39
9	2012/TOLCVRES-9	MR	250.87	220.12	30.75	12.26
10	2012/SPT/TOINDVAR-1	MR	205.48	176.48	29.00	14.11

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11	2012/SPT/TOINDVAR-2	S	173.43	136.42	37.01	21.34
12	2012/SPT/TOINDVAR-3	HS	196.96	131.00	65.96	33.49
13	2012/SPT/TOINDVAR-4	R	126.1	118.80	7.30	5.79
14	2012/SPT/TOINDVAR-5	S	116.55	94.42	22.13	18.99
15	2012/SPT/TOINDVAR-6	S	159.55	132.21	27.34	17.14
16	2012/SPT/TOINDVAR-7	S	145.93	116.98	28.95	19.84
17	2012/SPT/TOINDVAR-8	MR	190.69	166.59	24.10	12.64
18	2012/SPT/TOINDVAR-9	HS	192.78	110.37	82.41	42.75
19	2012/SPT/TOINDVAR-10	MR	211.9	186.21	25.69	12.12
20	2012/SPT/TODVAR-1	MR	224.22	198.42	25.80	11.51
21	2012/SPT/TODVAR-2	MR	240.13	214.24	25.89	10.78
22	2012/SPT/TODVAR-3	S	264.67	212.11	52.56	19.86
23	2012/SPT/TODVAR-4	S	168.15	135.00	33.15	19.71
24	2012/SPT/TODVAR-5	R	188.52	170.21	18.31	9.71
25	2012/SPT/TODVAR-6	R	244.2	230.12	14.08	5.77
26	2012/SPT/TODVAR-7	MR	211.2	184.21	26.99	12.78
27	2012/SPT/TODVAR-8	MR	195.68	172.42	23.26	11.89
28	2012/SPT/TODVAR-9	S	235.62	189.00	46.62	19.79
29	2012/SPT/TODVAR10	HS	194.7	121.72	72.98	37.48
30	10/TOLCVRES-1	MR	201.29	178.34	22.95	11.40
31	10/TOLCVRES-2	S	208.67	168.21	40.46	19.39
32	10/TOLCVRES-3	R	268.82	250.12	18.70	6.96
33	10/TOLCVRES-5	S	196.78	158.21	38.57	19.60
34	10/TOLCVRES-6	MR	142.74	123.35	19.39	13.58
35	Sel-35	HR	80	77.56	2.44	3.05
36	Sel-19	HR	77.67	76.00	1.67	2.15
37	Sel-46	R	95.67	86.24	9.43	9.86
38	Sel-16	R	90.67	83.46	7.21	7.95
39	Sel-9	R	81.33	74.56	6.77	8.32
40	Arka vikas	S	194.48	156.22	38.26	19.67
41	Hisar Arun	MR	205.56	182.24	23.32	11.34
42	H-86	S	146.27	118.21	28.06	19.18
43	Punjab Chhuhara (C)	MR	221.76	198.24	23.52	10.61
44	H-24 (C)	S	303	252.21	50.79	16.76
45	NDT-3 (C)	S	258.19	212.12	46.07	17.84
45	NDT-3 (C)	S	258.19	212.12	46.07	17.84

It could be suggested from the result that the genotypes Sel-35 and Sel-19 were highly resistant and these genotypes could be utilized in any tomato breeding programmes as a donor parent for developing early blight resistant variety.

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