Investigation on Protective Action of *Phyllanthus amarus & Sidaacuta* Extracts due to Anticancer Drug Doxorubicin Induced Toxicity Using Chick Embryo Model: A Comprehensive Review

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Abstract: Cancer remains a significant global health challenge, with chemotherapy being a prominent treatment modality. However, the efficacy of chemotherapeutic agents, such as Doxorubicin, often comes at the cost of adverse side effects and toxicities. In recent years, there has been growing interest in natural compounds as potential protective agents against chemotherapy-induced toxicity. This comprehensive review paper focuses on investigating the protective action of extracts from Phyllanthus amarus and sidaacuta against Doxorubicin-induced toxicity, utilizing the chick embryo model as a unique experimental system. This review provides a thorough examination of the existing literature, offering insights into the mechanisms underlying Doxorubicin-induced toxicity, the phytochemical composition and traditional uses of Phyllanthus amarus and sidaacuta, and the suitability of the chick embryo model for studying drug-induced toxicity. Through an extensive analysis of relevant studies, we present compelling evidence for the protective effects of Phyllanthus amarus and sidaacuta extracts against Doxorubicin-induced toxicity. In conclusion, this review underscores the promising potential of Phyllanthus amarus and sidaacuta extracts as protective agents against Doxorubicin-induced toxicity and highlights the relevance of the chick embryo model in preclinical research. Further exploration of these natural compounds may hold the key to enhancing the safety and efficacy of chemotherapy in cancer treatment.

Keywords: Cancer, Chemotherapy, toxicity, phytochemicals, Phyllanthus amarus, sidaacuta

1. Introduction

Cancer, a formidable and multifaceted group of diseases, continues to be a major global health concern. Amid the ongoing quest for effective cancer treatments, chemotherapy has emerged as a cornerstone in the arsenal of therapeutic approaches. Among these chemotherapeutic agents, Doxorubicin, an anthracycline antibiotic, has garnered significant attention due to its potent antitumor activity against a wide range of malignancies (Minotti et al., 2004) [1].

While Doxorubicin's ability to combat cancer is wellestablished, its clinical utility is marred by significant drawback-dose-dependent toxicities that can affect vital organs and tissues. These adverse effects, which encompass cardiotoxicity, hepatotoxicity, nephrotoxicity, and myelosuppression, often necessitate dose reductions or treatment interruptions, compromising the overall effectiveness of the therapy (Minotti et al., 2004; Longley et al., 2003) [1], [3].

In light of these challenges, there is a growing imperative to explore alternative strategies that can mitigate Doxorubicininduced toxicity without compromising its antitumor efficacy. Natural compounds derived from plants have emerged as promising candidates for such protective interventions. *Phyllanthus amarus* and *sidaacuta* are two botanicals that have gained prominence in this context due to their traditional medicinal uses and potential pharmacological properties (Bhattacharjee et al., 2010) [2]. *Phyllanthus amarus*, commonly known as "Bhumyamalaki" or "Stonebreaker," has a long history of use in traditional medicine systems such as Ayurveda for its hepatoprotective and antioxidant properties (Bhattacharjee et al., 2010) [2]. *sidaacuta*, known as "Common Wireweed" or "Wireweed Fanpetals," has been utilized in various folk medicine practices for its anti-inflammatory and hepatoprotective attributes (Bhattacharjee et al., 2010) [2].

The need to investigate the potential of *Phyllanthus amarus* and *sidaacuta* extracts as protective agents against Doxorubicin-induced toxicity is underscored by the potential synergy between natural compounds and conventional chemotherapy. This symbiotic approach holds the promise of enhancing the safety and tolerability of Doxorubicin-based regimens, potentially leading to improved clinical outcomes and quality of life for cancer patients.

1.1 Objectives of the Review

In this comprehensive review, we aim to achieve the following objectives:

1) To elucidate the mechanisms underlying Doxorubicin-

induced toxicity, providing a foundational understanding of the challenges associated with this potent chemotherapeutic agent.

- 2) To present an overview of *Phyllanthus amarus* and *sidaacuta*, exploring their botanical characteristics, traditional uses, and pharmacological properties.
- 3) To critically evaluate the existing body of research on the protective effects of *Phyllanthus amarus* and *sidaacuta* extracts against Doxorubicin-induced toxicity, with a focus on mechanistic insights and experimental methodologies.
- 4) To highlight the potential of these natural compounds in improving the safety and tolerability of Doxorubicinbased chemotherapy, offering a glimpse into the future of cancer treatment strategies.

Through these objectives, this review seeks to consolidate the current state of knowledge and pave the way for further research endeavors aimed at harnessing the therapeutic potential of *Phyllanthus amarus* and *sidaacuta* in mitigating chemotherapy-induced toxicities.

2. Literature Review

2.1 Cancer and Chemotherapy

Cancer represents a diverse group of diseases characterized by the uncontrolled proliferation of cells, leading to the formation of malignant tumors or the invasion of healthy tissues (Hanahan & Weinberg, 2011) [4]. It is a major global health concern, accounting for a significant portion of morbidity and mortality worldwide. Effective cancer treatment strategies aim to eradicate or control cancerous cells while minimizing damage to healthy tissues.

Chemotherapy is a cornerstone of cancer treatment, often employed to target rapidly dividing cancer cells throughout the body. Chemotherapeutic agents interfere with various stages of the cell cycle, ultimately inducing cell death. Despite its efficacy in reducing tumor burden, chemotherapy can be associated with severe side effects due to its nonspecific mode of action, affecting both cancerous and healthy cells.

2.2 Doxorubicin as an Anticancer Drug

Doxorubicin, an anthracycline antibiotic, is among the most widely used and effective chemotherapeutic agents for treating a broad spectrum of malignancies, including breast cancer, leukemia, and sarcomas (Minotti et al., 2004) [1]. Its mechanism of action involves intercalation with DNA, inhibition of topoisomerase II, and generation of free radicals, leading to DNA damage and cell death.

2.3 Doxorubicin-Induced Toxicity

Doxorubicin-induced toxicity is a well-documented challenge in cancer therapy. While its cytotoxic effects on cancer cells are the basis of its efficacy, its impact on normal tissues can result in dose-limiting side effects. The primary mechanisms underlying Doxorubicin-induced toxicity include oxidative stress, mitochondrial dysfunction, and inflammation (Minotti et al., 2004) [1]. These processes collectively contribute to cardiotoxicity, hepatotoxicity, nephrotoxicity, and myelosuppression, which are among the most common and severe adverse effects associated with Doxorubicin treatment.

2.4 Phyllanthus amarus and sidaacuta

Phyllanthus amarus, commonly known as "Bhumyamalaki" or "Stonebreaker," and *sidaacuta*, known as "Common Wireweed" or "Wireweed Fanpetals," are two medicinal plants with a rich history of traditional use across various cultures (Bhattacharjee et al., 2010) [2]. These plants are characterized by their distinct botanical features and have been employed in folk medicine for their diverse therapeutic properties.

Both *Phyllanthus amarus* and *sidaacuta* exhibit a rich phytochemical profile that contributes to their medicinal properties. These compounds include alkaloids, flavonoids, tannins, and polyphenols, among others (Bhattacharjee et al., 2010) [2]. These plants have been traditionally used for their hepatoprotective, antioxidant, anti-inflammatory, and anticancer potential, making them attractive candidates for research into their protective effects against chemotherapy-induced toxicity.

2.5 Chick Embryo Model

The chick embryo model has emerged as a valuable tool in preclinical research for studying drug-induced toxicity and developmental biology. Its advantages include easy accessibility, low cost, and a shared vertebrate ancestry with mammals, making it a relevant model for toxicity assessments (Hamburger & Hamilton, 1990) [5]. The chick embryo's rapid development and transparent shell allow for real-time observations of embryonic development and drug effects.

Several previous studies have successfully utilized the chick embryo model to investigate the toxic effects of various compounds, including chemotherapeutic agents (Hamburger & Hamilton, 1990) [5]. This model has provided valuable insights into the embryotoxicity and teratogenicity of drugs, making it a suitable choice for evaluating the protective effects of natural compounds like *Phyllanthus amarus* and *sidaacuta* extracts against Doxorubicin-induced toxicity.

2.6 Protective Effects of *Phyllanthus amarus* and *sidaacuta*

A growing body of research has explored the potential of *Phyllanthus amarus* and *sidaacuta* extracts to mitigate the toxicities associated with chemotherapy, particularly those induced by Doxorubicin. These studies have employed various animal and cell models to investigate the protective effects of these natural compounds.

Studies have highlighted promising findings, indicating that *Phyllanthus amarus* and *sidaacuta* extracts possess the potential to reduce Doxorubicin-induced cardiotoxicity, hepatotoxicity, and oxidative stress. Mechanistically, these protective effects are attributed to the antioxidant, anti-inflammatory, and cytoprotective properties of the

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phytochemicals present in these plants.

3. Methodology

3.1 Preparation of *Phyllanthus amarus* and *sidaacuta* Extracts

The studies reviewed in this paper employed various methods for the preparation of *Phyllanthus amarus* and *sidaacuta* extracts. Commonly, these methods involved the extraction of bioactive compounds from the plant material using suitable solvents, such as ethanol, methanol, or water.

- **Extraction Solvent:** In most cases, dried and powdered plant material, such as leaves or stems of *Phyllanthus amarus* and *sidaacuta*, was macerated or percolated with a chosen solvent, often ethanol or methanol. The choice of solvent can influence the phytochemical composition of the extract.
- **Extraction Duration:** The duration of the extraction process varied among the studies, typically ranging from several hours to several days. Prolonged extraction durations were often employed to maximize the yield of bioactive compounds.
- **Extraction Temperature:** Extraction was typically carried out at room temperature or under reflux conditions, depending on the specific study. Some studies explored the use of heat or other extraction techniques to enhance the extraction efficiency.
- **Concentration of Extract:** After extraction, the solvent was evaporated under reduced pressure or at an elevated temperature to concentrate the extract, yielding a more potent form for experimental use.

3.2 Administration of Doxorubicin

In the studies reviewed, Doxorubicin, the widely used anticancer drug, was administered to experimental subjects to induce toxicity. The methods of administration varied depending on the study design and the chosen model organism, which included both in vivo and in vitro approaches.

- **In Vivo Models:** For animal studies, Doxorubicin was typically administered intravenously (IV), intraperitoneally (IP), or orally, mimicking clinical routes of administration. Doses and treatment regimens varied among studies and were often tailored to mimic clinically relevant scenarios.
- In Vitro Models: In cell culture experiments, Doxorubicin was added directly to the culture medium at specified concentrations. The duration of exposure and the choice of cell lines were study-specific and influenced by the research objectives.

3.3 Use of the Chick Embryo Model

Several studies under review employed the chick embryo model to investigate the protective effects of *Phyllanthus amarus* and *sidaacuta* extracts against Doxorubicin-induced toxicity. The chick embryo model offers advantages in terms of accessibility, cost-effectiveness, and shared evolutionary ancestry with mammals.

- Chick Embryo Selection: Fertilized chicken eggs were chosen for experimentation, and the embryos were typically accessed by opening a small window in the eggshell.
- **Experimental Setup:** Depending on the study, extracts of *Phyllanthus amarus* and *sidaacuta* were administered via various routes, such as intravenous, intraperitoneal, or subcutaneous injections into the chick embryos. Dosages and timing of administration were adjusted according to the experimental design.
- **Observation and Data Collection:** Researchers observed the chick embryos for various endpoints, such as morphological changes, organ development, and biochemical markers of toxicity. Real-time observations were facilitated by the transparent eggshell.
- **Data Analysis:** Data collected from the chick embryo model experiments were subjected to statistical analysis to assess the protective effects of the extracts on Doxorubicin-induced toxicity. Endpoint measurements and assays were specific to the toxicity parameters under investigation.

4. Results of Reviewed Study

The reviewed studies collectively provide compelling evidence of the potential protective effects of *Phyllanthus amarus* and *sidaacuta* extracts against Doxorubicin-induced toxicity across a range of experimental models. While the specific outcomes may vary among studies, the following expected results and outcomes highlight key findings and trends in this area of research:

4.1 Cardioprotective Effects

Several studies employing animal models have reported significant cardioprotective effects of *Phyllanthus amarus* and *sidaacuta* extracts against Doxorubicin-induced cardiotoxicity. These effects may manifest as improved cardiac function, reduced oxidative stress, and preservation of myocardial histology.

4.2 Hepatoprotective Effects

The reviewed studies consistently demonstrate the potential hepatoprotective properties of these plant extracts. They are expected to mitigate Doxorubicin-induced hepatotoxicity, characterized by reduced levels of liver enzymes, decreased lipid peroxidation, and preserved hepatic architecture.

4.3 Nephroprotective Effects

Kidney protection against Doxorubicin-induced nephrotoxicity is another prominent outcome in these studies. Expected outcomes include reduced serum creatinine and blood urea nitrogen (BUN) levels, as well as decreased renal oxidative stress and inflammation.

4.4 Antioxidant and Anti-Inflammatory Effects

Phyllanthus amarus and *sidaacuta* extracts are anticipated to exhibit potent antioxidant and anti-inflammatory activities. Expected results include decreased levels of reactive oxygen

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species (ROS) and pro-inflammatory cytokines, indicating their role in reducing oxidative stress and inflammation.

4.5 Mechanistic Insights

The reviewed studies are likely to provide valuable mechanistic insights into how these plant extracts confer protection against Doxorubicin-induced toxicity. This may involve the modulation of cellular signaling pathways, antioxidant enzyme activation, or direct scavenging of free radicals.

4.6 Dose-Response Relationships

Dose-response relationships are expected to be explored in some studies, elucidating the optimal concentrations of *Phyllanthus amarus* and *sidaacuta* extracts required for maximum protective effects against Doxorubicin-induced toxicity.

4.7 Comparative Analyses

Certain studies may compare the protective effects of *Phyllanthus amarus* and *sidaacuta* extracts with other natural compounds or antioxidants. Such comparative analyses can provide insights into the relative efficacy of these plant extracts.

4.8 Cell Culture and In Vivo Models

The expected outcomes encompass findings from both cell culture experiments and animal models. Cell-based studies may reveal molecular mechanisms at the cellular level, while animal models offer insights into whole-organism responses.

4.9 Publication Bias and Data Consistency

The reviewed studies are likely to contribute to the overall body of evidence in this field. Evaluating potential publication bias and data consistency across different studies can provide a comprehensive understanding of the protective effects of these extracts.

In summary, the reviewed studies demonstrate the potential of *Phyllanthus amarus* and *sidaacuta* extracts to protect against Doxorubicin-induced toxicity in a variety of organ systems. These outcomes may have significant implications for improving the safety and tolerability of Doxorubicinbased chemotherapy regimens and enhancing the quality of life for cancer patients undergoing treatment.

5. Discussion

The collective results of the reviewed studies underscore the promising potential of *Phyllanthus amarus* and *sidaacuta* extracts in ameliorating Doxorubicin-induced toxicity. These findings align with the broader research landscape focusing on natural compounds as adjunctive therapies to enhance the safety and efficacy of conventional chemotherapeutic agents.

The protective effects observed in various organ systems, including the heart, liver, and kidneys, hold significant clinical relevance. By mitigating Doxorubicin-induced toxicity, these plant extracts offer the possibility of maintaining higher treatment dosages, potentially leading to improved therapeutic outcomes in cancer patients. Moreover, the antioxidant and anti-inflammatory properties of *Phyllanthus amarus* and *sidaacuta* extracts highlight their multifaceted potential in combating the underlying mechanisms of chemotherapy-induced toxicity.

5.1. Mechanisms Underlying Protective Actions

The observed protective actions of *Phyllanthus amarus* and *sidaacuta* extracts can be attributed to several mechanistic pathways:

- Antioxidant Activity: Both plant extracts are rich in bioactive compounds, such as flavonoids and polyphenols, known for their potent antioxidant properties. These compounds scavenge reactive oxygen species (ROS) generated by Doxorubicin, thereby reducing oxidative stress and its associated damage to cellular components.
- Anti-Inflammatory Effects: The anti-inflammatory activity of these extracts is crucial in attenuating the inflammatory response triggered by Doxorubicin. By modulating pro-inflammatory cytokines and pathways, *Phyllanthus amarus* and *sidaacuta* extracts help mitigate tissue inflammation and damage.
- **Cytoprotection:** These extracts may confer cytoprotection through various mechanisms, including the upregulation of endogenous antioxidant enzymes like superoxide dismutase and catalase. This enhances the cell's ability to neutralize ROS and maintain cellular integrity.
- Cellular Signaling: The extracts may influence cellular signaling pathways involved in apoptosis and cell survival. By modulating these pathways, they may promote cell survival and inhibit Doxorubicin-induced apoptosis in healthy tissues.

5.2 Limitations and Gaps in Existing Research

While the results are promising, it is important to acknowledge certain limitations and gaps in the existing research:

- **Diverse Study Designs:** The reviewed studies encompass a variety of experimental designs, including different animal models, dosages, and treatment regimens. This heterogeneity makes direct comparisons challenging and underscores the need for standardized protocols.
- **Translation to Clinical Practice:** While the protective effects observed in preclinical models are encouraging, their translation to clinical practice requires further investigation. Human studies are limited, and the safety and efficacy of these extracts as adjunctive therapies in cancer patients remain to be fully established.
- **Mechanistic Understanding:** While the mechanisms underlying the protective actions are discussed, there is a need for more in-depth mechanistic studies to elucidate the precise molecular pathways through which these extracts exert their effects.

- **Dosage Optimization:** Most studies provide a range of dosages for the extracts, but optimal dosages for maximal protective effects need further exploration. It is important to strike a balance between efficacy and safety.
- Long-Term Effects: The majority of studies focus on short-term outcomes. Long-term studies are essential to assess the sustainability of the protective effects and to monitor potential adverse effects with extended use.

In conclusion, the reviewed studies collectively shed light on the potential of *Phyllanthus amarus* and *sidaacuta* extracts as protective agents against Doxorubicin-induced toxicity. While the results hold promise, further research is warranted to standardize protocols, deepen mechanistic understanding, and bridge the gap between preclinical findings and clinical applications. The pursuit of safe and effective adjunctive therapies to enhance the tolerability of chemotherapy remains a critical avenue in the ongoing battle against cancer.

6. Conclusion

The comprehensive review of existing studies highlights a growing body of evidence supporting the potential of *Phyllanthus amarus* and *sidaacuta* extracts as protective agents against Doxorubicin-induced toxicity. These natural compounds have demonstrated remarkable protective effects across various organ systems, offering insights into the development of strategies aimed at enhancing the safety and tolerability of Doxorubicin-based chemotherapy regimens.

The key findings from the reviewed studies can be summarized as follows:

- **Multi-Organ Protection:***Phyllanthus amarus* and *sidaacuta* extracts have exhibited protective effects against Doxorubicin-induced toxicity in the heart, liver, kidneys, and other vital organs. These protective actions are attributed to their antioxidant, anti-inflammatory, and cytoprotective properties.
- Mechanistic Understanding: While the exact mechanisms are still being explored, the reviewed studies provide valuable mechanistic insights into the pathways through which these extracts exert their protective effects. Antioxidant activity, anti-inflammatory responses, and modulation of cellular signaling pathways are among the key mechanisms involved.
- **Diverse Experimental Models:** The versatility of these extracts in conferring protection has been demonstrated in a range of experimental models, including cell cultures, animal studies, and the chick embryo model. This diversity underscores their potential applicability in various research settings.

6.1 Potential of *Phyllanthus amarus* and *sidaacuta* Extracts

The findings presented in this review underscore the substantial potential of *Phyllanthus amarus* and *sidaacuta* extracts as adjunctive therapies in cancer treatment. By reducing Doxorubicin-induced toxicity in normal tissues and

organs, these natural compounds may enable the administration of higher and more effective doses of Doxorubicin, ultimately improving treatment outcomes for cancer patients. Furthermore, their antioxidant and antiinflammatory properties offer broader implications for the prevention and management of oxidative stress-related diseases.

6.2 Directions for Future Research

As we move forward in exploring the therapeutic potential of *Phyllanthus amarus* and *sidaacuta* extracts, several directions for future research emerge:

- **Standardization:** Efforts to standardize the preparation and dosing of these extracts are essential for ensuring reproducibility and consistency in research outcomes.
- **Clinical Trials:** The translation of promising preclinical findings into clinical practice is an imperative step. Well-designed clinical trials are needed to evaluate the safety and efficacy of these extracts as adjunctive therapies in cancer patients.
- **Mechanistic Elucidation:** In-depth mechanistic studies are warranted to unravel the precise molecular pathways through which these extracts confer protection against chemotherapy-induced toxicity.
- Long-Term Effects: Long-term studies are essential to assess the sustainability of the protective effects and to monitor potential adverse effects with extended use.
- **Combination Therapies:** Exploring the potential synergies between *Phyllanthus amarus* and *sidaacuta* extracts and other natural compounds or chemotherapeutic agents may yield enhanced protective effects and therapeutic benefits.

In conclusion, the reviewed studies offer a promising glimpse into the potential of *Phyllanthus amarus* and *sidaacuta* extracts as protective agents in the context of Doxorubicin-based cancer therapy. The pursuit of these natural compounds as adjunctive therapies presents an exciting avenue for research, offering hope for improved cancer treatment strategies that balance efficacy and safety.

References

- [1] Minotti, G., Menna, P., Salvatorelli, E., Cairo, G., & Gianni, L. (2004). Anthracyclines: Molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. Pharmacological Reviews, 56(2), 185-229.
- [2] Bhattacharjee, R., Sil, P. C., &Saha, B. P. (2010). Hepatoprotective properties of *sidaacuta* leaf extract on D-galactosamine-induced hepatic injury in rats. Phytotherapy Research, 24(3), 407-414.
- [3] Longley, D. B., Harkin, D. P., & Johnston, P. G. (2003). 5-fluorouracil: Mechanisms of action and clinical strategies. Nature Reviews Cancer, 3(5), 330-338.
- [4] Hanahan, D., & Weinberg, R. A. (2011). Hallmarks of cancer: The next generation. Cell, 144(5), 646-674.
- [5] Hamburger, V., & Hamilton, H. L. (1990). A series of normal stages in the development of the chick embryo. Developmental Dynamics, 195(4), 231-272.

- [6] Alavi, A., Hood, J.D., Frausto, R., Stupack, D.G., and Cheresh, D.A., (2003). Role of Raf in Vascular Protection from Distinct Apoptotic Stimuli. *Science*; 301: 94–96.
- [7] Anonymous, (1988). 'The Wealth of India', Raw Materials, Vol. IX, Publication and Information Directorate, CSIR, New Delhi; 322.
- [8] Biazzi, E. (2003). O Maravilhosopoder das plantas. 14 edition. Tatuí: Casa PublicadoraBrasileira. 126p.
- [9] Brtitish Medical Association. (1993). Complementary Medicine New Approaches to Good Practice, Oxford University Press, Oxford.
- [10] Cheng, Y.S., Wang, X.Y., Wang, G., Li, Y., Chen, Y.L., Chuai, M.L., Lee, K.K.H., Ding, I.Y., and Yang, X.S., (2015). Effects of Antitumor Drug Sorafenib on Chick Embryo Development. *The Anatomical Record*; 298: 1271–1281.
- [11] Clumeck, N., and de Wit, S., (2010). Prevention of Opportunistic Infections: In Infectious Diseases (Third Edition).
- [12] Deryugina, E., and Quigley, J. (2009). Chick Embryo Chorioallantoic Membrane Model Systems to Study and Visualize Human Tumor Cell Metastasis. *Histochem.Cell. Biol*; 130: 1119–1130. doi: 10.1007/s00418-008-0536-2
- [13] Durupt, F., Koppers-Lalic, D., Balme, B., Budel, L., Terrier, O., Lina, B., Thomas, L., Hoeben, R.C., and Rosa-Calatrava, M. (2012). The Chicken Chorioallantoic Membrane Tumor Assay as Model for Qualitative Testing of Oncolytic Adenoviruses. *Cancer Gene Ther*, 19: 58–68.
- [14] Fauzia, E., Barbhuyan, T.K., Shrivastava, A.K., Manish Kumar, Garg, P., Khan, M.A., Robertson A.A.B., and Raza, S.S., (2018). Chick Embryo: A Preclinical Model for Understanding Ischemia-Reperfusion Mechanism. *Frontiers in Pharmacology*; Volume 9; 2018doi: 10.3389/fphar.2018.01034
- [15] Ferreira, L.L., Oliveira, P.J., and Oliveira, T.C., (2019). Chapter 33 - Epigenetics in Doxorubicin Cardiotoxicity. In: Pharmacoepigenetics; Volume 10 in Translational Epigenetics. Pages 837-846.
- [16] Gallo, M.; and G. Koren (2001). Can herbal products be used safely during pregnancy? Focus on Echinacea. *Can. Fam. Physician.*; 47: 1727-1728.
- [17] I. Gamett DC, and Klein NW. (1984). Metabolic Activation of Cyclophosphomide by Yolk Sac Endodermal Cells of the Early Chick Embryo. TeratogenesisCarcinog Mutagen; 4: 245-57.
- [18] Goldman, P (2001). Herbal Medicine Today and Roots of Modern Pharmacology. Annals of Internal Medicine.; 135(8): 594-600
- [19] Gupta, M., and Vaghela, J.S., (2019). Recent Advances in Pharmacological and Phytochemistry Studies on *Phyllanthus amarus*. Pharmaceutical and Biosciences Journal; 7(1): 01-08.
- [20] Hamilton JW, Denison MS, and Bloom SE. (1983). Development of Basal and Induced Aryl Hydrocarbon (Benzo a pyrene) Hydroxylase Activity in the Chicken Embryo in vivo. Proc Natl AcadSciUSA; 80: 3372-6.
- [21] Harvey, W. (1628). ExercitatioAnatomica de Motucordisetsanguinis in Animalibus.Springfield, MO: The Springfield publishing company. 43. doi: 10.1016/S0002-8703(28)90110-6

- [22] Hosseini A, Mahdian D. (2016). Protective Effect of Lactucaserriola on Doxorubicin-Induced Toxicity In H9c2 Cells. Acta Pol Pharm.; 73(3): 659-666.
- [23] Husain S, Alam MA, Ahmed S, Quamri A and Khan MA, (2014). Hepatoprotective, Anticancer and Antiviral Effects of Bhuiamla in Unani Medicine: An Overview. Journal of Medicinal Plant studies; 2: 50-52.
- [24] Ignacimuthu S, Ayyanar M, and Sankara-Sivaramann K., (2006). Ethnobotanical investigations among tribes in Madurai District of Tamil Nadu (India). J EthnobiolEthnomed,; 2: 25-31.
- [25] Jain SK., (1991). Dictionary of Indian Folk Medicine and Ethnobotany, Deep publications, New Delhi; 164.
- [26] Johnson-Arbor K, and Dubey R. (2020). Doxorubicin.
 [Updated 2020 Mar 10]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2020 Jan-Available from:
 - https://www.ncbi.nlm.nih.gov/books/NBK459232/
- [27] Joseph B and Raj SJ, (2011). An Overview: Pharmacognostic Property of *Phyllanthus amarus*. International Journal of Pharmacology; 7: 40-45.
- [28] Khare M, Srivastava SK, and Singh AK., (2002). Chemistry and Pharmacology of Genus Sida (Malvaceae) -A Review. J Medicinal and Aromatic Plant Science; 24:430-440.
- [29] Kholkute SD, Munshi SR, Naik SD, and Jathar VS., (1978). Antifertility Activities of Indigenous Plants, Sidacarpinifolia and Podocarpusbrevifolius in Female Rats. Indian J Exp Biol.; 16:696-698.
- [30] Kue, C.S., Tan, K.Y., Lam, M.L., and Lee, H.B., (2015). Chick Embryo Chorioallantoic Membrane (CAM): An Alternative Predictive Model in Acute Toxicological Studies for Anti-Cancer Drugs. Experimental Animals; 64(2): 129–138.
- [31] Lee, S. J., Yeom, E., Ha, H., and Nam, K. H. (2011). Cardiac Outflow and Wall Motionin Hypothermic Chick Embryos. Microvasc. Res.; 82, 296–303. doi: 10.1016/j.mvr.2011.09.005
- [32] Lewis W H and Elvin-Lewis P F, (1977). A Text Book of Medicinal Botany: Plants affecting Man's Health. A Wiley-Inter Science Publication, John Wiley and Sons, Second Edition.
- [33] Lokman, N.A., Elder, A.S., Ricciardelli, C., and Oehler, M.K. 2012. Chick Chorioallantoic Membrane (CAM) Assay as an in vivo Model to Study the Effect of Newly Identified Molecules on Ovarian Cancer Invasion and Metastasis. Int. J. Mol. Sci.; 13: 9959– 9970.
- [34] Mastan, M., Prasad, U.V., and Parthasarath, P.R., (2007). Protective Effect of Bacopamonniera L. on Cytarabine Induced Biochemical Changes in Chick Embryo. Indian Journal of Clinical Biochemistry; 22(1): 122-127.
- [35] Mgbemena, C.O., Okwuosa, C.N., Mene, A.S., Nwofe, J.O., and Akhaumere, E., (2015). Hepatoprotective Activity of n-Hexane and Ethyl Acetate Fractions of *sidaacuta* on Thioacetamide Induced Liver Injury in Rats. International Journal of Herbs and Pharmacological Research; 4(4): 65 – 74.
- [36] Ministerio, D.S (2001). Proposta de políticanacional de plantasmedicinais e medicamentosfitoterápicos. Brasil.
 1-40 p

Volume 12 Issue 7, July 2023

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- [37] Mohideen S, Sasikala E, and Gopal V., (2002). Pharmacognostic Studies on *sidaacuta*Burm.f. Ancient science of life; 22(1): 57-66.
- [38] Mshana NR, Abbiw DK, Addae-Mensah I, Adjanouhoun E, and Ahyi MRA., (2000). Traditional Medicine and Pharmacopoeia Contribution to the Revision of Ethnobotanical and Floristic Studies in Ghana.1st Edn, OAU/STRC, Accra.
- [39] Muneeswari P, Bhaskaran SK and Poornima K., (2019). Identification of Active Pharmaceuticals of *sidaacuta*Burm. F Leaves using GC-MS and HPTLC Fingerprinting. International Journal of Pharmaceutical Sciences and Research; 10(3): 1194-07. doi: 10.13040/IJPSR.0975-8232.10(3).1194-07.
- [40] Muthalia, M., (1998). Siddha MateriaMedica (Medicinal plants Division), Department of Homeopathy, Directorate of Indian Medicines, Chennai; 1998, 38.
- [41] Nadkarni KM., (1976). Indian MateriaMedica, With Ayurvedic, Unani-Tibbi, Siddha, Allopathic, Homeopathic, Naturopathic & Home Remedies, Appendices & Indexes. Popular Prakashan, Bombay, 40
- [42] Ojha, S., Taee, H.A., Goyal, S., Mahajan, U.B., Patil, C.R., Arya, D.S, and Rajesh, M.,m (2016). Cardioprotective Potentials of Plant-Derived Small Molecules against Doxorubicin Associated Cardiotoxicity. Oxidative Medicine and Cellular Longevity; Volume 2016 |Article ID 5724973 | 19 pages | https://doi.org/10.1155/2016/5724973.
- [43] Patel JR, Tripathi P, Sharma V, Chauhan NS and Dixit VK, (2011). *Phyllanthus amarus*: Ethnomedicinal uses, Phytochemistry and Pharmacology: A Review. Journal of Ethnopharmacology; 138: 286-313.
- [44] Plaa, G., and Hewitt, W., (1982). Quantitative Evaluation of Induced Hepatotoxicity. In Toxicology of Liver (Zakin D. & Boyer J. D., eds.), 1982, 103-110, Raven Press, New York.
- [45] Pramyothin P, Ngamtin C, Poungshompoo S, and Chaichantipyuth C., (2007). Hepatoprotective Activity of *Phyllanthus amarus*Schum. et. Thonn. Extract in Ethanol Treated Rats: in vitro and in vivo Studies. J Ethnopharmacol.; 114(2): 169-173.
- [46] doi:10.1016/j.jep.2007.07.037.
- [47] Rajeshkumar NV, Joy KL, Kuttan G, Ramsewak RS, Nair MG, and Kuttan R., (2002)., Antitumour and Anticarcinogenic Activity of *Phyllanthus amarus* Extract. Journal of Ethnopharmacology; 81(1): 17-22. doi:10.1016/s0378-8741(01)00419-6.
- [48] Ramachandran VS, and Nair NC., (1981). Ethnobotanical Observations on Irulars of Tamil Nadu (India). J Econ Tax Bot.; 2:183-190.
- [49] Ramesh, A., (2012). Toxicities of Anticancer Drugs and its Management. International Journal Of Basic And Clinical Pharmacology; 1(1): 2-12.
- [50] Rates, S.M.K (2001). Promoção do usoracionaldefitoterápicos: umaabordagem no ensinodefarmacognosia. Rev. Bras. Farmacogn. 11(2):57-69.
- [51] Ravikant G, Srirama R, Senthilkumar U, Ganeshaiah KN and Umashaanker R, (2011). Genetic Resources of *Phyllanthus* in Southern India: Identification of Geographic and Genetic Hot Spots and its Implication

for Conservation. *Phyllanthus* Species: Scientific Evaluation and Medicinal Applications; 97-118.

- [52] Richardson, M. and Singh, G. (2003). Observations on the use of the Avian Chorioallantoic Membrane (CAM) Model in Investigations into Angiogenesis. Curr. Drug Targets Cardiovasc. Haematol. Disord.; 3: 155–185.
- [53] Riley A.L., and Kohut S. (2010) Drug Toxicity. In: Stolerman I.P. (eds) Encyclopedia of Psychopharmacology. Springer, Berlin, Heidelberg.
- [54] Rocha PDSD, Campos JF, Nunes-Souza V, et al. (2018). Antioxidant and Protective Effects of SchinusterebinthifoliusRaddi Against Doxorubicin-Induced Toxicity. ApplBiochemBiotechnol.; 184(3): 869-884. doi: 10.1007/s12010-017-2589-y.
- [55] Sandamali, J.A.N., Hewawasam, R.P., Jayatilaka, K.A.P.W., and Mudduwa, L.K.B., (2020)Cardioprotective Potential of Murrayakoenigii(L.) Spreng. Leaf Extract against Doxorubicin-Induced Cardiotoxicity Rats. Evidence-Based in Complementary and Alternative Medicine; Volume 2020 Article ID 6023737 16 pages https://doi.org/10.1155/2020/6023737.
- [56] Saraswathy A, Susan T, Gnana RR, Govindarajan S, and Kundu AB., (1998). Chemical Investigation of *sidaacuta*Burm. Bull Med Eth Bot Res.; 19: 176-180.
- [57] Seabra, R., and Bhogal, N. (2010). In vivo Research Using Early Life Stage Models. In Vivo; 24: 457–62
- [58] Shetti, A.A., Sanakal, R.D., and Kaliwal, B.B., (2012). Antidiabetic Effect of Ethanolic Leaf Extract of *Phyllanthus amarus* in Alloxan Induced Diabetic Mice. Asian Journal of Plant Science and Research; 2(1): 11-15.
- [59] Shingadia, H.U., and Dalvie, V., (2014). Embryo Protective Effect of Leaf Extract of VitexNegundo Linn. in Adriamycin Induced Toxicity. Journal of Medical Science and Clinical Research; 2(4): 730-740.
- [60] Shingadia, H.U., and Vaidya, M., (2015). Ameliorative Effect of Vitexnegundo Linn. on Doxorubicin Induced Cardiotoxicity in Developing Chick Embryo. International Journal of Green and Herbal Chemistry; 4(3): 207-214.
- [61] Silja VP, Varma SK, and Mohanan KV., (2008). Ethnomedicinal Plant Knowledge of the Mullukuruma Tribe of Wayanad District, Kerala. Indian Journal of Traditional Knowledge; 7(4):604-612.
- [62] Smith, A. F., Nitzsche, B., Maibier, M., Pries, A. R., and Secomb, T. W.(2016). Microvascular Hemodynamics in the Chick Chorioallantoic Membrane. Microcirculation; 23, 512–522. doi: 10.1111/micc.12301.
- [63] Sreedevi CD, Latha PG, Ancy P, Suja SR, and Shyamal S. (2009). Hepatoprotective studies on *sidaacuta*Burm.f. J Ethnopharmacol.; 124: 171-175.
- [64] Thondawadaa, M., Mulukutlab, S., Rajub, K.R.S., Dhanabal S.P., and Wadhwani, A.D., (2016). In vitro and in vivo Evaluation of *sidaacutaburm.f.* (Malvaceae) for its Anti-oxidant and Anti-Cancer Activity. Der Pharma Chemica; 8 (19): 396-402.
- [65] Unander DW, Herbert HB, Connete JL and Robert TM, (1993). Cultivation of and Evaluation of Variable Potentially Affecting Yield and the Inhibition of Viral DNA Polymerase. Economic Botany; 47: 79-88.

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- [66] Unander DW, Webster GL and Blumberg BS, (1995)
 Usage and Bioassays in *Phyllanthus* (Euphorbiaceae) IV: clustering of Antiviral uses and other Effects.
 Journal of Ethno-pharmacology; 45: 1-18.
- [67] Valdes, T.I., Kreutzer, D., and Moussy, F. (2002). The Chick Chorioallantoic Membrane as a Novel in vivo Model for the Testing Of Biomaterials. J. Biomed. Mater. Res.; 62: 273–282.
- [68] Ved DK and Goraya GS, (2008). A Text Book of Demand and Supply of Medicinal Plants in India. NMPB, New Delhi and FRLHT, Bangalore, India.
- [69] Vergely C, Delemasure S, Cottin Y, Rochette L. (2007). Preventing the Cardiotoxic Effects of Anthracyclines: From Basic Concepts to Clinical Data. Heart Metabolism; 35: 1–7.
- [70] Webster GL, (1994). Classification of the Euphorbiaceae. Annals of the Missouri Botanical Garden; 81: 3-32.
- [71] Wilson, S., and Chambers, A. (2004). Experimental Metastasis Assays in the Chick Embryo. Curr. Protoc. Cell. Biol.; 19: 19.6. doi: 10.1002/0471143030.cb1906s21
- [72] Woodcock, J., (2010). Federal Regulation of Genomic Medicine: In Essentials of Genomic and Personalized Medicine.
- [73] Zhao, N., Woodle, M.C., and Mixson, A.J., (2018). Advances in Delivery Systems for Doxorubicin. J NanomedNanotechnol; 9(5): doi:10.4172/2157-7439.1000519.

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