

Impact of ZnO NPs, Pi Availability and the Interaction of Pi x NaCl in Reducing the Negative Effect of Salinity and Pi Deficiency on Growth of Arabidopsis Plant

Azizah M. Nahari¹, Al-Zahrani, H. S.²

Department of Biological Sciences, Faculty of Science, King Abdulaziz University, Jeddah, Saudi Arabia

Email: amnahary@uj.edu.sa

Abstract: Most crop production is decreased by salinity in agriculture, which is one of the main abiotic stressors that lowers crop production. One of the detrimental effects of salinity on plant growth and productivity is known as nutritional imbalance such as phosphate (Pi). Approximately 54% of the cultivated area of Saudi Arabia suffers from salinization. While its soils characterized by low available Pi. Such regions in Saudi Arabia, the factors of soil salinity and Pi deficiency pose major challenge to agricultural production. Nowadays, zinc oxide nanoparticles (ZnO NPs) are widely used in agriculture, and many investigations were carried out to determine the impact of ZnO NPs on plants under abiotic stress. So far, few research investigated into the potential impact of ZnO NPs on the growth of Arabidopsis plants under salt and Pi deficiency, and their interaction. This study aimed to investigate the effect of ZnO NPs application (10 mg/L), Pi availability and the interaction of Pi x NaCl in reducing the negative effect of the salinity and Pi deficiency on growth of Arabidopsis. Arabidopsis plants were grown hydroponically under different treatments of Pi and NaCl, and the interaction of Pi x NaCl with/without ZnO NPs and ZnO BPs. The results indicated that salinized plants had a reduction in plant growth traits, but, the values of these traits increased with high Pi (100 and 250 μ M). Pi at 10 μ M is more detrimental than NaCl at 100 mM for plant growth. Salt-treated plants (50, 100 mM) exposed to high Pi (100 and 250 μ M) showed higher salt tolerance. In contrast, plants subjected to NaCl (150 mM) and Pi (250 μ M) exhibited reduce in the values of growth parameter. Results also showed that ZnO NPs and ZnO BPs was sufficient to protect plant against salinity and Pi deficiency, however ZnO NPs was more efficient than ZnO BPs. The Pi availability, the combined treatment of Pi x NaCl, and ZnO NPs can be a promising application to alleviate the negative impacts of salinity and Pi deficiency on plant growth.

Keywords: Arabidopsis thaliana, ZnO-NPs application, salinity stress, deficiency and availability of Pi, Interaction of Pi x NaCl

1. Introduction

The sustainability of agriculture is seriously threatened by both the loss of arable land and the growth of the human population (Shokat and Grosskinsky, 2019). One of the primary abiotic stresses restricting crop production is high salt content in soils, which is a significant environmental issue for agriculture globally (Srivastava, 2020). By the entry of both Na and Cl into the cells result in severe ion imbalance and excessive uptake may result in significant physiological disorder. Where High levels of Na prevent K uptake which is a necessary component for growth and development of plant (James *et al.*, 2011).

It is known that only 10-30% of the applied Pi fertilizer can be recovered by the crop grown (Syers *et al.*, 2008), while above 80% of the Pi remains immobile and is not taken up by plants (Kapri and Tewari 2010). The world reliance on fertiliser of Pi for production of food results in decreasing resources for rock Pi and increasing in expenses of the production of Pi fertiliser in the next decade. All indicate that a crisis is about to occur which will have disastrous effects for the world's food security (Cordell *et al.*, 2009). Soil nutrients must be managed carefully, and feasible strategies must be in place for enhancing agricultural productivity in low Pi soils. One of these strategies is to utilise Pi fertiliser more wisely, efficiently, and in an

adequate quantity without causing any environmental effect (Mahapatra *et al.*, 2022).

The salinity factor, which is rising in many agricultural soils, particularly dry and semiarid, is well recognized to increase the loss of phosphorus (P) availability (Tchakounté *et al.*, 2020). Thus, in these types of soils, plants experience both salinity and Pi deficiency (Dey *et al.*, 2021). Thus, nutrient supplementation in the form of chemical fertilizer is needed for optimum plant growth and high yields since nutrients are crucial for development and growth of plant (Usman *et al.*, 2020)⁵. However, the excessive use of chemical fertilizers in agricultural activities causes several environmental problems. Nowadays, the important issue is to find proper and sustainable technique to help plants adapt more rapidly to environmental factors without harming ecosystems (Vermeulen *et al.*, 2012). Using nanoparticles (NPs) application for agricultural purposes is a new and promising approach to enhance agricultural practices. By optimising the administration of nutrients, the use of NPs aims to minimise nutrient losses during fertilisation, lower the quantity of plant protection chemicals use, and boost yields (Das *et al.*, 2015). One of the most widely applied nanoparticles in agriculture is zinc oxide nanoparticles (ZnO NPs) (Lang *et al.*, 2021). ZnO NPs as nano-fertilizer has a favourable effect on plants and it can be effectively applied in

agriculture without damaging the ecology or releasing nutrients into the environment (Thounaojam *et al.*, 2021). We evaluated the effect of ZnO NPs application, appropriate amount of Pi and the interaction of Pi and NaCl in reducing the negative effect of the salinity stress and Pi deficiency on *Arabidopsis* growth.

2. Material and Methods

2.1 Model plant

In the experiments of this project, we used *Arabidopsis thaliana* Columbia-0 plant. The seeds were obtained from King Abdullah University of Science and Technology (KAUST), in Saudi Arabia. The growth was carried out in cabinet room by using an ideal hydroponic growth system we designed to adapt to *Arabidopsis* growth habit.

2.2 General growth conditions for *Arabidopsis*

Arabidopsis thaliana plant was used in all experiments involving: (1) 4 different levels of NaCl (0.50, 100 and 150 mM), (2) 4 different levels of Pi (0, 10, 100 and 250 μ M) and (3) and their interaction Pi x NaCl (4x NaCl and 4xPi, 16

processing) with/without applications of ZnO NPs and ZnO BPs (10 mg/l). All experiments were performed in the hydroponic system containing Gibeaut's solution which consisted of macronutrients and micronutrients (Table 1.1), all solutions were autoclaved (Gibeaut *et al.*, 1997). The controlled hydroponic experiments were carried out in growth rooms at 24 °C with a 16/8 hr day/night cycle.

2.3 Hydroponic growth

2.3.1 Preparation of hydroponic growth

The purpose of this hydroponic system was to minimize physical limitations in the hydroponic system and Pi effectively became a mobile nutrient. Also, this system is faster and more accurate for analyzing a range of growth trait *Arabidopsis thaliana*. Seed holders were prepared by cutting off the bottom and gluing on a piece of grey mesh (1 cm) to avoid the loss of agar. The lids were made of a rectangular plastic sheet. 56 holes (1.3 cm diameter) were drilled in these lids to carry the seed-holders during growth. Black plastic containers) filled up with Gibeaut's solution. In such designs for hydroponic growth, the growth of algae is prevented as well as the media solution remains clear, because the media containers are made of black plastic (Figure 1).

Table 1: Chemical composition of Gibeaut's solution was used in the experiment of hydroponic growth system. Gibeaut's solution consisted of Gibeaut's Macronutrients, and Gibeaut's Micronutrients

Gibeaut's Macronutrients						
Molecule	MW	Stock [M]	[g]/ [L]	[g]/250 [ml]	Final conc. [mM]	Volume of stock in media [ml/10l]
Ca(NO ₃) ₂ x 4H ₂ O	236.15	1	236.15	59.0375	1.5	15
KNO ₃	101.11	1	101.11	25.2775	1.25	12.5
MgSO ₄	120.37	1	120.37	30.0925	0.75	7.5
Na ₂ SiO ₄ · 9H ₂ O	284.2	0.1	28.42	7.105	0.1	10
Fe-EDDHA	435.2	0.036	15.6672	3.917	0.036	10
Gibeaut's Micronutrients						
Molecule	MW	Stock [mM]	[g]/ [L]	[g]/250 [ml]	Final conc. [μM]	Volume of stock in media [ml/10l]
KCl	74.56	50	3.728	0.932	50	10
MnSO ₄ · H ₂ O	169.01	10	1.6901	0.4225	10	
CuSO ₄ · 5H ₂ O	249.68	1.5	0.37452	0.0936	1.5	
ZnSO ₄ · 7H ₂ O	287.54	2	0.57508	0.1437	2	
H ₃ BO ₃	61.83	50	3.0915	0.7728	50	
(NH ₄) ₆ Mo ₇ O ₂₄	1235.9	0.075	0.092693	0.0231	0.075	

2.3.2 Preparation of germination

Before germination, seeds of *Arabidopsis thaliana* were imbibed in 0.1% agar at 4 °C for 5 days before transferring to growth room where the hydroponic system will be placed. Around 1ml of melted 0.4% agar (germination medium) was poured into each seed-holders and left to solidify for 20 mins. Once it cooled down, the seed-holders were placed on the lid hole of the containers. Stratified seeds were placed on the top once the agar was solid. To enhance humidity, the containers covered with cling film for a few days (4 days) until the seed germinate.

2.3.3 Stage of seedling pre-growth

When plants were directly germinated and grown in different treatments, we found that the results were unreliable due to the very large differences in size. Thus, we decided that in order to

obtain reliable results for this study, uniform growth was of great importance and should be taken into account for the experimental design. Thus, we used the pre-growth stage before treating our plants with different levels of treatments in Gibeaut's solution. 56 seeds of *Arabidopsis* plant were used for the early growth process, where the seeds grown for 10 days in hydroponic solution containing 20 μ M Pi (control) with the aim of obtaining plants showing uniform morphology. Also, to achieve a large enough set of plants for further growths under different treatments (Figure 1).

2.3.4 Stage of Treatments

In this stage, 12 uniform 10-d-old of *Arabidopsis* plants were transferred into different hydroponic systems containing various treatments: (1) 4 levels of NaCl (0.50, 100 and 150

mM), (2) 4 levels of Pi (0, 10, 100 and 250 μ M) and (3) and their interaction Pi x NaCl (4x NaCl and 4xPi, 16 processing) with or without applications of ZnO NPs and ZnO BPs (10 mg/L) in Gibeaut's solution as shown in (Table. 2). Growth parameters (root lengths, fresh and dry weight per plant and leaf area) were measured to as the role of ZnO NPs application and the role of combined effect of Pi x NaCl in reducing the negative effect of the salinity stress and Pi deficiency in *Arabidopsis* plant. To maintain constant Pi, NaCl and ZnO concentrations, if necessary, media was added every 3-4 days except on the day of nutrient application. Approximately 2 weeks after growing in Gibeaut's solution, the mature plants were harvested for the following growth parameters.

2.4 Plant growth measurements

These measurements include root lengths (cm), leaf area square centimetre (cm^2) and weights of fresh and dry per plant (g) per plant, with 3 replicates for each treatment.

Table 2: Different treatments were used in the experiments of plant growth for *Arabidopsis thaliana* Colombia-0 in hydroponic system containing Gibeaut's solution.

Treatments	
Pi treatments	
Control - Pi (0, 10, 100 and 250 μ M)	
Pi (0, 10, 100 and 250 μ M) + ZnO BPs (10 mg/L).	
Pi (0, 10, 100 and 250 μ M) + ZnO NPs (10 mg/L).	
NaCl treatments	
Control - NaCl (0, 50, 100 and 150 mM).	
NaCl (0, 50, 100 and 150 mM) + ZnO BPs (10 mg/L).	
NaCl (0, 50, 100 and 150 mM) + ZnO NPs (10 mg/L).	
Interaction between NaCl x Pi	
NaCl (50 mM)	Control - Pi (0, 10, 100 and 250 μ M)
	NaCl (50 mM) with Pi (0, 10, 100 and 250 μ M) + ZnO BPs (10 mg/L).
	NaCl (50 mM) with Pi (0, 10, 100 and 250 μ M) + ZnO NPs (10 mg/L).
NaCl (100 mM)	Control - Pi (0, 10, 100 and 250 μ M)
	NaCl (100 mM) with Pi (0, 10, 100 and 250 μ M) + ZnO BPs (10 mg/L).
	NaCl (100 mM) with Pi (0, 10, 100 and 250 μ M) + ZnO NPs (10 mg/L).
NaCl (150 mM)	Control - Pi (0, 10, 100 and 250 μ M)
	NaCl (150 mM) Pi with (0, 10, 100 and 250 μ M) + ZnO BPs (10 mg/L).
	NaCl (150 mM) with Pi (0, 10, 100 and 250 μ M) + ZnO NPs (10 mg/L).

2.4.1 Determination of root lengths

A metric scale was used to measure the plants lengths, and the results were represented in centimetres (cm).

2.4.2 Leaf area measurements

The measurement of the total rosette (leaf) surface was made using ImageJ analysis in accordance with Schnaubelt *et al.*, (2013)¹⁸. ImageJ is open-source software for processing and analysing scientific images.

2.4.3 Determination of fresh and dry weight per plant

From the hydroponic system, plants were harvested together, and the root tissues were carefully separated from the shoot tissues and washed 3 times with distilled water to remove media residue and the fresh weight was immediately determined by using analytical balance [HR-200] and expressed in grams (g). To dry shoot and root tissues, samples of dry shoots and roots were moved to a crucible that had been previously weighed and heated to 85°C in the oven for 18 hr.

2.5 Statistical analyses

Statistical Package for the Social Sciences (SPSS) version 25 was used to enter and analyse our data. All data presented are the mean values, and the standard error of the mean (\pm SE) of 3 replicates, t-test and Two-way ANOVA (Analysis of Variance) were applied to compare the average values between treatments and to analyse the effect and interaction between Pi and NaCl, with statistical significance shown at a confidence level of significant differences in $P < 0.05$.

3. Results

3.1 Root length

3.1.1 Effect of different levels of NaCl (\pm ZnO NPs-BPs) on Root length

Figure 2a shows the impact of different NaCl levels (0, 50, 100, and 150 mM) with or without 10 mg/L of ZnO NPs and ZnO BPs on the length of the roots of *Arabidopsis thaliana*. The measured values of root length significantly decreased as the NaCl concentration increased in Gibeaut's solution. A considerable reduction was found at high NaCl levels (100 and 150 mM) (5.4073 and 5.4073 cm), respectively. In comparison to the controls, the addition of ZnO BPs and ZnO NPs (10 mg/L) led to highly significant increase in root length. The root length of the plant given ZnO NPs treatment reached (5.4653 cm) at the high level of NaCl (150 mM), whereas the root length of the ZnO BPs -treated plants reached (3.9287 cm) in comparison to the control (3.3223 cm).

3.1.2 Effect of different levels of Pi (\pm ZnO NPs-BPs) on root length

At varying Pi concentrations (0, 100, and 250 μ M), *Arabidopsis thaliana* roots grew longer (Figure 2b). After 2 weeks of treatment, the root length was seen to be suppressed as Pi concentrations decreased below 10 μ M, whereas root length increased significantly at 100 and 250 μ M. Using application of ZnO BPs helped to lengthen the roots, however, ZnO NPs considerably lengthened the roots at 0, 10, and 250 μ M Pi in contrast to the control group (Figure 2b).

3.1.3 Effect of NaCl-Pi Interaction (\pm ZnO NPs-BPs) on root length

Different amounts of Pi, NaCl, and their combination had a significant impact on root length, according to a two-way ANOVA (Figure 3). Root length was significantly decreased by low Pi and salt, with salinity having a stronger impact on root growth. Salinity significantly decreased the length of *Arabidopsis thaliana*'s roots regardless of the Pi supply,

however at low Pi (10 μM) supply than at control, 100, and 250 μM Pi supply, the loss in root length was more pronounced. It is interesting to note that increasing the Pi dose in Gibeaut's solution prevents the average root height from decreasing under saline conditions. The maximum average root length values were found at the saline concentration levels of 100 and 250 μM for each saline concentration (50 and 100 mM). While, the lowest values were found at the saline concentration levels of 10 μM for each saline concentration (50, 100, and 150 mM). When ZnO BPs were added to a Gibeaut's solution that had various amounts of NaCl and Pi, the lengths of the roots significantly increased. In comparison to control, the increase was 7.9533 cm at 100 μM Pi in 50 mM NaCl and 5.8170 cm at 250 μM Pi in 100 mM NaCl, respectively (Table 2). In the Gibeaut's solution containing ZnO NPs, the maximal root length growth rates were 8.6513 cm and 5.8300 cm at 250 μM Pi in 50 and 100 mM NaCl, respectively.

3.2 Leaf area measurements

3.2.1 Effect of different levels of NaCl ($\pm\text{ZnO NPs-BPs}$) on Leaf area

The values of leaf area has decreased with all treatments of NaCl (50, 100, and 150 mM) compared to the control in treated plants (Figure 4a), indicating the negative impacts of salt stress. We observed an increase in leaf area after adding ZnO BPs and ZnO NPs at a level of 10 mg/L of each. In all NaCl concentrations, added ZnO BPs increased leaf area, however, addition of ZnO NPs significantly increased leaf area. In comparison to the control leaf area (6.4757 cm^2), ZnO BPs and ZnO NPs treatments at (100 mM NaCl) increased leaf areas to be recorded at 7.0863 cm^2 and 8.8023 cm^2 , respectively.

3.2.2 Effect of different levels of Pi ($\pm\text{ZnO NPs-BPs}$) on leaf area

The findings in Figure 4b demonstrate that *Arabidopsis* plants treated with various concentrations of Pi (10, 100, and 250 μM) tend to increase significantly in (100 and 250 μM) Pi reaching (10.3320 and 12.1080 cm^2). Whereas, the decrease was highly significant at concentrations (10 μM) Pi reaching (5.4680 cm^2), respectively. Except for the control condition, where the increase in leaf area was highly significant, the leaf area rose non-significantly for plants treated with different levels of Pi (10, 100, and 250 μM) and ZnO BPs (Figure 4b).

However, with ZnO NPs, the increase in leaf area at all concentrations was high significant, with the exception of the lowest concentration of Pi (10 μM).

3.2.3 Effect of NaCl-Pi Interaction ($\pm\text{ZnO NPs-BPs}$) on leaf area

At the 50, 100, and 150 mM NaCl levels, respectively, the leaf area rose by (9.1960, 7.0057, and 5.2693 cm^2) at Pi level (100 μM) as compared to the lowest Pi level (10 μM) (Figure 5). At the 50 and 100 mM NaCl levels, the highest Pi level (250 μM) enhanced the leaf area of plants, but not to the same extent as the 100 μM Pi did with 150 mM of NaCl. Our findings show that the leaf area decreased, when NaCl (150 mM) was applied

with the highest Pi level (250 μM). We observed that the addition of ZnO BPs and ZnO NPs (10 mg/L) markedly decreased the detrimental effects of NaCl stress and Pi deficiency (10 μM) on *Arabidopsis* plants, resulting in a notable rise in leaf area. However, plants treated with ZnO NPs showed the greatest significant increase in leaf area (Figure 5).

3.3 Fresh and dry weight per plant

3.3.1 Effect of different levels of NaCl ($\pm\text{ZnO NPs-BPs}$) on Fresh and dry weight

The fresh weight per plant was significantly decreased in all NaCl treatments after two weeks of treatment in hydroponic system (Figure 6a). In comparison to control plants (0.2637 g), plants under 150 mM of NaCl saw more losses of for fresh weight (0.0626 g). During the experiment, we observed that ZnO BPs raised the fresh weight of the shoot non-significantly in all NaCl concentrations. Whereas ZnO NPs frequently enhanced this measure by a considerable margin of significance in comparison to their respective controls (Figure 6a).

In NaCl stressed plants (0, 50, 100, and 150 mM), Figure 6b demonstrates a significant decrease in the dry weight per plant in response to salt treatments as compared to untreated plants. The reduction in the dry weight at 50 mM was significant, while it was highly significant at 100 and 150 mM. The NaCl stressed plants and treated plants with ZnO NPs or ZnO BPs significantly varied from those without both ZnO where the dry weight increased in the treated plant tissues after application of ZnO BPs and ZnO NPs. However, ZnO NP had the higher values of dry weight than ZnO BP application,

3.3.2 Effect of different levels of Pi ($\pm\text{ZnO NPs-BPs}$) on Fresh and dry weight

Figure 7a shows the impact of different Pi concentrations (0, 10, 100, and 250 μM) on the fresh weight of *Arabidopsis* plants in the presence or absence of ZnO BPs and ZnO NPs. The findings in Figure 7a demonstrate that the fresh weight per plant increased with increasing Pi level to be recorded at (0.1143, 2043, and 0.2887 g) at (0, 100, and 250 μM Pi), respectively, and subsequently dropped to (0.1060 g) at (10 μM), this value was increased to be at (0.1597 g) by applying ZnO NPs. Comparing fresh weights of *Arabidopsis* plants grown in Gibeaut's solution containing ZnO NPs and ZnO BPs (10 mg/L) to those grown in ZnO-free media, the plants grown in media with ZnO NPs and ZnO BPs showed higher fresh weights. However, plants exposed to ZnO NPs had the greatest fresh weights (0.4890 g).

The dry weight per plant significantly decreased at the lower dose (10 μM), where it was recorded at (0.01153 g). then, it significantly increased at the higher doses (100 and 250 μM), where it was recorded at (0.03190 g and 0.03553 g, respectively, in accordance with values shown in (Figure 7b). When ZnO BPs had been added to Gibeaut's solution, the dry weight increased significantly over the corresponding controls, whereas ZnO NPs significantly raised these values above the controls. At (100 and 250 μM Pi), the dry weight of plants

treated with ZnO BPs reached (0.03843 and 0.04707 g) and plants treated with ZnO NPs reached (0.04433 and 0.05277 g).

3.3.3 Effect of NaCl-Pi Interaction (\pm ZnO NPs-BPs) on Fresh and dry weight

With increasing Pi-addition treatments at all NaCl treatments, *Arabidopsis* plant fresh weight obviously increased (Figure 8). The interaction of NaCl (150 mM) and Pi (250 μ M) resulted in a fresh weight drop of (0.0470 g). while the interaction between NaCl (150 mM) and 100 μ M Pi resulted in increasing fresh weight to reach at (and 0.1057 g). Low Pi (10 μ M) significantly decreased fresh weight compared to high Pi (250 μ M) at all NaCl treatments. Applications of 10 mg/L ZnO NPs and ZnO BPs with NaCl and Pi in Gibeaut's solution had significant effects on fresh weight (Figure 8). The findings showed that the treated plants with ZnO BPs had increased fresh weight. However, plants exposed to ZnO NPs showed the greatest improvement in fresh weight (2.4 times).

With the exception of 250 μ M Pi, the dry weight per plant increased at all NaCl levels as the treatments of Pi increased (Figure 9). The interaction between NaCl (150 mM) and Pi (100 μ M) led to increase the dry weight by (0.0130 g). Whereas treatments of NaCl (150 mM) and Pi (250 μ M) decreased the dry weight by (0.0090 g). The applications of ZnO BPs and ZnO NPs along with treatments of NaCl and Pi in Gibeaut's solution had significant effects on the dry weight per plant. The results showed that plants given 10 mg/L of ZnO BP had higher dry weight values than plants who weren't given ZnO BP treatments. Obviously, plants treated with ZnO NPs and exposed to NaCl-Pi interaction had the largest improvement in dry weight per plant (Figure 9).

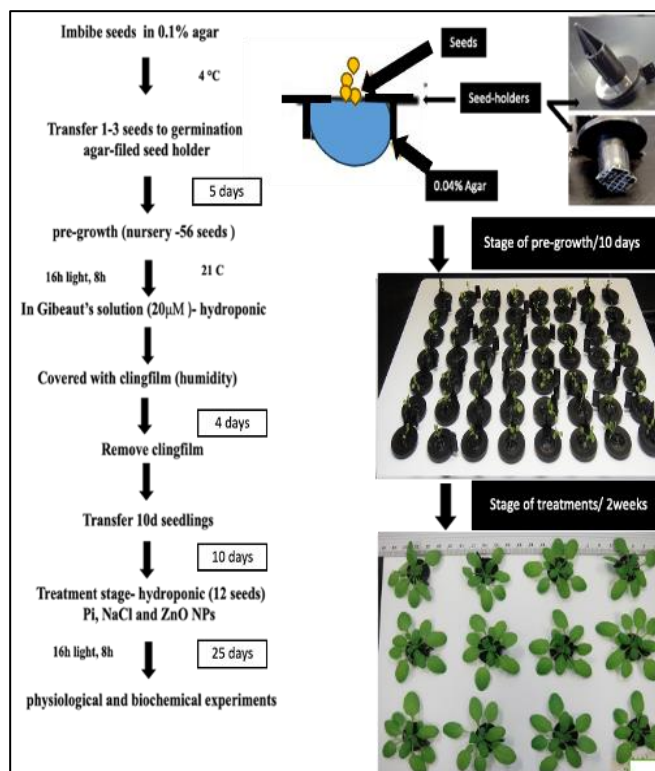


Figure 1: Method for hydroponically growing *Arabidopsis*. A flowchart detailing the process's major phases and timing. Images in the right-hand side depict the seed germination process in action as well as typical shots of seedlings and mature plants.

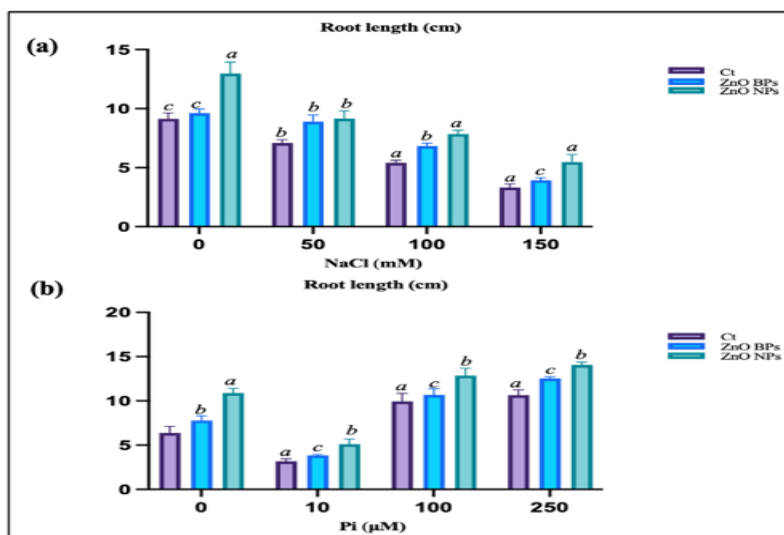


Figure 2: Effect of different concentrations of (a) NaCl (mM) and (b) Pi (μ M) +/- (ZnO NPs - ZnO BPs) on Root length (cm) of *Arabidopsis thaliana* (Col) plants grown in Gibeaut's solution in hydroponic system (means of 3 replicates \pm SE, $P \leq 0.05$), a = (highly significant), b = (significant), c = (not significant).

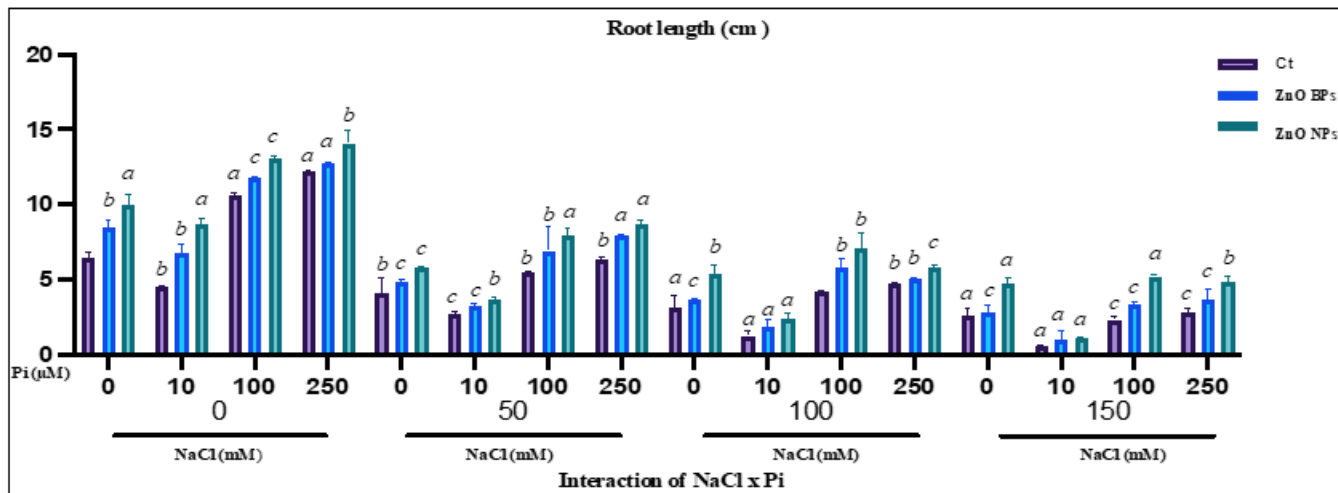


Figure 3: Effect of NaCl x Pi Interaction +/- ZnO NPs and ZnO BPs on Root length (cm) of *Arabidopsis thaliana* (Col) plants grown in Gibeaut's solution in hydroponic system (means of 3 replicates ± SE, $P \leq 0.05$), a= (highly significant), b= (significant), c= (not significant)

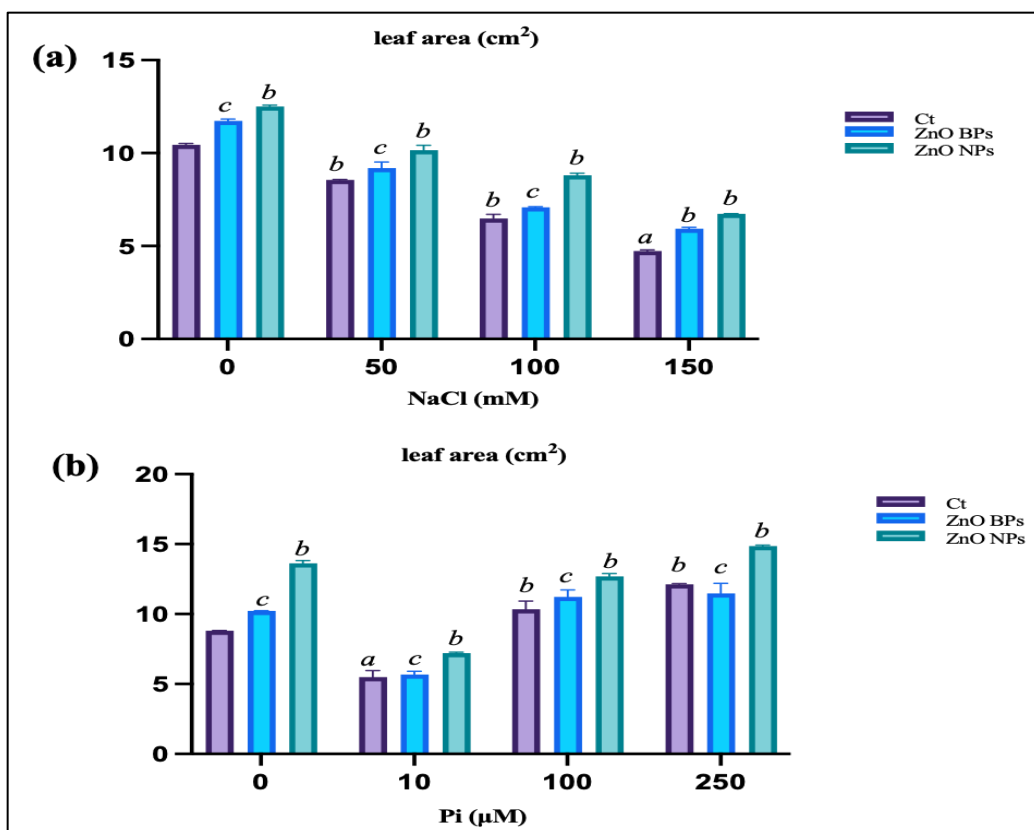


Figure 4: Effect of different concentrations of (a) NaCl (mM) and (b) Pi (μM) +/- (ZnO NPs - ZnO BPs) on Leaf area (cm²) of *Arabidopsis thaliana* (Col) plants grown in Gibeaut's solution in hydroponic system (means of 3 replicates ± SE, $P \leq 0.05$), a= (highly significant), b= (significant), c= (not significant).

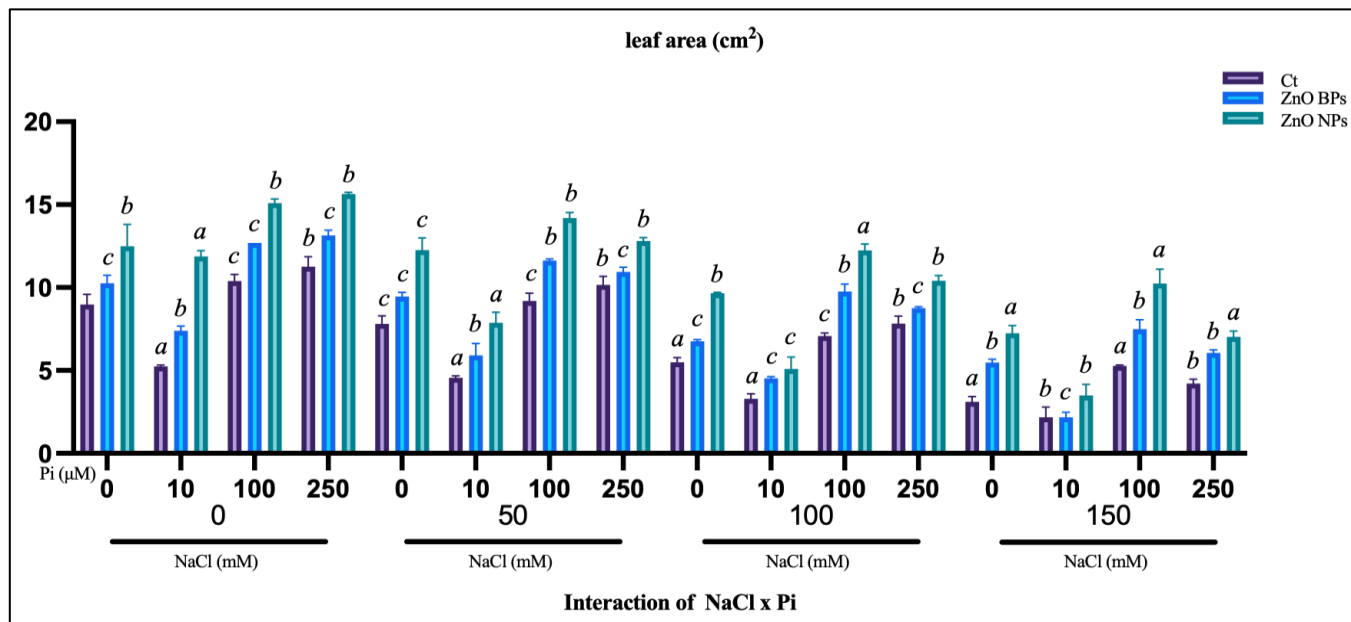


Figure 5: Effect of NaCl x Pi Interaction +/- ZnO NPs and ZnO BPs on Leaf area (cm²) of *Arabidopsis thaliana* (Col) plants grown in Gibeaut's solution in hydroponic system (means of 3 replicates ± SE, P ≤ 0.05), a= (highly significant), b= (significant), c= (not significant).

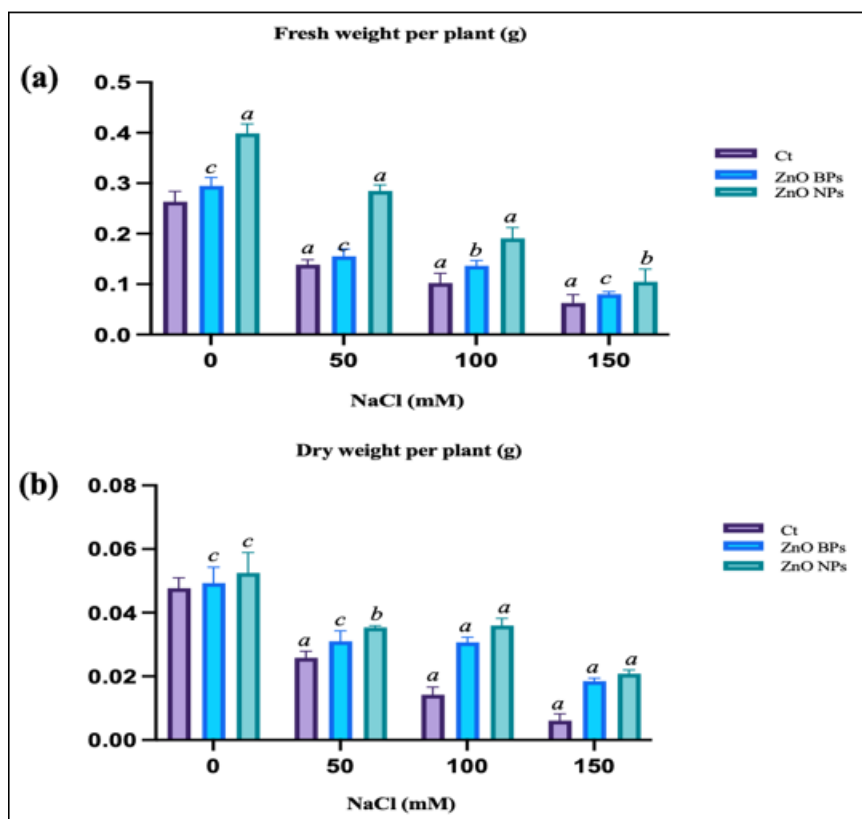


Figure 6: Effect of different concentrations of NaCl (mM) +/- (ZnO NPs) and (ZnO BPs) on (a) Fresh weight (g) and (b) Dry weight of *Arabidopsis thaliana* (Col) plants grown in Gibeaut's solution in hydroponic system (means of 3 replicates ± SE, P ≤ 0.05), a= (highly significant), b= (significant), c= (not significant).

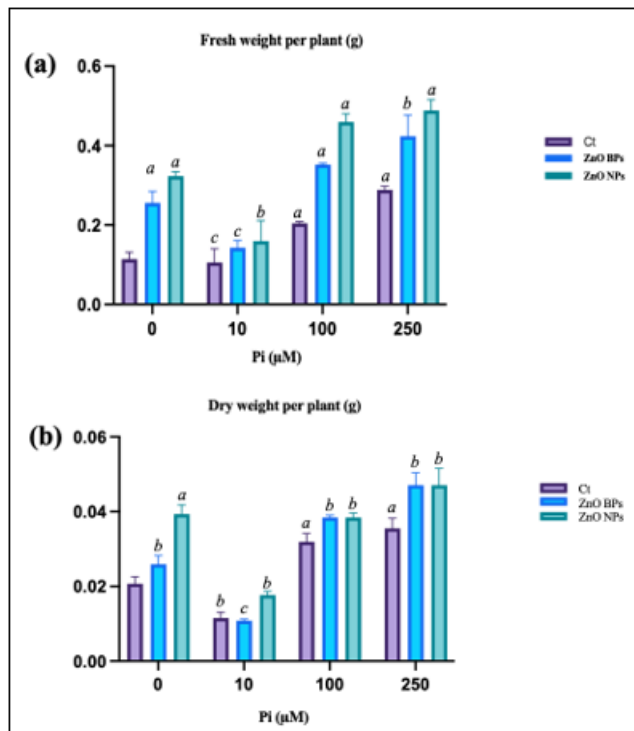


Figure 7: Effect of different concentrations of Pi (μM +/- (ZnO NPs) and (ZnO BPs) on (a) Fresh weight (g) and (b) Dry weight of *Arabidopsis thaliana* (Col) plants grown in Gibeaut's solution in hydroponic system (means of 3 replicates \pm SE, $P \leq 0.05$), $a =$ (highly significant), $b =$ (significant), $c =$ (not significant).

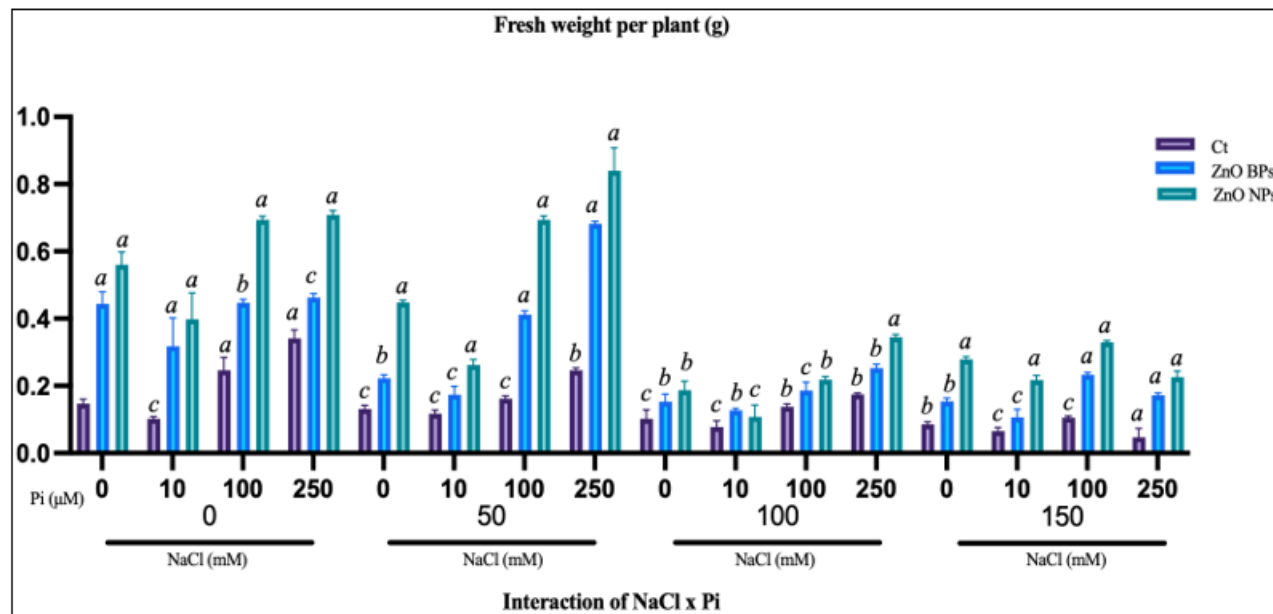


Figure 8: Effect of NaCl x Pi Interaction +/- ZnO NPs and ZnO BPs on Fresh weight (g) of *Arabidopsis thaliana* (Col) plants grown in Gibeaut's solution in hydroponic system (means of 3 replicates \pm SE, $P \leq 0.05$), $a =$ (highly significant), $b =$ (significant), $c =$ (not significant).

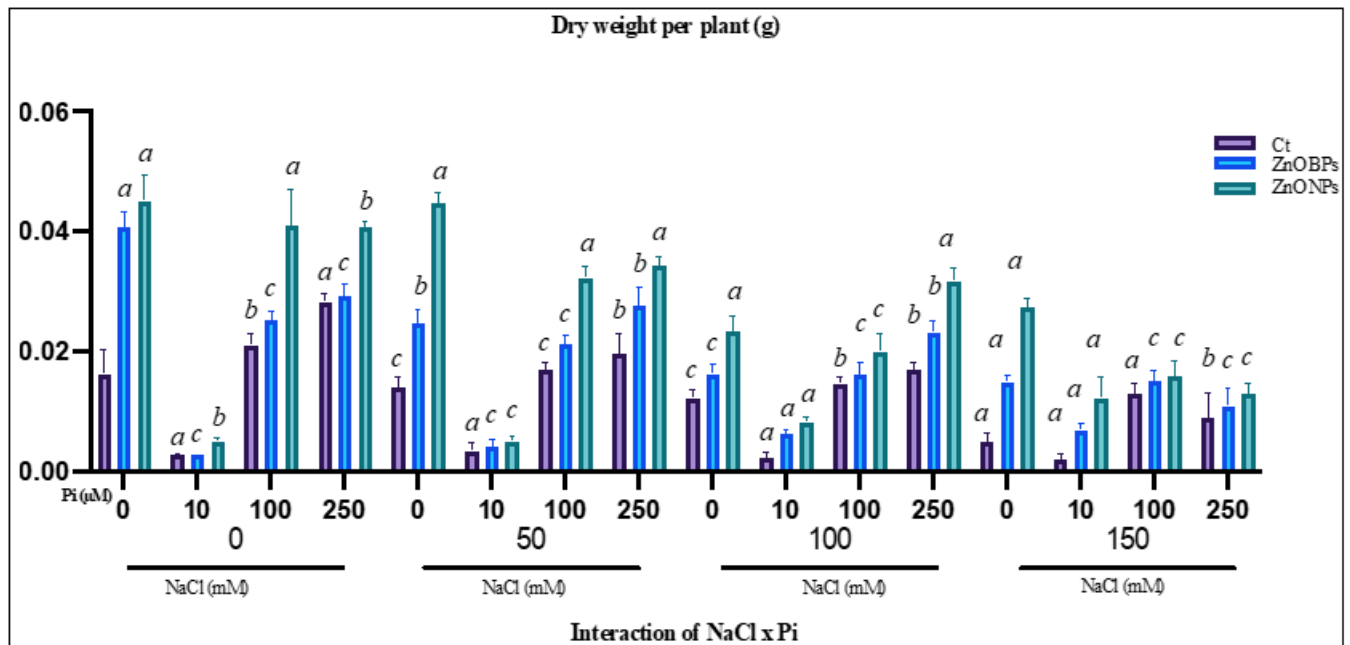


Figure 9: Effect of NaCl x Pi Interaction +/- ZnO NPs and ZnO BPs on Dry weight (g) of *Arabidopsis thaliana* (Col) plants grown in Gibeaut's solution in hydroponic system (means of 3 replicates \pm SE, $P \leq 0.05$), a= (highly significant), b= (significant), c= (not significant)

4. Discussion

Morphological traits in plants include lengths of root and shoot, weights of shoot and root and leaf area which are indicators of plant health (Rizwan *et al.*, 2017). The findings of this study indicated that different concentrations of NaCl (50, 100, and 150 mM) inhibited the development of *Arabidopsis thaliana* plants in hydroponic culture in terms of the parameters of root lengths, fresh and dry weight per plant, and leaf area. The current results aligns with some previous studies such as (Mahmoud *et al.*, 2020) on barley, (Panwar *et al.*, 2016) on chickpea, (Lai *et al.*, 2014) on alfalfa and (Abd El-Mageed *et al.*, 2020) on hot pepper. Moreover, the application of different saline water concentrations (0, 1000, 2000, and 3000 ppm) resulted in a decrease in dry weight and plant height of the herb, and root in sweet basil (Ibrahim *et al.*, 2019). In accordance with García-Caparrós and Lao, (2018), a decline in plant height and number of leaves result in a decrease in fresh or dry weight. A reduction in the total leaf area is one of the most common reactions to salt stress. As reported by Munns and Tester, (2008), changes in both cell wall properties and a decrease in photosynthetic rate are all reasons for a reduction in leaf area.

In our study, *Arabidopsis* plants cultivated hydroponically under the low Pi (10 μ M) condition failed to show much growth. However, it was noticed that the plants treated with high Pi (100 and 250 μ M) had a larger proportion of biomass in the shoot and roots. This reaction has been documented for many plants with Pi deficit as a method of overcoming the depleted zone and to obtain more P (YAO *et al.*, 2007). From the cellular to the whole plant level, Pi is a crucial component influencing the growth of plants. As stated by Assuero *et al.* (2004), Pi is essential for cell growth and division. It discovered that P-deficient leaf cells are smaller than P-sufficient leaf cells.

Because certain plants utilise Pi more effectively than others, the amount that different plants needs for growth and development vary (Vickers, 2017 ; Reich, *et al.*, 2009). Similar statistics for several species have been published by GUO *et al.*, (2012). Also, Yoneyama *et al.* (2012) reported that low levels of Pi considerably reduced the dry weight of several crops. Likewise, barley (Nadira *et al.*, 2014), Chinese fir (Xianhua *et al.*, 2018), strawberry (Valentinuzzi *et al.*, 2015), Stylosanthes (Luo *et al.*, 2020), maize (Li *et al.*, 2012), and tomato (Schausberger, 2018) all had decreased growth in their shoots due to a Pi deficiency.

Plants are frequently forced to choose between two stressors when they are simultaneously subjected to salt and deficiency in nutrients, and their growth and development are governed by the most limiting factor of growth (Van der Ploeg, 1999). The results of our study support this argument since growth of *Arabidopsis thaliana* was significantly reduced under high salinity compared to 10 μ M of Pi deficiency, indicating that salinity stress affects *Arabidopsis* growth more than Pi shortage. Our findings are corresponding with earlier research in *Hordeum vulgare* (Talbi Zribi *et al.*, 2011; Zribi *et al.*, 2015) and in *Lycopersicon esculentum* (Mohammad *et al.*, 1998). However, our findings do not coincide with those reported in previous published research by Rogers *et al.*, (2003) which revealed that organs of shoots are the more negatively impacted by deficiency of nutrient than salinity stress. On other side, our findings indicated that the growth parameters values of *Arabidopsis* plant was higher when it was treated with high Pi (100 μ M) at all NaCl concentrations, these results are in line with those of Kaya *et al.* (2003), who found that adding additional Pi to soil at a concentration of 272 mg might partially alleviate the detrimental effects of excessive salinity on pepper and cucumber. According to Erdal *et al.* (2011), the decreased

in fresh and dry weight, leaf area and root length at high NaCl concentration may be the result of changes in plant metabolism in reaction to salt. Consequently, numerous researchers (Kaya *et al.*, 2001) ; Gulmezoglu and Daghan, 2017) have reported on the impact of Pi treatment in reducing the negative effects of salinity on plants. Aslam *et al.* (1996) stated that Pi had positive impact on rice development when salt is present in the media. Likewise, Kaya *et al.* (2001) insured that Pi treatment reduced the detrimental impacts of salinity stress on growth of plants. These researchers observed that the growth of rice at (50 mM NaCl) increased by increasing exogenous P, level up to 100 (mM). Application of 18 kg of P to pots with saline soil also led to a notable increase in paddy production (Singh, 1988). Any further addition of P, led to a decrease in growth plant (paddy and straw). In the findings of Roberts *et al.* (1984), this is a consequence of the build-up of toxic amounts of P in plant tissues, which is considered to be an evidence of P toxicity caused by salt. These outcomes partly support the conclusions we reached in our investigation. In contrast, as stated by Abbas *et al.* (2018), that low P (10 M) is more damaging to shoot and root development than 100 mM NaCl (medium salinity). This conclusion validates the results we obtained from treating our plants under the same conditions. According to (Singh *et al.*, 2018), plants with P amendments in greenhouse circumstances exhibited better growth metrics than plants under salt stress without P fertilization. Therefore, it is necessary to increase the availability of Pi to plants under salinity conditions for osmotic adjustment and growth. Sadjji-Ait Kaci *et al.* (2017) indicated that applying Pi to salinized soil may improve the field's situation by restoring ionic and osmotic balance in the rhizosphere. Also, they stated that applying P to salinized soil may improve the field's situation by restoring ionic and osmotic balance in the rhizosphere, which may have lessened and/or inhibited the salinity's severe impacts.

In our findings, the treatment of ZnO NPs and ZnO BPs under salt stress enhanced the measured growth parameters in Arabidopsis plants. However, application of ZnO NPs performed more effective than ZnO BPs which are consistent with conclusion of study carried out by Torabian *et al.* (2016). They noticed that the use of ZnO NPs boosted the leaf area and shoot dry weight in salt-stressed sunflower plants and produced greater results than the normal ZnO application approach. Also, our findings concur with those reported in other studies that have been published, including those on strawberries (Luksiene *et al.*, 2020), and maize (Shinde *et al.*, 2020). As previously stated by Sturikova *et al.* (2018)⁸⁷, ZnO NPs may be able to reduce the harmful impacts of abiotic stress on development and growth of plants.

This increased effect of application is attributed to the essential function of zinc in biological and metabolic processes, including enzyme activity, nitrogen metabolism, photosynthetic pigments and accumulation of phospholipids (Mehrabani, *et al.*, 2018). Regarding interaction between Pi and ZnONPs, our findings are at contrast with those of Dimkpa *et al.* (2012); Stewart *et al.* (2015), and Watson *et al.* (2015), who reported that *T. aestivum's* growth decreased when it was treated with Pi and exposed to ZnONPs. Furthermore, studies

by Zafar *et al.* (2016) and Mukherjee *et al.* (2016) demonstrated similar growth inhibition in *Pisum sativum* and Brassica species treated to ZnONPs. However, in accordance with the combination of P and Zn caused a significant increase in the dry matter of chia due to the treatment of P at 100 mg kg⁻¹ soil coupled with Zn at 10 mg kg⁻¹ soil, which is consistent with our findings. Similar findings indicated that when Zn and P were applied together rather than separately, growth and yield characteristics in maize genotypes were enhanced (Imran *et al.*, 2016). These results are consistent with the findings of Ova *et al.* (2015), who demonstrated that P and Zn had beneficial interactions in wheat yield. They also demonstrated that Zn application did not increase dry matter at low P levels, but plants generated greater yield when exposed to increased Zn at high P levels. Similar findings were found in other crops as well, including maize (Amanullah *et al.*, 2016), rice (Amanullah and Inamullah, 2016), bean (Gianquinto *et al.*, 2000) and cowpea (Kumar *et al.*, 2016), showing that combined Zn and Pi enhanced dry biomass production.

5. Conclusion

Proper amount of Pi improved the salt tolerance of *Arabidopsis* plants by enhancing exclusion of Na. Also, the results showed that in the combined treatment of Pi and NaCl was more effective than in the single treatment. ZnO-NPs application in low doses (10 mg/L) is an eco-friendly, cheap cost, and can be considered as a promising application to alleviate the harmful impacts of salt and stress of mineral on plants. In sum, our result provides new alternative strategies for increasing crop productivity which are hydroponic growth allowing for more effective mineral absorption, interaction NaCl x Pi and Pi treatments as the biochemical fertilizers and ZnO NPs as nanofertilizers, all can be considered as powerful tools to boost plant growth under abiotic stress. This study is promising to open a new path for nanotechnology particularly in Saudi Arabian context as per the kingdom's vision 2030 and Saudi green initiative.

References

- [1] Abbas, Ghulam, Yinglong Chen, Faisal Younus Khan, Yupeng Feng, Jairo A. Palta, and Kadambot H. M. Siddique. 2018. "Salinity and Low Phosphorus Differentially Affect Shoot and Root Traits in Two Wheat Cultivars with Contrasting Tolerance to Salt." *Agronomy* 8(8):155.
- [2] Abd El-Mageed, Taia A., Mostafa M. Rady, Ragab S. Taha, Sayed Abd El Azeam, Catherine R. Simpson, and Wael M. Semida. 2020. "Effects of Integrated Use of Residual Sulfur-Enhanced Biochar with Effective Microorganisms on Soil Properties, Plant Growth and Short-Term Productivity of Capsicum Annuum under Salt Stress." *Scientia Horticulturae* 261:108930.
- [3] Amanullah, Inamullah, and X. Inamullah. 2016. "Dry Matter Partitioning and Harvest Index Differ in Rice Genotypes with Variable Rates of Phosphorus and Zinc Nutrition." *Rice Sci* 23(2):78–87.
- [4] Amanullah, Saleem A., A. Iqbal, and S. Fahad. 2016.

- “Foliar Phosphorus and Zinc Application Improve Growth and Productivity of Maize (*Zea Mays* L.) under Moisture Stress Conditions in Semi-Arid Climates.” *J Microb Biochem Technol* 8(5).
- [5] Aslam, M., T. J. Flowers, R. H. Qureshi, and A. R. Yeo. 1996. “Interaction of Phosphate and Salinity on the Growth and Yield of Rice (*Oryza Sativa* L.)” *Journal of Agronomy and Crop Science* 176(4):249–58.
- [6] Assuero, S. G., Alain Mollier, and Sylvain Pellerin. 2004. “The Decrease in Growth of Phosphorus-deficient Maize Leaves Is Related to a Lower Cell Production.” *Plant, Cell & Environment* 27(7):887–95.
- [7] Cordell, Dana, Jan-Olof Drangert, and Stuart White. 2009. “The Story of Phosphorus: Global Food Security and Food for Thought.” *Global Environmental Change* 19(2):292–305.
- [8] Das, Sumistha, Biswarup Sen, and Nitai Debnath. 2015. “Recent Trends in Nanomaterials Applications in Environmental Monitoring and Remediation.” *Environmental Science and Pollution Research* 22(23):18333–44.
- [9] Dey, Gobinda, Pritam Banerjee, Raju Kumar Sharma, Jyoti Prakash Maity, Hassan Etesami, Arun Kumar Shaw, Yi Hsun Huang, Hsien Bin Huang, and Chien Yen Chen. 2021. “Management of Phosphorus in Salinity-Stressed Agriculture for Sustainable Crop Production by Salt-Tolerant Phosphate-Solubilizing Bacteria—a Review.” *Agronomy* 11(8). doi: 10.3390/agronomy11081552.
- [10] Dimkpa, Christian O., Joan E. McLean, Drew E. Latta, Eliana Manangón, David W. Britt, William P. Johnson, Maxim I. Boyanov, and Anne J. Anderson. 2012. “CuO and ZnO Nanoparticles: Phytotoxicity, Metal Speciation, and Induction of Oxidative Stress in Sand-Grown Wheat.” *Journal of Nanoparticle Research* 14:1–15.
- [11] Elemike, Elias E., Ifeyinwa Monica Uzoh, Damian C. Onwudiwe, and Olubukola Oluranti Babalola. 2019. “The Role of Nanotechnology in the Fortification of Plant Nutrients and Improvement of Crop Production.” *Applied Sciences* 9(3):499.
- [12] Erdal, Serkan, M. Aydın, M. Genisel, M. S. Taspınar, Rahmi Dumlupınar, O. Kaya, and Z. Gorcek. 2011. “Effects of Salicylic Acid on Wheat Salt Sensitivity.” *African Journal of Biotechnology* 10(30):5713–18.
- [13] García-Caparrós, Pedro, and María Teresa Lao. 2018. “The Effects of Salt Stress on Ornamental Plants and Integrative Cultivation Practices.” *Scientia Horticulturae* 240:430–39.
- [14] Gianquinto, Giorgio, Azmi Abu-Rayyan, Livia Di Tola, Diletta Piccotino, and Beatrice Pezzarossa. 2000. “Interaction Effects of Phosphorus and Zinc on Photosynthesis, Growth and Yield of Dwarf Bean Grown in Two Environments.” *Plant and Soil* 220(1–2):219–28.
- [15] Gibeaut, David M., John Hulett, Grant R. Cramer, and Jeffrey R. Seemann. 1997. “Maximal Biomass of *Arabidopsis Thaliana* Using a Simple, Low-Maintenance Hydroponic Method and Favorable Environmental Conditions.” *Plant Physiology* 115(2):317.
- [16] Gulmezoglu, N., and H. Daghan. 2017. “The Interactive Effects of Phosphorus and Salt on Growth, Water Potential and Phosphorus Uptake in Green Beans.” *Appl. Ecol. Environ. Res* 15(3):1831–42.
- [17] GUO, Tian-rong, Peng-cheng YAO, Zi-dong ZHANG, Jiang-jia WANG, and WANG Mei. 2012. “Involvement of Antioxidative Defense System in Rice Seedlings Exposed to Aluminum Toxicity and Phosphorus Deficiency.” *Rice Science* 19(3):207–12.
- [18] Ibrahim, Ayat M. M., A. E. Awad, A. S. H. Gendy, and M. A. I. Abdelkader. 2019. “Effect of Proline Foliar Spray on Growth and Productivity of Sweet Basil (*Ocimum Basilicum*, L.) Plant under Salinity Stress Conditions.” *Zagazig Journal of Agricultural Research* 46(6):1877–89.
- [19] Imran, Muhammad, Abdur Rehim, Shahid Hussain, Muhammad Zafar ul Hye, and Hafeez Ur Rehman. 2016. “Efficiency of Zinc and Phosphorus Applied to Open-Pollinated and Hybrid Cultivars of Maize.” *Int J Agric Biol* 18(6):1249–55.
- [20] James, Richard A., Carol Blake, Caitlin S. Byrt, and Rana Munns. 2011. “Major Genes for Na⁺ Exclusion, Nax1 and Nax2 (Wheat HKT1; 4 and HKT1; 5), Decrease Na⁺ Accumulation in Bread Wheat Leaves under Saline and Waterlogged Conditions.” *Journal of Experimental Botany* 62(8):2939–47.
- [21] Kapri, Anil, and Lakshmi Tewari. 2010. “Phosphate Solubilization Potential and Phosphatase Activity of Rhizospheric *Trichoderma* Spp.” *Brazilian Journal of Microbiology* 41(3):787–95.
- [22] Kaya, Cengiz, David Higgs, Faruk Ince, Bernado Murillo Amador, Atilla Cakir, and Ebru Sakar. 2003. “Ameliorative Effects of Potassium Phosphate on Salt-stressed Pepper and Cucumber.” *Journal of Plant Nutrition* 26(4):807–20.
- [23] Kaya, Cengiz, David Higgs, and Halil Kirnak. 2001. “The Effects of High Salinity (NaCl) and Supplementary Phosphorus and Potassium on Physiology and Nutrition Development of Spinach.” *Bulg. J. Plant Physiol* 27(3–4):47–59.
- [24] Kumar, Rakesh, Deepak Kumar Rathore, Magan Singh, Parveen Kumar, and Anil Khippal. 2016. “Effect of Phosphorus and Zinc Nutrition on Growth and Yield of Fodder Cowpea.” *Legume Research-An International Journal* 39(2):262–67.
- [25] Lai, Diwen, Yu Mao, Heng Zhou, Feng Li, Mingzhu Wu, Jing Zhang, Ziyi He, Weiti Cui, and Yanjie Xie. 2014. “Endogenous Hydrogen Sulfide Enhances Salt Tolerance by Coupling the Reestablishment of Redox Homeostasis and Preventing Salt-Induced K⁺ Loss in Seedlings of *Medicago Sativa*.” *Plant Science* 225:117–29.
- [26] Lang, Claudia, Elaine Gabutin Mission, Abdullah Al-Hadi Ahmad Fuaad, and Mohamed Shaalan. 2021. “Nanoparticle Tools to Improve and Advance Precision Practices in the Agrifoods Sector towards Sustainability—A Review.” *Journal of Cleaner Production* 293:126063.
- [27] Li, Zhaoxia, Changzheng Xu, Kunpeng Li, Shi Yan, Xun Qu, and Juren Zhang. 2012. “Phosphate Starvation of

- Maize Inhibits Lateral Root Formation and Alters Gene Expression in the Lateral Root Primordium Zone.” *BMC Plant Biology* 12:1–17.
- [28] Luksiene, Zivile, Neringa Rasiukeviciute, Bernadeta Zudyte, and Nobertas Uselis. 2020. “Innovative Approach to Sunlight Activated Biofungicides for Strawberry Crop Protection: ZnO Nanoparticles.” *Journal of Photochemistry and Photobiology B: Biology* 203:111656.
- [29] Luo, Jiajia, Yunxi Liu, Huikai Zhang, Jinpeng Wang, Zhijian Chen, Lijuan Luo, Guodao Liu, and Pandao Liu. 2020. “Metabolic Alterations Provide Insights into Stylosanthes Roots Responding to Phosphorus Deficiency.” *BMC Plant Biology* 20(1):1–16.
- [30] Mahapatra, Durga Madhab, Kanhu Charan Satapathy, and Bhabatarini Panda. 2022. “Biofertilizers and Nanofertilizers for Sustainable Agriculture: Phycopropects and Challenges.” *Science of the Total Environment* 803:149990.
- [31] Mahmoud, O. Metoui Ben, R. Hidri, O. Talbi-Zribi, W. Taamalli, C. Abdelly, and N. Djébal. 2020. “Auxin and Proline Producing Rhizobacteria Mitigate Salt-Induced Growth Inhibition of Barley Plants by Enhancing Water and Nutrient Status.” *South African Journal of Botany* 128:209–17.
- [32] Mehrabani, Lamia Vojodi, Mohammad Bagher Hassanpouraghdam, and Tahereh Shamsi-Khotab. 2018. “The Effects of Common and Nano-Zinc Foliar Application on the Alleviation of Salinity Stress in *Rosmarinus Officinalis* L.” *ACTA Scientiarum Polonorum Hortorum Cultus* 17(6):65–73.
- [33] Mohammad, M., R. Shibli, M. Ajlouni, and L. Nimri. 1998. “Tomato Root and Shoot Responses to Salt Stress under Different Levels of Phosphorus Nutrition.” *Journal of Plant Nutrition* 21(8):1667–80.
- [34] Mukherjee, Arnab, Youping Sun, Erving Morelius, Carlos Tamez, Susmita Bandyopadhyay, Genhua Niu, Jason C. White, Jose R. Peralta-Videa, and Jorge L. Gardea-Torresdey. 2016.
- [35] “Differential Toxicity of Bare and Hybrid ZnO Nanoparticles in Green Pea (*Pisum Sativum* L.): A Life Cycle Study.” *Frontiers in Plant Science* 6:1242.
- [36] Munns, Rana, and Mark Tester. 2008. “Mechanisms of Salinity Tolerance.” *Annu. Rev. Plant Biol.* 59:651–81.
- [37] Nadira, Umme Aktari, Imrul Mosaddek Ahmed, Jianbin Zeng, Noreen Bibi, Shengguan Cai, Feibo Wu, and Guoping Zhang. 2014. “The Changes in Physiological and Biochemical Traits of Tibetan Wild and Cultivated Barley in Response to Low Phosphorus Stress.” *Soil Science and Plant Nutrition* 60(6):832–42.
- [38] Ova, Emir Ali, Umit Baris Kutman, Levent Ozturk, and Ismail Cakmak. 2015. “High Phosphorus Supply Reduced Zinc Concentration of Wheat in Native Soil but Not in Autoclaved Soil or Nutrient Solution.” *Plant and Soil* 393:147–62.
- [39] Panwar, Meenu, Rupinder Tewari, Arvind Gulati, and Harsh Nayyar. 2016. “Indigenous Salt-Tolerant Rhizobacterium *Pantoea Dispersa* (PSB3) Reduces Sodium Uptake and Mitigates the Effects of Salt Stress on Growth and Yield of Chickpea.” *Acta Physiologiae Plantarum* 38:1–12.
- [40] Van der Ploeg, R. R. n.d. “Bé Ohm, W., and Kirkham, MB (1999) On the Origin of the Theory of Mineral Nutrition of Plants and the Law of the Minimum.” *Soil Sci Soc Am J* 63:1055–62.
- [41] Reich, Peter B., Jacek Oleksyn, and Ian J. Wright. 2009. “Leaf Phosphorus Influences the Photosynthesis–Nitrogen Relation: A Cross-Biome Analysis of 314 Species.” *Oecologia* 160:207–12.
- [42] Rizwan, Muhammad, Shafaqat Ali, Muhammad Farooq Qayyum, Yong Sik Ok, Muhammad Adrees, Muhammad Ibrahim, Muhammad Zia-ur-Rehman, Mujahid Farid, and Farhat Abbas. 2017. “Effect of Metal and Metal Oxide Nanoparticles on Growth and Physiology of Globally Important Food Crops: A Critical Review.” *Journal of Hazardous Materials* 322:2–16.
- [43] Roberts, Justin K. M., Carey S. Linker, Anthony G. Benoit, Oleg Jardetzky, and Richard H. Nieman. 1984. “Salt Stimulation of Phosphate Uptake in Maize Root Tips Studied by ³¹P Nuclear Magnetic Resonance.” *Plant Physiology* 75(4):947–50.
- [44] Rogers, M. E., C. M. Grieve, and M. C. Shannon. 2003. “Plant Growth and Ion Relations in Lucerne (*Medicago Sativa* L.) in Response to the Combined Effects of NaCl and P.” *Plant and Soil* 253:187–94.
- [45] Sadji-Ait Kaci, H., A. Chaker-Haddadj, and F. Aid. 2017. “Interactive Effects of Salinity and Two Phosphorus Fertilizers on Growth and Grain Yield of *Cicer Arietinum* L.” *Acta Agriculturae Scandinavica, Section B—Soil & Plant Science* 67(3):208–16.
- [46] Schausberger, Peter. 2018. “Herbivore-Associated Bacteria as Potential Mediators and Modifiers of Induced Plant Defense against Spider Mites and Thrips.” *Frontiers in Plant Science* 9:1107.
- [47] Schnaubelt, Daniel, Philipp Schulz, Matthew A. Hannah, Rosita E. Yocgo, and Christine H. Foyer. 2013. “A Phenomics Approach to the Analysis of the Influence of Glutathione on Leaf Area and Abiotic Stress Tolerance in *Arabidopsis Thaliana*.” *Frontiers in Plant Science* 4:416.
- [48] Shinde, Surbhi, Priti Paralikar, Avinash P. Ingle, and Mahendra Rai. 2020. “Promotion of Seed Germination and Seedling Growth of *Zea Mays* by Magnesium Hydroxide Nanoparticles Synthesized by the Filtrate from *Aspergillus Niger*.” *Arabian Journal of Chemistry* 13(1):3172–82.
- [49] Shokat, Sajid, and Dominik K. Großkinsky. 2019. “Tackling Salinity in Sustainable Agriculture—What Developing Countries May Learn from Approaches of the Developed World.” *Sustainability* 11(17):4558.
- [50] Singh, M. V. 1988. “Response of Rice (*Oryza Sativa*) and Wheat (*Triticum Aestivum*) to Phosphorus and Zinc Fertilization in Sodic Soil.” *Indian Journal of Agricultural Science* 58(11):823–27.
- [51] Singh, Shardendu K., Vangimalla R. Reddy, David H. Fleisher, and Dennis J. Timlin. 2018. “Phosphorus Nutrition Affects
- [52] Temperature Response of Soybean Growth and Canopy

- Photosynthesis.” *Frontiers in Plant Science* 9:1116.
- [53] Srivastava, Neerja. 2020. “Reclamation of Saline and Sodic Soil through Phytoremediation.” Pp. 279–306 in *Environmental concerns and sustainable development*. Springer.
- [54] Stewart, Jacob, Trevor Hansen, Joan E. McLean, Paul McManus, Siddhartha Das, David W. Britt, Anne J. Anderson, and Christian O. Dimkpa. 2015. “Salts Affect the Interaction of ZnO or CuO Nanoparticles with Wheat.” *Environmental Toxicology and Chemistry* 34(9):2116–25.
- [55] Sturikova, Helena, Olga Krystofova, Dalibor Huska, and Vojtech Adam. 2018. “Zinc, Zinc Nanoparticles and Plants.” *Journal of Hazardous Materials* 349:101–10.
- [56] Syers, J. K., A. E. Johnston, and D. Curtin. 2008. “Efficiency of Soil and Fertilizer Phosphorus Use.” *FAO Fertilizer and Plant Nutrition Bulletin* 18.
- [57] Talbi Zribi, O., C. Abdelly, and A. Debez. 2011. “Interactive Effects of Salinity and Phosphorus Availability on Growth, Water Relations, Nutritional Status and Photosynthetic Activity of Barley (*Hordeum Vulgare* L.)” *Plant Biology* 13(6):872–80.
- [58] Tchakounté, Gylaine Vanissa Tchuisseu, Beatrice Berger, Sascha Patz, Matthias Becker, Henri Fankem, Victor Désiré Taffouo, and Silke Ruppel. 2020. “Selected Rhizosphere Bacteria Help Tomato Plants Cope with Combined Phosphorus and Salt Stresses.” *Microorganisms* 8(11):1844.
- [59] Thounaojam, Thorny Chanu, Thounaojam Thomas Meetei, Yumnam Bijilaxmi Devi, Sanjib Kumar Panda, and Hrishikesh Upadhyaya. 2021. “Zinc Oxide Nanoparticles (ZnO-NPs): A Promising Nanoparticle in Renovating Plant Science.” *Acta Physiologiae Plantarum* 43(10):1–21.
- [60] Torabian, Shahram, Morteza Zahedi, and Amir Hossein Khoshgoftar. 2016. “Effects of Foliar Spray of Two Kinds of Zinc Oxide on the Growth and Ion Concentration of Sunflower Cultivars under Salt Stress.” *Journal of Plant Nutrition* 39(2):172–80.
- [61] Usman, Muhammad, Muhammad Farooq, Abdul Wakeel, Ahmad Nawaz, Sardar Alam Cheema, Hafeez ur Rehman, Imran Ashraf, and Muhammad Sanaullah. 2020. “Nanotechnology in Agriculture: Current Status, Challenges and Future Opportunities.” *Science of the Total Environment* 721:137778.
- [62] Valentinuzzi, Fabio, Youry Pii, Gianpiero Vigani, Martin Lehmann, Stefano Cesco, and Tanja Mimmo. 2015. “Phosphorus and Iron Deficiencies Induce a Metabolic Reprogramming and Affect the Exudation Traits of the Woody Plant *Fragaria* × *Ananassa*.” *Journal of Experimental Botany* 66(20):6483–95.
- [63] Vermeulen, Sonja Joy, Pramod K. Aggarwal, Andrew Ainslie, C. Angelone, Bruce M. Campbell, Andrew J. Challinor, James W. Hansen, J. S. I. Ingram, Andrew Jarvis, and P. Kristjansson. 2012. “Options for Support to Agriculture and Food Security under Climate Change.” *Environmental Science & Policy* 15(1):136–44.
- [64] Vickers, Neil J. 2017. “Animal Communication: When i’m Calling You, Will You Answer Too?” *Current Biology* 27(14):R713–15.
- [65] Watson, Jean-Luc, Tommy Fang, Christian O. Dimkpa, David W. Britt, Joan E. McLean, Astrid Jacobson, and Anne J. Anderson. 2015. “The Phytotoxicity of ZnO Nanoparticles on Wheat Varies with Soil Properties.” *Biometals* 28:101–12.
- [66] Xianhua, Zou, Wei Dan, Wu Pengfei, Zhang Ying, Hu Yanan, Chen Sitong, and Ma Xiangqing. 2018. “Strategies of Organic Acid Production and Exudation in Response to Low-Phosphorus Stress in Chinese Fir Genotypes Differing in Phosphorus-Use Efficiencies.” *Trees* 32:897–912.
- [67] YAO, Qi-lun, Ke-cheng YANG, Guang-tang PAN, and Ting-zhao RONG. 2007. “The Effects of Low Phosphorus Stress on Morphological and Physiological Characteristics of Maize (*Zea Mays* L.) Landraces.” *Agricultural Sciences in China* 6(5):559–66.
- [68] Yoneyama, Kaori, Xiaonan Xie, Hyun Il Kim, Takaya Kisugi, Takahito Nomura, Hitoshi Sekimoto, Takao Yokota, and Koichi Yoneyama. 2012. “How Do Nitrogen and Phosphorus Deficiencies Affect Strigolactone Production and Exudation?” *Planta* 235:1197–1207.
- [69] Zafar, Hira, Attarad Ali, Joham S. Ali, Ihsan U. Haq, and Muhammad Zia. 2016. “Effect of ZnO Nanoparticles on Brassica Nigra Seedlings and Stem Explants: Growth Dynamics and Antioxidative Response.” *Frontiers in Plant Science* 7:535.
- [70] Zribi, Ons Talbi, Zouhaier Barhoumi, Saber Kouas, Mohamed Ghandour, Ines Slama, and Chedly Abdelly. 2015. “Insights into the Physiological Responses of the Facultative Halophyte *Aeluropus Littoralis* to the Combined Effects of Salinity and Phosphorus Availability.” *Journal of Plant Physiology* 189:1–10