

Optimizing Amphotericin B Invasomes: Critical Quality Attributes using Plackett Burman Design

Priyank Kumar M. Patel¹, Mansiben M. Patel², Chairesh N. Shah³, Umesh M. Upadhyay⁴

^{1, 2, 3, 4}Sigma Institute of Pharmacy, Sigma University, Vadodara - 390019, Gujarat, India

³Corresponding Author Email: [drdrchaireshshah\[at\]gmail.com](mailto:drdrchaireshshah[at]gmail.com)

Abstract: *This study investigates the development and optimization of Amphotericin B loaded invasomes for enhanced transdermal drug delivery. Invasomes (IVS) are the novel vesicular carrier of drugs for transdermal delivery and enhanced penetration than conventional liposomes. IVS are flexible vesicles and composed of phospholipids, alcohol, and terpenes. Invasomes were prepared using the mechanical Dispersion method. Plackett - Burman design is used as screening method to identify the critical factors using 12 runs with 7 variables. Hence, critical quality attributes (CQAs) such as phospholipid concentration, terpene concentration, and sonication speed were identified and optimized to maximize entrapment efficiency and control vesicle size. Results showed that the entrapment efficiency (%) and vesicle size (nm) across the 12 batches ranged from 60.63 to 96.82% and 73 to 306 nm, respectively which demonstrated significant improvements in drug entrapment and vesicle size, offering a promising approach for effective transdermal delivery of Amphotericin B.*

Keywords: Quality by Design, Invasomes, Amphotericin B, Topical Candidiasis, Mechanical Dispersion, Plackett Burman Design, Critical Quality Attributes, Critical Process Parameters, Critical Material Attributes

1. Introduction

Pharmaceutical QbD is a systematic, scientific, risk - based, holistic and proactive approach to pharmaceutical development that begins with predefined objectives and emphasizes product and processes understanding and process control. [1] Amphotericin B is an effective polyene antifungal considered as a “gold standard” in the management of fungal infections and BCS class – IV drug. Amphotericin B is having oral bioavailability 0.2 to 0.9% and 100% upon IV administration. Oral Half - life is 7 - 8 hours. Amphotericin B Aqueous Solubility is 0.086 mg/ml and Log P value of Amphotericin B is 5.83. [2] Intravenous administration paradoxically leads to side effects that have not been fully mitigated, even with newer formulations. Thus, the need for alternative formulations/route of administration for amphotericin B remains crucial. [3] However, the commercial topical preparation is not significantly properly permeated or absorbed through the skin due to low residence time or less crossing of the stratum corneum layer of the skin. Thus, it has been required to develop a novel drug delivery system (nanotechnology - based) for the improvement of the therapeutic activity of drugs. [4]

Invasomes (IVS) are the novel vesicular carrier of drugs for transdermal delivery and enhanced penetration than conventional liposomes. [5 - 6] IVS is flexible vesicles and composed of phospholipids, alcohol, and terpenes. Alcohol has the property that enhances the flexibility of lipids bilayer of IVS, producing soft vesicles leading to increasing skin permeability of the drug. [7] Ethanol can fluidize and disturb the stratum corneum lipid leading to an increase in the permeability of the drug. Terpenes also enhance the skin permeation of both lipophilic and hydrophilic drugs by re - arrangement of the stratum corneum layer of skin. Incorporating IVS into the gel system provided the sustained drug release, increased skin contact time and enhanced penetration. [8] There are many reports published on IVS gel, which improved the transdermal delivery of drugs.

Purpose Statement

This study aims to optimize the formulation of Amphotericin B loaded invasomes by identifying and controlling critical quality attributes using the Plackett Burman design.

Significance of research work

The study significance lies in its potential to enhance the effectiveness of Amphotericin B as a transdermal drug, offering an alternative to traditional administration routes, thereby reducing side effects and improving patient outcomes.

2. Materials and Methods

Amphotericin B was received as gift sample from Kivi Lab, Por, India. Phospholipids such as Soyaphosphatidylcholine and Dimyristoylphosphatidylcholine as well as Terpenes such as D - Limonene and Fenchone were procured from Kivi Lab, Por, India. All other reagents were purchased from local suppliers.

Method of Preparation of Amphotericin B Invasomes:

Drug and terpene or mixtures of terpenes are dissolved in ethanolic phospholipid solution. The mixture is vortexed for 5 min and then sonicated for 5 min in order to obtain a clear solution. Phosphate buffer saline (PBS) (pH: 7.4) is added to the solution by a syringe under constant vortexing. The vortexing is continued for an additional 5 min. The last step is the extrusion of multilamellar vesicles through polycarbonate membranes of different pore sizes. The invasome dispersions are extruded through each polycarbonate membrane for several times. [9 - 11]

Characterization of Amphotericin B Invasomes

Entrapment efficiency

Entrapment efficiency of invasomal formulation determined by centrifuging 10 mL invasomal suspension for 10 min. The clear fraction was further used for the determination of free

drug by using UV/visible spectrophotometer at 406 nm.¹³ The entrapment efficiency was calculated using the following formula:

$$\text{Entrapment efficiency} = (C_t - C_f) / C_t \times 100$$

Where, C_t : concentration of total drug and; C_f : concentration of unentrapped (free) drug

Vesicle size

It is determined by Malvern Instrument at Kivi Labs, Por, Vadodara.

Statistical analysis:

All data were subjected to analysis of variance (ANOVA) using Design Expert 13.0.01.

3. Results and Discussion

Calibration curve of Amphotericin B

A stock solution was prepared by dissolving 100mg of drug in small quantity of methanol & sonicated for few minutes & diluted with 100 ml of phosphate buffer (pH 7.4). Stock solution was serially diluted to get solutions in range of 2 - 10µg/ml & λ_{max} of solution was found out. Absorbance of different diluted solution was measured in UV - visible spectrophotometer scan. Calibration curve was plotted by taking concentration in X axis & absorbance in Y axis & correlation coefficient 'R²' was calculated.^[12]

Identification of Amphotericin B by FTIR Spectroscopy

Amphotericin B had identified by using FTIR spectroscopy in which it was confirmed from the observed functional groups stated in spectra.

Formulation of Amphotericin B Invasomes

Amphotericin B Invasomes was prepared by using Mechanical Dispersion method with QbD Approach. In order to assess risk, the identified QTPPs were %E. E and Vesicle size whereas CQAs were Conc. and Type of Phospholipids, Conc. and Type of terpene, ethanol conc., sonication speed and sonication time displayed in table 1.^[14 - 15]

Screening of significant risk factors using Plackett - Burman design^[16 - 17]

- Plackett - Burman design is an efficient screening method to identify the critical factors using as few experimental runs as possible.
- Design is used for screening of independent variables: significant (critical) or non - significant (non - critical).
- It is two - level design i. e. Low (- 1) level & High (+1) level.
- PB design gives 12 runs that may be used for an experiment containing up to 11 factors.
- The parameter level selection was based on preliminary studies and literature survey.
- In this experiment, as per the method of preparation of Invasomes, to screen critical material attributes (CQAs) and process parameters (CPPs) are conducted to run the screening design of 12 - run Plackett - Burman design utilizing Design Expert® software (Version 13.0.1) displayed in the table 2.

Characterization of Amphotericin B Invasomal Batches

Amphotericin B loaded Invasomes were prepared by using mechanical dispersion method which is characterized on the basis of %E. E & Vesicle size of each batch. Results were showed in the figure 3.

Statistical Discussion

ANOVA interpretation of Y1 Response (%E. E.)

The analysis of variance (ANOVA) revealed a statistical difference between the batches from the figure 4. A regression coefficient is said to be significant if p - value is less than 0.05. R² value was 0.9482 indicating a good fit. From the result, it is evident that phospholipid conc. (0.0073) and Terpene conc. (0.6506) significantly affect the entrapment efficiency which is again confirmed by Pareto chart as both the graphs are crossing the t - value limit which is 2.776. When the higher EE is desired within selected factor range, factor X1 and X2 have positive coefficients which indicate that increasing factor value increases the response which means that increasing phospholipid and terpene concentration increases the %EE of Invasomes.

ANOVA interpretation of Y2 Response (V. S)

The analysis of variance (ANOVA) revealed a statistical difference between the batches from the figure 5. From the result, it is evident that phospholipid concentration (0.0066), terpene conc. (0.0144) and sonication speed (0.0347) significantly affect the V. S which is again confirmed by Pareto chart. When the concentration of phospholipid and terpenes increases with rise in sonication speed, then desired vesicle size within selected range can be obtained.

Summary Critical Quality Attributes Analysis of various CMAs & CPPs based on PBD Screening Design Analysis & Pareto Chart. Based on the Characterization data and Pareto charts, following summary has been made which is depicted in table 4.

4. Conclusion

The study successfully identified key critical quality attributes CQAs influencing the formulation of Amphotericin B invasomes. Optimizing these factors, particularly phospholipid and terpene concentrations, as well as sonication speed, resulted in a formulation with enhanced entrapment efficiency and controlled vesicle size, demonstrating its potential for improved transdermal delivery of Amphotericin B.

Acknowledgements:

We would like to thank Kivi Labs Pvt. Ltd., Por, Vadodara for their kind support for Particle size assessment and other testing.

References

- [1] Jain S., "Quality by Design (QbD): a comprehensive understanding of implementation and challenges in pharmaceuticals development, " Int J Pharm Sci (6), pp.29 - 35, 2013.
- [2] Utz J., Louria D., Feder N., Emmons C., McCullough

- N., "A Report of Clinical Studies on the Use of Amphotericin in Patients with Systemic Fungal Diseases," CABI, Wallingford, UK, 1958.
- [3] Banshoya K., Kaneo Y., Tanaka T., Yamamoto S., Maeda H., "Development of an amphotericin B micellar formulation using cholesterol - conjugated styrene - maleic acid copolymer for enhancement of blood circulation and antifungal selectivity," *Int. J. Pharm.*, 589, 119813, 2020.
- [4] doi: 10.1016/j.ijpharm.2020.119813.
- [5] Baig, Reesha & Wais, Mohammad, "Formulation and Development of Proniosomal Gel for Topical Delivery of Amphotericin B," *International Journal of Pharmacy and Pharmaceutical Sciences* 4 (1), pp.37 - 49, 2022.
- [6] Ashtikar M., Langelüddecke L., Fahr A., Deckert V., "Tip - enhanced Raman scattering for tracking of invasomes in the stratum corneum," *Biochim. Et Biophys. Acta (Bba) - Gen.*, 1861, pp.2630–2639, 2017. doi: 10.1016/j.bbagen.2017.07.003.
- [7] Kumari et al, "Development of Soft Luliconazole Invasomes Gel for Effective Transdermal Delivery: Optimization to In - Vivo Antifungal Activity," *Gels* (9), pp.626 - 634, 2023.
- [8] Fouad A. et al Design, "optimization, and in vivo evaluation of invasome mediated candesartan for the control of diabetes-associated atherosclerosis," *Drug Delivery and Translational Research* (8), pp.1 - 17, 2023.
- [9] Albash, et al, "Development and Optimization of Terpene - Enriched Vesicles (Terpesomes) for Effective Ocular Delivery of Fenticonazole Nitrate: In vitro Characterization and in vivo Assessment," *International Journal of Nanomedicine*, 16, pp.609–621, 2021.
- [10] Q. Hoda, "Optimization of valencene containing lipid vesicles for boosting the transungual delivery of itraconazole," *Biotech*, 11 (3), pp.137 - 143, 2021.
- [11] Vidya and Lakshmi, "Cytotoxic effect of transdermal invasomal anastrozole gel on MCF - 7 breast cancer cell line," *Journal of Applied Pharmaceutical Science*, 9 (03), pp.050 - 058, 2019.
- [12] Dhimmar, B. et al, "Newfangled Topical Film - Forming Solution for Facilitated Antifungal Therapy: Design, Development, Characterization, and In Vitro Evaluation," *Polymers*, 15 (1003), pp.1 - 20, 2023.
- [13] Barbalata C. I. et al, "Application of the QbD Approach in the Development of a Liposomal Formulation with EGCG," *Journal of Pharmaceutical Innovation* (17), pp.867–880, 2022.
- [14] Patel R. N. and Upadhyay U, "formulation and development of luliconazole microsponges for topical delivery system using QbD approach," *International Journal of Creative Research Thoughts*, 1 (5), pp. M707 – M739, 2023.
- [15] Alnaim, A. S. et al, "Qbd - Based Approach to Optimize Niosomal Gel of Levosulpride for Transdermal Drug Delivery," *Gels* (9), pp.213 - 217, 2023.
- [16] Almotairi, N. et. al., "Design and Optimization of Lornoxicam Dispersible Tablets Using Quality by Design (QbD) Approach," *Pharmaceuticals*, 15 (1463), pp.1 - 18, 2022.
- [17] Mehta J. et al, "Formulation and Development of Luliconazole loaded Microemulgel using QBD Approach," *International Journal of Pharmaceutical Research and Applications*, 6 (3), pp.209 - 221, 2021.
- [18] Mohapatra, S. et al, "Quality by Design Assisted Optimization and Risk Assessment of Black Cohosh Loaded Ethosomal Gel for Menopause: Investigating Different Formulation and Process Variables," *Pharmaceutics*, 15 (465), pp.1 - 25, 2023.
- [19] Rapallia V. K. et al, "QbD - driven formulation development and evaluation of topical hydrogel containing ketoconazole loaded cubosomes." *Materials Science & Engineering*, 119 (11), pp.15 - 48, 2021.
- [20] Arora D. et al, "Quality by design driven development of resveratrol loaded ethosomal hydrogel for improved dermatological benefits via enhanced skin permeation and retention," *International Journal of Pharmaceutics*, 567: 118448, 2019.

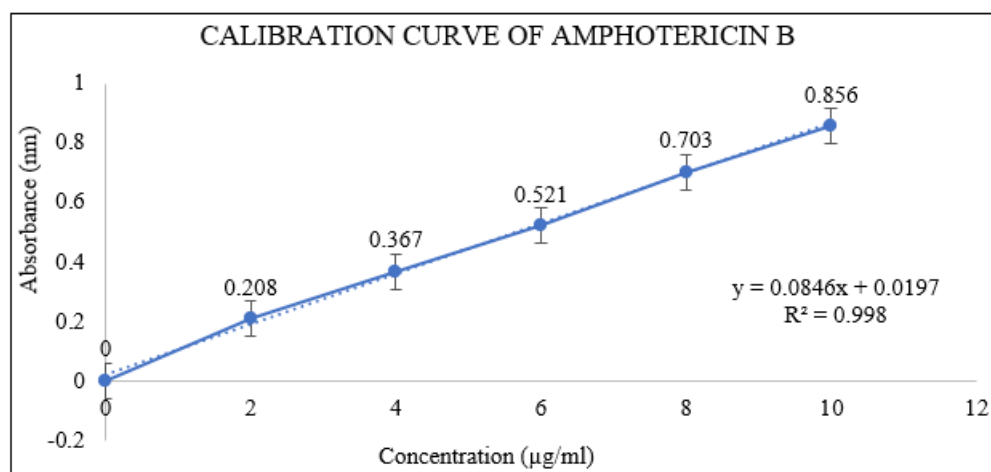


Figure 1: Calibration curve of Amphotericin B.

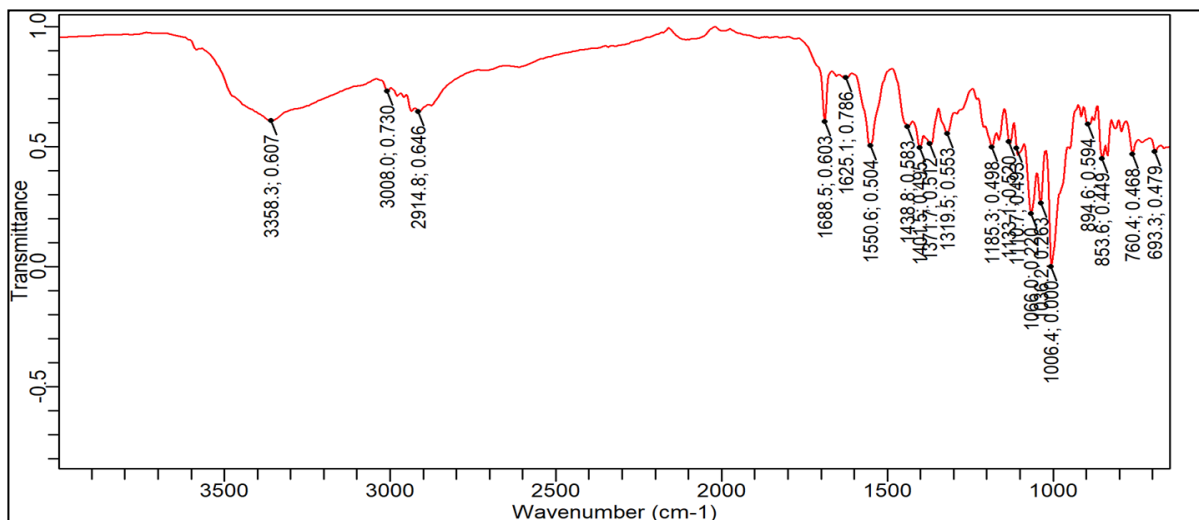


Figure 2: FTIR spectra of Amphotericin B.

Table 1: Selection of Independent Variables and reason for selection

Independent Variables (CQAs)	Range	Reason for Selection
CRITICAL MATERIAL ATTRIBUTES (CMAs)		
Concentration of Phospholipid	100 - 200 mg	<ul style="list-style-type: none"> Disintegrates on skin penetrate deep into the intercellular matrix and get mixed with the lipid bilayer of SC\ changes its structural integrity, thus making it easier for active moieties to permeate deep into the skin.
Type of Phospholipid	<ul style="list-style-type: none"> DMPC (Dimyristoylphosphatidylcholine), SPC (soya phosphatidylcholine) 	<ul style="list-style-type: none"> To form stable vesicles To prevent leakage To form solid, rigid & vesicular structure
Concentration of Terpene	5 - 10 mL	<ul style="list-style-type: none"> Permeability enhancement is their ability to disturb or break the hydrogen bonds between the components of cellular bilayer membrane.
Type of Terpene	Fenchone, D - Limonene	<ul style="list-style-type: none"> Terpenes with high lipophilic character act as strong boosters for sorption of lipophilic drug. Terpenes with non - polar hydrogen (e. g. limonene) are much more efficient boosters than oxygen - rich polarized terpenes.
Concentration of Ethanol	3 - 4 % v/v	<ul style="list-style-type: none"> It improves vesicular ability to penetrate the SC Provides net (- ve) surface charge and prevents aggregation of vesicle due to electrostatic repulsion. Ethanol interacts with lipid moiety in the polar head group region, resulting in a decrease in the transition temperature of the lipids thus improved their fluidity.
CRITICAL PROCESS PARAMETERS (CPPs)		
Sonication time	3 - 5 min	when increasing the sonication time <ul style="list-style-type: none"> PDI is decreased due to decreased the VS.
Sonication Speed	500 - 700 RPM	Particle size decreased when speed of sonication increased.

Table 2: Parameters feed in Design Expert to run Plackett Burman Design

Variable Code	Variables	UNITS	LEVELS	
			LOW (-1)	HIGH (+1)
Independent Variables: X - Inputs				
X1	Concentration of Phospholipid	mg	100	200
X2	Type of Phospholipid	Categoric	DMPC	SPC
X3	Concentration of Terpene	mL	5	10
X4	Type of Terpene	Categoric	Fenchone	D - Limonene
X5	Concentration of Ethanol	% v/v	3	4
X6	Sonication time	Min	3	5
X7	Sonication Speed	RPM	500	700
DEPENDENT VARIABLES: Y - RESPONSES				
Y1	Entrapment Efficiency		%	
Y2	Vesicle Size		nm	

Table 3: Compositions of Variables in decoded forms

RUN	PL. Conc. (mg)	PL Type	Terpene Conc. (ml)	Terpene Type	Eth Conc. (%v/v)	ST (Min)	SS (RPM)
AIN 1	200	SPC	10	fenchone	3	3	700
AIN 2	100	SPC	10	D - limonene	3	3	500
AIN 3	200	DMPC	10	D - limonene	3	5	700
AIN 4	100	SPC	5	D - limonene	4	3	700
AIN 5	100	DMPC	10	fenchone	4	5	500
AIN 6	100	SPC	10	fenchone	4	5	700
AIN 7	200	SPC	5	D - limonene	4	5	500
AIN 8	200	DMPC	10	D - limonene	4	3	500
AIN 9	200	DMPC	5	fenchone	4	3	700
AIN 10	100	DMPC	5	D - limonene	3	5	700
AIN 11	200	SPC	5	fenchone	3	5	500
AIN 12	100	DMPC	5	fenchone	3	3	500

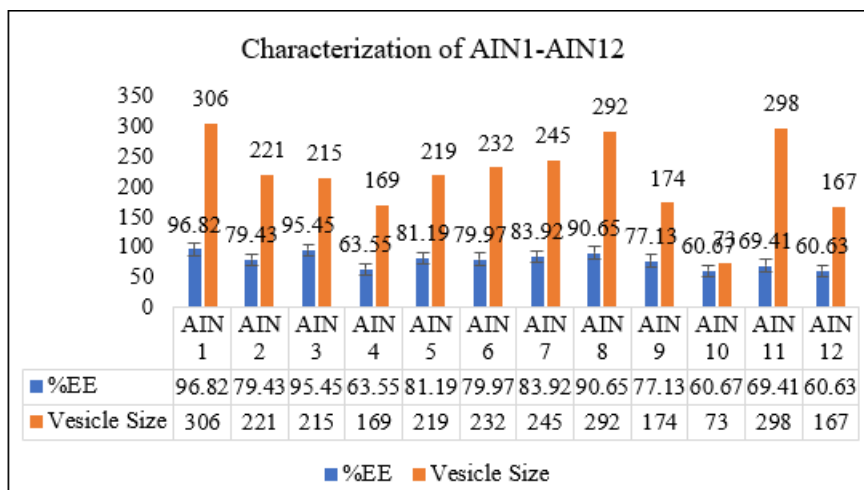


Figure 3: Characterization of AIN1 - AIN12

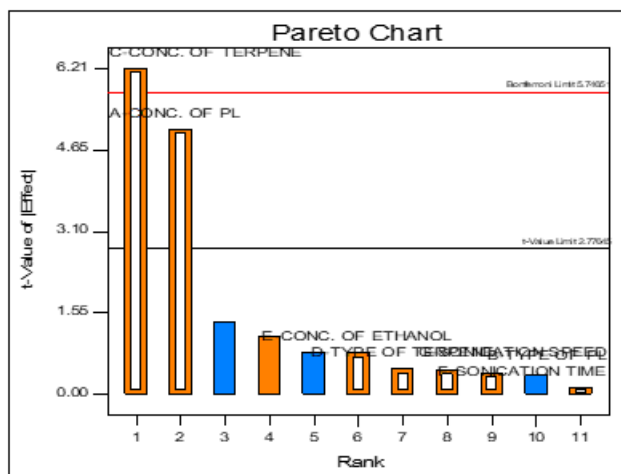


Figure 2: Pareto chart of Y1 Response - %E. E.

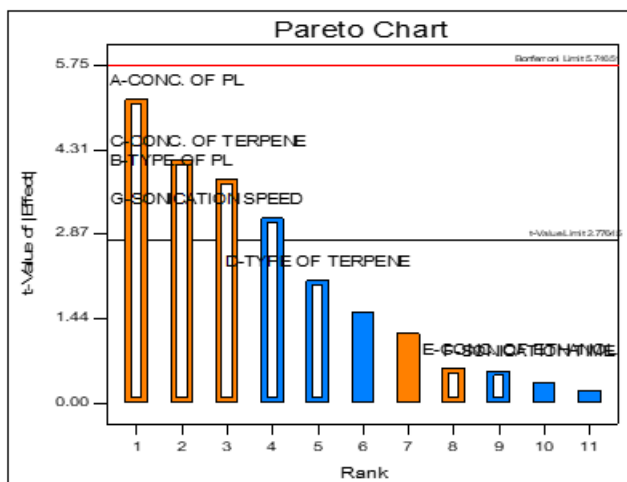


Figure 3: Pareto Chart of Y2 Response - Vesicular Size (nm)

Table 4: Critical Quality Attributes Analysis of various CMA & CPPs based on PBD Screening Design Analysis & Pareto Chart

Critical Quality Attributes Analysis of various CMA & CPPs	Effect on % E. E.	Reason	Effect on Vesicular Size	Reason
A - CONC. OF PL	++	Higher the concentration of phospholipid, more the drug incorporated into it.	++	Changes its structural integrity, thus making it easier for active moieties to permeate
B - TYPE OF PL	00	--	0+	Depends internal structure.
C - CONC. OF TERPENE	++	Higher the lipophilicity of the terpene, higher the solubilization of drug which increases space for drug incorporation.	++	Terpenes with high lipophilic character act as strong boosters for sorption of lipophilic drug
D - TYPE OF TERPENE	00	-	00	-
E - CONC. OF ETHANOL	+0	Ethanol interacts with lipid moiety in the polar head group region which results in decrease in transition temperature of lipids	00	Higher the concentration of ethanol, higher the fluidity in vesicles.
F - SONICATION TIME	00	-	+0	when increasing sonication time, PDI is decreased due to decreased the VS.
G - SONICATION SPEED	00	-	--	Particle size decreased when speed of sonication increased.

Note: ++: Positive Effect, +0: Moderate Effect, 00: No effect, --: Negative Effect

Tables and figure titles and legend:

Figure 1: Calibration curve of Amphotericin B.

Each reading has been performed thrice (n=3) to obtain standard deviation plot.

Figure 2: FTIR spectra of Amphotericin B.

Table 1: Selection of Independent Variables and reason for selection

Table 2: Parameters feed in Design Expert to run Plackett Burman Design

Parameters feed was gathered from the design expert software version 13.0.1

Table 3: Compositions of Variables in decoded forms

Parameters feed was gathered from the design expert software version 13.0.1

Figure 3: Characterization of AIN1 - AIN12

Each reading of %E. E has been performed thrice (n=3) to obtain standard deviation plot.

%E. Vesicle size

Figure 4: Pareto chart of Y1 Response - %E. E.

Figure 5: Pareto Chart of Y2 Response - Vesicular Size (nm)

Table 4: Critical Quality Attributes Analysis of various CMA & CPPs based on PBD Screening Design Analysis & Pareto Chart

++: Positive Effect, +0: Moderate Effect, 00: No effect, --: Negative Effect