Effect of Precursor β-phenylalanine on Production of Flavonoids of Maytenus emarginata in vitro

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Abstract: Medicinally important Maytenus emarginata is an ever green tree of desert useful against various diseases. Flavonoids have been reported in its plant parts. In present study unorganized tissue raised on MS medium supplemented with 1.5 mg/L BAP + 1 mg/L NAA, harvested at maximum GI, were analysed for qualitative and quantitative estimation of flavonoids by Subramanian and Nagarajan (1969) method. These tissues were fed with different concentration of dl β-phenylalanine (25, 50, 75, 100 mg/100ml) to observe the effect on GI and quantitative production of flavonoids. Maximum GI and amount of flavonoids (leutiolin, kaempferol, quercetin and total flavonoid) was estimated in eight weeks old tissue fed with 75 mg/100ml phenylalanine.

Keywords: Maytenus emarginata, β-phenylalanine, flavonoids, medicinal plant, GI

1. Introduction

Maytenus emarginata (Willd.) is an ever green tree that tolerates various types of stresses of the desert and is found in drier parts of central, south – western and north western India. Maytenus have been used for fever, asthma, rheumatism and gastrointestinal disorders, carcoma and leukemia, gastrointestinal troubles etc. Medicinal plants are rich source of secondary metabolites, biosynthetically derived from primary metabolites but restricted to specific taxonomic genera of plant kingdom and specific part of plant body. Secondary plant products are of major interest because of their biological activities ranging from antibacterial, antibiotic, insecticidal, hormonal, pharmacological and pharmaceutical.

β-phenylalanine amino acid is the precursor of flavonoids (Harborne, 1965 Barz, 1977). Biosynthetic scheme of flavonoids based mainly on C14 labelling experiments suggest that the C15 skeleton of the flavonoids is derived from phenylalanine through cinnamic acid, under the shikimik acid pathway. Effect of dl-β-phenylalanine on flavonoids has been studied in tissue cultures Tribulus terrestris (Sahuja, 1981), Agave wightii (Sharma, 1982), Cheiranthus cheiri (Agarwal, 1982), Tribulus alatus (Jit, 1985), Lycium barbarum (Shakawat, 1985), Zygophyllum simplex (Mathur, 1988), Seetzenia orientalis (Sethia, 1988), Abutilon pannosum, Ocimum americanum (Singh, 1988), Calligonum polygonoides, Lasius rudis (Bhojaj, 1991), Fagonia cretica (Kapoor, 1991), Lycium barbarum (Mukhi, 1995), Gossypium cultivars (Kaur, 1997, Goyal, 1997), Peganum harmala (Badia, 1999), Fragaria ananassa (Mori et al., 2001), Vigna aconitifolia (Tyagi, 2002), Withania somnifera (Bains, 2002), Zizyphus mauritiana (Chauhan, 2003), Spirodela intermedia (Gitz et al., 2004), Cassia angustifolia (Reddy, 2005), Balanites aegyptiaca (Bedawat, 2006), Ailanthus excelsa (Rao, 2007), Adhatoda vasica and Barleria prionitis (Deepa, 2009), Cocculus pendulus and Tinospora cordifolia (Yadav, 2010), Moringa oleifera (Talreja, 2010).

2. Material and Method

Murashige and Skoog’s medium (1962) supplemented with 1.5 mg/L BAP and 1.0 mg/L NAA was used for establishment of unorganized tissue of M. emarginata. Callus was maintained by frequent sub culturing at interval of 6 to 8 weeks at 26 ±1ºC, 60% relative humidity and diffused light conditions (3000 lux). Growth Indices (GI) of tissues were calculated at 2,4,6,8 and 10 weeks time intervals. Tissues were harvested at different age intervals and their growth indices (GI) were calculated.

Growth Index (GI)

Final fresh weight of tissue - Initial fresh weight of tissue

Initial fresh weight of tissue

These tissues established on MS medium (Standrized by 1.5 mg/L BAP and 1.0 mg/ L NAA) were supplemented with various concentrations of dl β-phenylalanine (25, 50, 75 and 100mg/100ml) to observe the effect on GI and quantitative amount of flavonoids. GI was calculated at the age of 2,4,6,8, 10 and 12 weeks separately for each concentration of phenylalanine. Tissues were harvested at maximum GI separately, dried, powdered and analysed for flavonoids by Subramanian and Nagarajan, (1969) method.

3. Analysis of flavonoids

Tissue samples of M. emarginata supplemented with various concentrations of phenylalanine were air dried, weighed, powdered, soxhlet extracted separately with 80% hot ethanol on a water bath for 24 hr to extract flavonoids and filtered. Filtrate was re-extracted with petroleum ether (Fr I), ethyl ether (Fr II) and ethyl acetate (Fr III) in succession following the method of Subramanian and Nagarajan (1969). Each step was repeated three times to ensure complete extraction in each case. Petroleum ether (Fr I) was rejected due to its richness in fatty substances where as Ethyl ether (Fr II) was analyzed for free flavonoids while the Ethyl acetate fraction was hydrolyzed with 7% H₂SO₄ for 2 hr. The mixture was filtered, the filtrate extracted with ethyl acetate was
neutralized with distilled water and then dried in vacuum and analyzed for bound flavonoids.

3.1 Qualitative and quantitative estimation

Plates were developed in an air tight chromatography chamber containing about 200 ml of solvent mixture of n-butanol, acetic acid and water 4:1:5 (upper layer). Developed plates were air dried and visualized under UV light 254 nm which showed one fluorescent spot in ethyl ether fraction second and two spots in ethyl acetate fraction third. The plates were also placed in a chamber saturated with ammonia vapours to observe the colours of the spots. The developed plates were also sprayed with 5% ethanolic ferric chloride solution for further confirmation. Each of the isolates was purified by preparative TLC in similar solvent system. Isolates were eluted with ethanol, crystallized by CHCl₃ and further confirmed by melting points, UV maxima on spectrophotometer and infra red spectral studies. Quantitative estimation of the identified flavonoids was carried out colorimetrically following method of Kariyon et al. 1953 and Naghshtet al. 1975 in case of quercetin and of Mabry et al. 1970 in case of luteolin and kaempferol.

4. Results and Discussion

Growth indices (GI) of unorganized tissues of *M. Emarginata* established and multiplied on standardized MS medium (MS + 1.5 mg/L BAP + 1.0 mg/L NAA) and supplemented with different concentrations of β-phenylalanine (25, 50, 75 and 100 mg/100ml) separately, were calculated.

GI of callus was found to be increased from standardized MS medium to STMS medium fed with 25mg PA/100ml to 50mg PA/100ml (3.43 and 4.15 respectively) and maximum (4.42) with 75 mg/100ml phenylalanine fed medium at all growth intervals (2, 4, 6, 8, 10, and 12 weeks). Maximum GI was observed at the age of eight weeks old tissues fed with all concentrations of phenylalanine. (Table.1)

<table>
<thead>
<tr>
<th>MEDIUM</th>
<th>2 WEE KS</th>
<th>4 WEE KS</th>
<th>6 WEE KS</th>
<th>8 WEE KS</th>
<th>10 WEE KS</th>
<th>12 WEE KS</th>
</tr>
</thead>
<tbody>
<tr>
<td>ST MS Medium</td>
<td>0.26± 0.24</td>
<td>0.49± 0.19</td>
<td>1.89± 0.20</td>
<td>3.24± 0.33</td>
<td>3.03± 0.17</td>
<td>2.11± 0.23</td>
</tr>
<tr>
<td>MS + 25 mg PA/100 ml</td>
<td>0.29± 0.21</td>
<td>0.54± 0.12</td>
<td>2.00± 0.11</td>
<td>3.43± 0.10</td>
<td>3.28± 0.11</td>
<td>2.03± 0.20</td>
</tr>
<tr>
<td>MS + 50 mg PA/100 ml</td>
<td>0.36± 0.18</td>
<td>0.73± 0.13</td>
<td>2.69± 0.15</td>
<td>4.15± 0.13</td>
<td>3.97± 0.18</td>
<td>3.03± 0.19</td>
</tr>
<tr>
<td>MS + 75 mg PA/100 ml</td>
<td>0.41± 0.15</td>
<td>0.92± 0.12</td>
<td>2.98± 0.13</td>
<td>4.42± 0.19</td>
<td>4.01± 0.20</td>
<td>3.51± 0.16</td>
</tr>
<tr>
<td>MS +100 mg PA/100 ml</td>
<td>0.32± 0.09</td>
<td>0.69± 0.12</td>
<td>2.53± 0.39</td>
<td>3.89± 0.17</td>
<td>3.72± 0.33</td>
<td>3.33± 0.16</td>
</tr>
</tbody>
</table>

Individual as well as total flavonoid content showed gradual increase in amount present in tissue grown on standardized medium (MS) to tissues fed with 25 mg PA/100ml medium, up to maximum in tissue fed with 75 mg PA/100ml. Maximum amount of leutiolin (0.70mg/100g.d.w.), kaempferol (0.68mg/100g.d.w.), quercetin (0.64mg/100g.d.w.) and total flavonoid (2.02 mg/100g.d.w.) was observed in the tissue grown on standardized MS medium fed with 75 mg PA/100ml. After that the amount started decreasing in tissues fed with 100 mg PA/100ml (0.60 mg/100 g.d.w, 0.54 mg/100 g.d.w, 0.50 mg/100 g.d.w and 1.64 mg/100 g.d.w. respectively) (Table 2).

<table>
<thead>
<tr>
<th>MEDIUM</th>
<th>Leuteolin</th>
<th>Kaempferol</th>
<th>Quercetin</th>
<th>Total Flavonoid</th>
</tr>
</thead>
<tbody>
<tr>
<td>ST MS Medium</td>
<td>0.60± 0.02</td>
<td>0.53± 0.03</td>
<td>0.48± 0.05</td>
<td>1.61± 0.06</td>
</tr>
<tr>
<td>MS + 25 mg PA/100 ml</td>
<td>0.62± 0.03</td>
<td>0.53± 0.04</td>
<td>0.52± 0.05</td>
<td>1.70± 0.04</td>
</tr>
<tr>
<td>MS + 50 mg PA/100 ml</td>
<td>0.65± 0.04</td>
<td>0.60± 0.03</td>
<td>0.57± 0.04</td>
<td>1.82± 0.03</td>
</tr>
<tr>
<td>MS + 75 mg PA/100 ml</td>
<td>0.70± 0.02</td>
<td>0.68± 0.03</td>
<td>0.64± 0.06</td>
<td>2.02± 0.05</td>
</tr>
<tr>
<td>MS +100 mg PA/100 ml</td>
<td>0.60± 0.05</td>
<td>0.54± 0.04</td>
<td>0.50± 0.03</td>
<td>1.64± 0.04</td>
</tr>
</tbody>
</table>
In all observations amount of leuclolin was slightly more than kaempferol and kaempferol than quercetin as in tissue grown on standardized MS medium as well as MS medium fed with kaempferol and kaempferol than quercetin as in tissue grown in all observations amount of leuclolin was slightly more than kaempferol and kaempferol than quercetin as in tissue grown on standardized MS medium as well as MS medium fed with

5. Conclusion

The present study shows that plants and their tissue cultures retain the potential to synthesize flavonoids in fair amount. Di-β-Phenylalanine being one of the precursors of flavonoids, when fed into the MS medium, increased the production of flavonoids appreciably, up to a certain limit. Hence, it can be concluded that specific concentration of phenylalanine can be fed into the medium, to obtain higher concentration of flavonoids, for large scale production being medicinally useful. All these findings strengthen the view of Barz (1977) that phenylalanine is a precursor of flavonoids and plays important role in flavonols biosynthesis.

References


Authors Profile

Shweta Mathur received her post graduation in Biotechnology from Allahabad Agriculture University in 2006. She did her Ph.D in Botany from Maharaja Ganga Singh University, (Bikaner). She worked as Senior Research Fellow in ICMR project for 2 years from 2007-2009.

Dr. Asha Goswami is Principal in Govt. College, Rajasthan. She received her post graduation in Botany, Ph.D in Botany and F.B.S degree. She has 35 years of teaching experience to U.G, P.G students. She had 20 M.Phil students and 10 Ph.D students under her guidance. Her papers are published in National and International Journals. Her research experience is of 20 years.