

PCR Amplification, Sequencing of 16S rRNA Genes with Universal Primers and Phylogenetic Analysis of *Pseudomonas aeruginosa*

K. Amutha¹, V. Kokila²

¹Associate Professor, Department of Biotechnology, Vels University, Chennai, Tamil Nadu, India

²Research Scholar, Department of Biotechnology, Vels University, Chennai, Tamil Nadu, India

Abstract: *Pseudomonas aeruginosa* is a gram negative bacterium isolated from soil. The isolate was cultured in a selective medium (*Pseudomonas* Agar base) at 37°C for 24 hours. For species identification, *Pseudomonas* like organisms creates lot of problems when identified with the help of morphological and biochemical characters. However, sequencing of 16S rRNA region is a suitable technique for species identification. The amplified product of 16S rRNA was submitted to NCBI database. Amplification of 16S rRNA gene region, and new sets of primer pairs were designed by NCBI database search tool. To study phylogenetic relationship between various strains of *Pseudomonas aeruginosa* have often been based on sequencing of 16S rRNA gene region. Distance tree was constructed to find out genetic similarity between the organisms. Hence gene sequencing of 16S rRNA region was a suitable technique to identify *Pseudomonas aeruginosa* at molecular level.

Keywords: *Pseudomonas aeruginosa*, Amplification, 16S rRNA, Phylogenetic relationship, NCBI, BLAST

1. Introduction

The genus *Pseudomonas* is a gram negative, rod shaped microorganisms [1] reported to have ecological, economic and health related importance's. Some *Pseudomonas* species are reported to be pathogenic for plants [2, 3], opportunistic pathogens of animals or humans [4, 5, 6] and some are used for bio control agent because it exhibit plant growth promoting and pathogen suppressing functions [7, 8].

Sequencing of 16S rRNA is a molecular technique for characterization of bacteria and tools involved is to analyse the phylogenetic relationship of an organism [9]. For species identification various molecular methods has been raised. According to Busse et al 1996 [10], the molecular biological methods such as nucleic acid analysis, protein pattern or fatty acid profiles has been proved for identification of bacteria rapidly. Identification of bacteria at species level, DNA sequences at genus-specific might be widely used. According to Barrey et al 1991 [11], Jensen et al 1993 [12], Gurtler & Stanisich 1996 [13] 16S-23S rRNA intergenic spacer of the ribosomal RNA operon (RRN) gene region is used for identification of strains and species.

The aim of this study was, sequencing of the 16S rRNA gene region of *Pseudomonas aeruginosa* isolated from soil. To design genus- specific new set of primer pairs from the NCBI database search tool and to study the phylogenetic relationship between the various strains of *Pseudomonas aeruginosa*.

2. Materials and Methods

Pseudomonas aeruginosa was isolated from the soil sample collected at Maduravoyal near chennai. It was cultured in the selective medium (*Pseudomonas* Agar Base) and incubated at 37°C for 24 hours. The isolate was identified based on the

morphological and biochemical characters [14]. For species identification sequencing of 16S rRNA gene region was carried out.

2.1 DNA Extraction

DNA was extracted from *Pseudomonas aeruginosa* based on the method described by Pitcher et al 2008 [15]. After extraction the DNA sample was run on 1% agarose gel at a constant voltage of 100V. The gel was examined on UV transilluminator.

2.2 PCR amplification and sequencing of 16S rRNA

16S rRNA gene region was amplified with the universal primers. For setting up PCR, the following reaction mixtures were added into the PCR tube. The reaction mixtures were 5µl of template, Primers: 1 µl of Forward primer- 27F (5' AGAGTTTGATCCTGGCTCAG 3'), 1 µl of Reverse primer- 1492R (5' TACCTTGTTACGACTT 3') [16], 6 µl of assay buffer, 2 µl of Taq DNA polymerase and 5 µl of dNTP mix (Applied Biosystems, Acme Progen Biotech (India) pvt. Ltd, Salem, Tamilnadu, India). The amplification was carried out in a thermal cycler for 40 cycles using the following reaction conditions, denaturation of DNA at 94°C for 1 minute, primer annealing at 56 °C for 30 seconds and primer extension at 72 °C for 1 minute. The amplified PCR product was mixed with 5 µl of gel loading buffer. 1.5% agarose gel was casted. The samples were loaded along with 5 µl of 1kb DNA ladder (HIMEDIA, Mumbai, Maharashtra, India) as a molecular marker. The gel was run and examined on UV transilluminator to visualize the bands. PCR products were purified by using the PCR Klenzol™ (Genei, Bangalore, India) and it was sequenced with an ABI Prism 3700 DNA Analyzer (Acme Progen Biotech (India) Pvt. Ltd., Salem, Tamilnadu, India).

2.3 Nucleotide sequence accession number and BLAST analysis

The nucleotide sequence 16S rRNA gene region data was submitted to NCBI nucleotide sequence database. Using BLAST tool, phylogentic tree, primer pairs were designed from NCBI database search tool.

3. Result

Pseudomonas aeruginosa was isolated from soil. Based on the morphological and biochemical characters it was identified as *Pseudomonas aeruginosa* (Table 1). *Pseudomonas aeruginosa* was gram negative, which shows positive result on catalase and oxidase test. Hence to identify and confirm the *Pseudomonas aeruginosa* at molecular level, 16S rRNA gene region was amplified and sequenced. Genomic DNA was extracted from *Pseudomonas aeruginosa* by the standard method. PCR amplification of 16S rRNA gene region by using universal primer, the obtained PCR product resulted in 1451 bp (Fig. 1). The sequence was submitted to NCBI database and the accession number is KC119195. By using BLAST analysis, 98 sequences of NCBI data gave 99% similarity. Phylogenetic tree generated by NCBI tool proves that this organism genetically related with other organisms (Fig. 2). NCBI primer blast tool help us to search new primer pairs to amplify 16S rRNA gene region (Table 2).

Table 1: Morphological And Biochemical Identification Of *Pseudomonas Aeruginosa*

S.No	Characteristics	
	Morphological Identification	
1.	Gram Staining	Gram negative, Rods
2.	Colony	Small, pigmented, circular
	Biochemical Identification	
1.	Catalase	+
2.	Oxidase	+
3.	Methyl Red	-
4.	Indole	+
5.	Citrate Test	+
6.	Nitrate Test	+

+Positive, - Negative

1 AGATTGAACG CTGGCGGCAG GCCTAACACA
 TGCAAGTCGA GCGGATGAAG
 51 GGAGCTTGCT CCTGGATTCA GCGGCGGACG
 GGTGAGTAAT GCCTAGGAAT
 101 CTGCCTGGTA GTGGGGGATA ACGTCCGGAA
 ACGGGCGCTA ATACCGCATA
 151 CGTCCTGAGG GAGAAAGTGG GGGATCTTCG
 GACCTCACGC TATCAGATGA
 201 GCCTAGGTCG GATTAGCTAG TTGGTGGGT
 AAAGGCCTAC CAAGGCGACG

251 ATCCGTA ACT GGTCTGAGAG GATGATCAGT
 CACACTGGAA CTGAGACACG
 301 GTCCAGACTC CTACGGGAGG CAGCAGTGGG
 GAATATTGGA CAATGGGCGA
 351 AAGCCTGATC CAGCCATGCC GCGTGTGTGA
 AGAAGGTCTT CGGATTGTAA
 401 AGCACTTTAA GTTGGGAGGA AGGGCAGTAA
 GTTAATACCT TGCTGTTTTG
 451 ACGTTACCAA CAGAATAAGC ACCGGCTAAC
 TTCGTGCCAG CAGCCGCGGT
 501 AATACGAAGG GTGCAAGCGT TAATCGGAAT
 TACTGGGCGT AAAGCGCGCG
 551 TAGGTGGTTC AGCAAGTTGG ATGTGAAATC
 CCCGGGCTCA ACCTGGGAAC
 601 TGCATCCAAA ACTACTGAGC TAGAGTACGG
 TAGAGGGTGG TGGAAATTTCC
 651 TGTGTAGCGG TGAAATGCGT AGATATAGGA
 AGGAACACCA GTGGCGAAGG
 701 CGACCACCTG GACTGACT GACACTGAGG
 TCGGAAAGCG TGGGAGACAA
 751 ACAGGATTAG ATACCCTGGT AGTCCACGCC
 GTAAACGATG TCGACTAGCC
 801 GTTGGGATCC TTGAGATCTT AGTGGCGCAG
 CTAACGCGAT AAGTCGACCG
 851 CCTGGGGAGT ACGGCCGCAA GGTTAAAAC
 CAAATGAATT GACGGGGGCC
 901 CGCACAAGCG GTGGAGCATG TGGTTTAATT
 CGAAGCAACG CGAAGAACCT
 951 TACCTGGCCT TGACATGCTG AGAACTTTCC
 AGAGATGGAT TGGTGCCTTC
 1001 GGGAACTCAG ACACAGGTGC TGCATGGCTG
 TCGTCAGCTC GTGTCGTGAG
 1051 ATGTTGGGTT AAGTCCCGTA ACGAGCGCAA
 CCCTTGTCCT TAGTTACCAG
 1101 CACCTCGGGT GGGCACTCTA AGGAGACTGC
 CGGTGACAAA CCGGAGGAAG
 1151 GTGGGGATGA CGTCAAGTCA TCATGGCCCT
 TACGGCCAGG GCTACACACG
 1201 TGCTACAATG GTCGGTACAA AGGGTTGCCA
 AGCCGCGAGG TGGAGCTAAT
 1251 CCCATAAAAC CGATCGTAGT CCGGATCGCA
 GTCTGCAACT CGACTGCGTG
 1301 AAGTCGGAAT CGTAGTAAT CGTGAATCAG
 AATGTCACCG TGAATACGTT
 1351 CCCGGGCCTT GTACACACCG CCCGTCACAC
 CATGGGGAGT GGGTTGCTCC
 1401 AGAAGTAGCT AGTCTAACCG CAAGGGGGGA
 CGGTTACCAC CGGAGTGATT
 1451 CATGACTGGG GGTGAAGTCG TAACAAGGTA
 GCCTAG
 //

Figure 1: Amplified 16S rRNA gene region

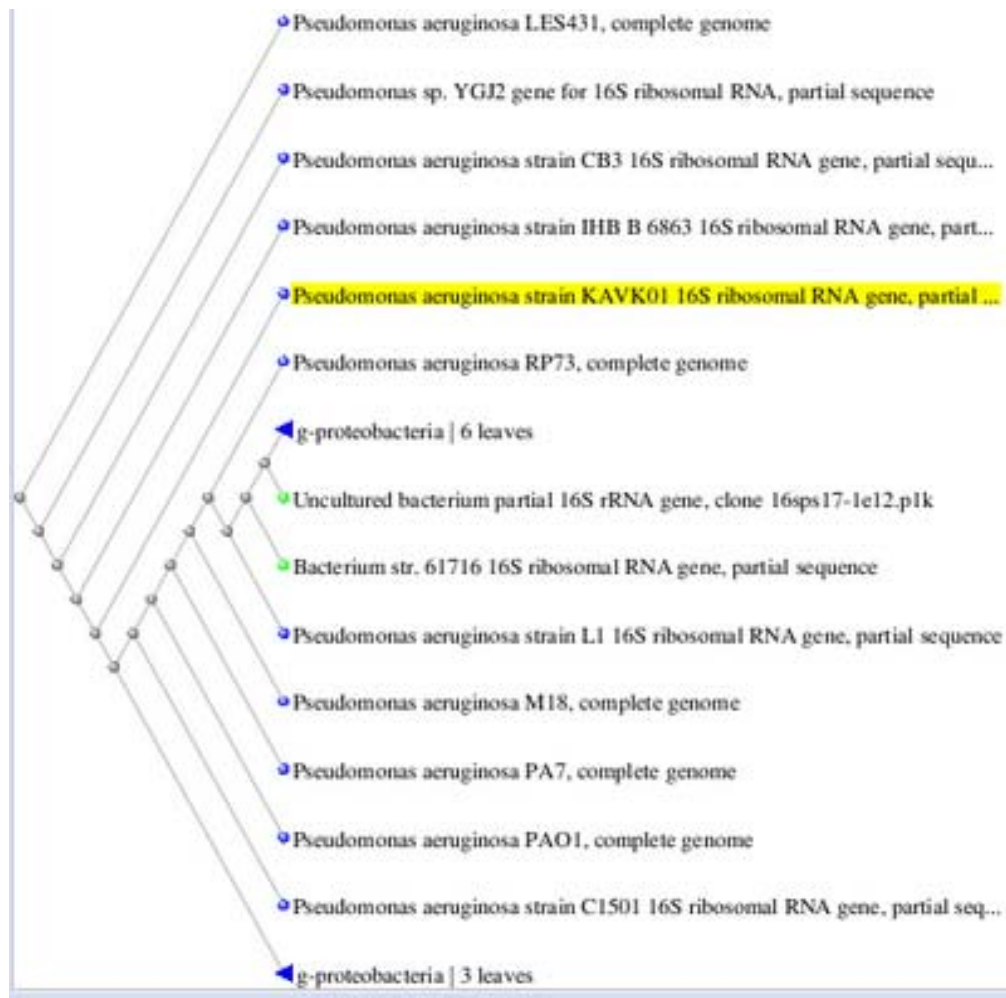


Figure 2: Distance Tree Using Blast Tool in the Ncbi Database Search Tool Of (Accession No: Kc119195) (SOURCE: <http://www.ncbi.nlm.nih.gov/blast/treeview/tree View>)

Table 2: Designing of Primer Pairs Using Primer Blast Tool In The Ncbi Database Search Tool Of (Accession No: Kc119195) (Source: <http://www.ncbi.nlm.nih.gov/tools/primer-blast/primertools>)

S.NO	Primer Pairs	Sequence (5'→3')	Template Strand	Length	Product Length
1.	Forward Primer	CGTCCTGAGGGAGAAAGTGG	Plus	20	111
	Reverse Primer	CAGTTACGGATCGTCGCCTT	Minus	20	
2.	Forward Primer	AGTTGGGAGGAAGGGCAGTA	Plus	20	237
	Reverse Primer	ATCCACCACCCTCTACCGT	Minus	20	
3.	Forward Primer	CAAGGCGACGATCCGTAAC	Plus	20	119
	Reverse Primer	ATCAGGCTTTCGCCATTGT	Minus	20	
4.	Forward Primer	TAAAGGCCTACCAAGGCGAC	Plus	20	414
	Reverse Primer	CCACCACCCTCTACCGTACT	Minus	20	
5.	Forward Primer	GTAGTGGGGATAACGTCCG	Plus	20	141
	Reverse Primer	TCGCCTTGGTAGGCCTTTAC	Minus	20	
6.	Forward Primer	GGGGTAAAGGCCTACCAAGG	Plus	20	135
	Reverse Primer	GATCAGGCTTTCGCCATTG	Minus	20	
7.	Forward Primer	GTACGGTAGAGGGTGGTGGGA	Plus	20	785
	Reverse Primer	GCTACTTCTGGAGCAACCCA	Minus	20	
8.	Forward Primer	CTGCCTGGTAGTGGGGATA	Plus	20	149
	Reverse Primer	GTCGCCTTGGTAGGCCTTTA	Minus	20	
9.	Forward Primer	ACAATGGGCGAAAGCCTGAT	Plus	20	137
	Reverse Primer	GCCGGTGCTTATTCTGTTGG	Minus	20	
10.	Forward Primer	TACGGTAGAGGGTGGTGGAA	Plus	20	781
	Reverse Primer	ACTTCTGGAGCAACCCACTC	Minus	20	

4. Discussion

In the past decade, the *Pseudomonas* classification had attracted more attention and it was reclassified by Brosch et al 1996, Kersters et al 1996, Palleroni 1992 [17, 18, 19]. Identification of *Pseudomonas* creates lot of difficulties [20, 21, and 4]. Morphologically similar species are alike to have biochemical characters. Sequence of highly conserved gene region 16S rRNA data helps us for the prediction of correct taxonomy. Our present study was carried out to sequence 16S rRNA based on PCR amplification for identification and genetic level conformation of *Pseudomonas aeruginosa*.

NCBI database provide us more information regarding nucleotide sequences. The revealed sequence accession number is KC119195 in NCBI provide us taxonomy report. Ninety eight organisms were evaluated with 99% similarity in BLAST analysis. In phylogenetic tree these two organism *Pseudomonas aeruginosa* strain IIIB B 6863 16S ribosomal RNA gene, partial sequence and *Pseudomonas aeruginosa* RP73, Complete genome were genetically very close to our strain of *Pseudomonas aeruginosa*. Primer pairs database search tool revealed that 10 new sets of primer pairs. These sets were very helpful for further sequencing of 16S rRNA species specific gene region.

5. Conclusion

The present study explained that some *Pseudomonas aeruginosa* strains are morphologically similar but different at genetic level. Hence amplification of 16S rRNA gene region is the suitable technique for identification of *Pseudomonas aeruginosa* is more accurate. By using nucleotide database search tool, analysing 16S rRNA gene sequence of *Pseudomonas aeruginosa* similarities with other strains, the phylogenetic relationship was constructed and new set of primer pairs were designed. Designing of new primers will help species level identification at molecular level in future.

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References

- [1] N.J. Palleroni, "Genus I. *Pseudomonas* (Migula 1894), p.141-199. In N.R. Krieg and J.G. Holt (ed.), *Bergey's manual of systematic bacteriology*, Vol.1, Williams & Wilkins, Baltimore, Md, 1984.
- [2] P. De Vos, M. Goor, M. Gillis and J. De Ley, "Ribosomal ribonucleic acid cistron similarities of phytopathogenic *Pseudomonas* species," *International Journal of Systematic Bacteriology*, 35: 169-184, 1985.
- [3] D. Stead, "Grouping of plant pathogenic and some other *Pseudomonas* spp. by using cellular fatty acid profiles," *International Journal of Systematic Bacteriology*, 42: 281-295, 1992.
- [4] P.H. Gilligan, "Microbiology of airway disease in patients with cystic fibrosis," *Clinical Microbiology Reviews*, 4:35-51, 1991.
- [5] N.J. Palleroni, "Human and animal pathogenic *Pseudomonads* p.3086-3103. In A. Balous, H.G. Truper, M. Dworkin, W. Harder and K.H. Schleifer (Ed), the prokaryotes. A handbook on the biology of bacteria: ecophysiology, isolation, identification, applications. Springer Verlag, New York, N.Y, 1992.
- [6] S.D. Tylaer, C.A. Strathdee, K.R. Rozee and W.M. Johnson, "Oligonucleotide primers designed to differentiate pathogenic *Pseudomonads* on the basis of the sequencing of genes coding for 16S-23S rRNA internal transcribed spacer," *Clinical Diagnostic Laboratory Immunology*, 2: 448-453, 1995.
- [7] C. Keel, D.M. Weller, A. Natsch, G. Defago, R.J. Cook, and L.S. Thomashow, "Conservation of the 2,4-diacetylphloroglucinol biosynthesis locus among fluorescent *Pseudomonas* strains from diverse geographic locations," *Applied and Environmental Microbiology*, 62:552-563, 1996.
- [8] D.J. O'Sullivan, and F. O'Gara," Traits of fluorescent *Pseudomonas* spp. involved in suppression of plant root pathogens," *Microbiology Reviews*, 56:662-676, 1992.
- [9] M.W. Gray, D. Sankoff, R.J. Cedergren, "On the evolutionary descent of organisms and organelles: a global phlogeny based on a highly conserved structural core in small subunit ribosomal RNA," *Nucleic Acids Research*, 12, 5837-5852, 1984.
- [10] H. J. Busse, E.B. M, Denner, & W. Lubitz, "Classification and identification of bacteria: Current approaches to an old problem. Overview of methods used in bacterial systematics." *Journal of Biotechnology*, 47:3-38, 1996.
- [11] T. Barry, G. Colleran, M. Glennon, L.K. Dunican, & F. Gannon,"The 16S-23S ribosomal spacer region as a target for DNA probes to identify eubacteria," *PCR Methods and Applications*, 1: 51-56, 1991.
- [12] M.A. Jensen, J.A. Webster, & N. Straus, "Rapid identification of bacteria on the basis of polymerase chain reaction amplified ribosomal DNA spacer polymorphism." *Applied Environment Microbiology*, 59: 945-952, 1993.
- [13] V. Gutler, & V.A Stanisich, "New approaches to typing and identification of bacteria using the 16S-23S rDNA spaces region, *Microbiology*, 142, 3-16, 1996.
- [14] Williams and Wilkins. 9th Edition. *Bergey's Manual of determinative bacteriology*, 1994.
- [15] D.G. Pitcher, N.A. Saunders and R.J. Owen, "Rapid extraction of bacterial genome DNA with guanidium thiocyanate," *Letters in applied Microbiology*, Vol 8, 4:151-156. 2008.
- [16] Jereny A. Frank, Claudia I. Reich, Shobha Sharma, Jon S. Weisbaum, Brenda A. Wilson and Gary J. Olsen, "Critical evaluation of two primers commonly used for amplification of bacterial 16S rRNA genes." *Applied and environmental microbiology*, 74(8): 2461-2470, 2008. Doi: 10.1128/AEM, 02272-07.
- [17] R. Brosch, M. Lefevre, F. Grimont and P.A.D.Grimont, " Taxonomic diversity of *Pseudomonads* revealed by computer-interpretation of ribotyping Data Systematic and Applied Microbiology, 19:541-555, 1996.
- [18] K. Kersters, W. Ludwig, M.Vancanneyt, P. De Vos, M.Gillis and K.H. Schleifer. "Recent changes in classification of *Pseudomonads*: an overview."

Systematic and Applied Microbiology, 19:465-477, 1996.

- [19] N.J. Palleroni, "Present situation of the taxonomy of aerobic Pseudomonads, p 105-115. In Galli, S. Silver and B. Witholt (ed.) Pseudomonas: Molecular biology and biotechnology. American Society for Microbiology, Washington, D.C, 1992.
- [20] J.L. Burns, J. Emerson, J.R. Stapp, D.L. Yim, J. Krzewinski, L. Lowden, B.W. Ransey, and C.R. Clausen. "Microbiology of sputum from patients at cystic fibrosis centres in the united states." Clinical Infectious Disease, 27:158-163, 1998.
- [21] A. Ferroni, I. Sermet-Gaudelus, E. Abachin, G. Querne, G. Lenoir, P. Berche, and J.L. Gaillard. Use of 16S rRNA gene sequencing for identification of nonfermenting gram-negative bacilli recovered from patients attending a single cystic fibrosis centre. Journal of Clinical Microbiology, 40: 3793-3797, 2002.

Author Profile



Dr. K. Amutha working as Associate professor in Department of Biotechnology, Vels University, Chennai, Tamil Nadu, India.



V. Kokila is a Ph.D Scholar in the Department of Biotechnology, Vels University, Chennai, Tamil Nadu, India.