

Enzymatic Synthesis of Micronutrients Rich Diacylglycerol from Rice Bran Oil Fatty Acid Distillate

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Abstract: Diacylglycerol (DAG) rich products with micronutrients like sterols, tocopherols and squalene is prepared from cheap raw material like rice bran oil fatty acid distillate (RBOFAD). RBOFAD is an important byproduct of vegetable oil refining industries in the physical refining process. DAG containing significant amount of unsaponifiable matters like sterols, tocopherols and hydrocarbons (mainly squalene) may certainly be considered as novel functional food ingredients. In the present study, free fatty acids present in the RBOFAD were esterified with glycerol of varying amount (FAD: Glycerol:: 2:1-1.5) for 8 h using non-specific enzyme NS 40013 (*Candida antarctica*). After bio esterification, the product mixture was purified by molecular distillation to remove FFA and undesirable components. The final DAG (72.86%) rich products containing sterol, tocopherol and squalene can be utilized in various functional food formulations. This is an extension of earlier work where present author made neutral glycerides from RBOFAD containing 53.8% DAG only.

Keywords: Fatty acid distillate, rice bran oil, sterols, tocopherols, squalene, *Candida antarctica*.

1. Introduction

Rice bran oil fatty acid distillate (RBOFAD) is a major by product of RBO refining industries which is mainly utilized in the soap manufacturing process. RBOFAD contains higher amount of unsaponifiable matters amongst which sterols, tocopherols and hydrocarbons (mainly squalene) are the main components. RBOFAD can be converted to diacylglycerol (DAG) rich products along with these unsaponifiable matters.

DAG oil was designated as Generally Recognized as Safe (GRAS) by an outside panel of scientific experts and their conclusion has been reviewed and accepted by the US Food and Drug Administration (FDA). The use of DAG oil in food products in the U.S. is limited to cooking oil and as an ingredient in salad dressing and mayonnaise. This GRAS determination is for use in vegetable oil spreads and home cooking oil. In Japan, the Ministry of Health, Labour and Welfare has approved DAG oil to manage serum triglycerides after a meal, which leads to less build-up of body fat.

DAG occurs naturally as a minor component of various edible fats and oils and has been used in foods as an emulsifier. DAG has been utilized as a cocoa butter anti blooming agent [1] and as an intermediate in the synthesis of structured lipid [2,3,4]. Studies on the nutritional properties and dietary effects of DAG [5,6,7] have revealed that DAG, of which 1,3-DAG is a major component, in contrast to TAG, had the ability to reduce serum TAG concentrations and as a result, to decrease both body weight and visceral fat mass in rats and humans [8,9]. Hidekatsu Yanai et. al. [10] has shown the therapeutic application of diacylglycerol oil for obesity and obesity-related metabolic disorder. Studies also indicate that DAG oil has numerous health benefits including reducing post-meal blood triglyceride levels. Clinical studies [9,11,12] in Japan have also shown that DAG oil may

increase overall metabolism, helping reduce the amount of fat already stored in the body [13,14,15].

Tocopherols and tocotrienols present in RBO can improve the blood circulation and reduce the oxygen demand of human body. α -tocopherol is used in pharmaceutical and cosmetic industries and a mixture of α -, γ - and δ - tocopherols is added to various foods including fats and oils. Squalene has the ability to assist the skin in retaining moisture. It helps to keep our skin soft and healthy and its antioxidant capabilities help to protect from the harsh effect of the environment. Squalene is being investigated as an adjunctive therapy in some cancers [16, 17] effective in inhibiting lung tumors and also demonstrated chemo protective effects against colon cancer in animal models [18]. So DAG rich oil along with these micronutrients has the immense potential health benefits to human beings.

By considering the above advantages and positive effects of DAG rich micronutrient oils and the specific importance of DAG rich products in the context of obesity and cholesterol reduction, attempts have been made to produce products rich in DAG by varying glycerol concentration, time course of reaction in the presence of enzyme. Zhong *et al.* [19] prepared solvent free 1,3-diacylglycerol by direct esterification of glycerol with saturated fatty acids in the presence of enzyme. Martinez *et al.* [20] prepared mono- and diacylglycerol enzymatically with conjugated linoleic acid in solvent hexane. In another study, Song *et al.* [21] prepared DAG enriched oil from high acid rice bran oil in the presence of Lipozyme RM-IM in a solvent free system. Santisawadi *et al.* [22] optimized a bioprocess for the preparation of DAG from palm fatty acid distillate using response surface methodology and yielded 70% DAG. In another work, Lo *et al.* [23] synthesized 70% DAG using *Rhizomucor miehei* lipase and analyzed the composition of DAG from corn oil deodorizer distillate. Earlier, present author studied the preparation of neutral glycerides form RBOFAD where

the final product contained 53.8% DAG along with TAG, MAG and other micronutrients [24]. But no study has been made for the preparation of higher amount of DAG in the oil with micronutrients like tocopherols, sterols and squalene from cheap raw materials. So, in the present investigation, an extended work has been made from our earlier work to produce DAG rich product (72.86%) from RBOFAD containing sterols, tocopherols and squalene using lipase catalyzed reaction in a solvent free system.

2. Materials and Methods

2.1 Materials

RBOFAD was collected from Sethia Oil Mill, Burdwan, West Bengal, India. Glycerol (A.R.) was purchased from E. Merck (India) Pvt. Limited. The lipase NS 40013 (*Candida antarctica*, a non-specific immobilized lipase) was a kind gift of Novozymes, South Asia Pvt. Ltd., Bangalore, India. Except otherwise specified all other chemicals used were A.R. Grade.

2.2 Methods

2.2.1 Bleaching of RBOFAD

About 500 gm of RBOFAD was taken in a 1 L round bottom flask and heated under vacuum (2-4 mm of Hg pressure) in a boiling water bath for 15 min. After that 5% Tonsil earth (Sud Chemical Company, Germany) and 0.6% activated charcoal (E. Merck, India) were added and shaken vigorously for 20 min under vacuum. Then it was cooled to 45°C and filtered under vacuum. The bleached FAD was stored in a refrigerator for further study.

2.2.2 Enzymatic esterification of bleached RBOFAD

Bleached RBOFAD and glycerol were taken in different molar ratios in a round bottom flask and stirred by a magnetic stirrer under vacuum of 4 mm Hg for 8 h using 5% (by weight of substrates) lipase catalyst (NS 40013). The temperature of the reaction was maintained at 65±2°C. The esterification reaction was monitored by estimating the free fatty acids in the reaction mixture periodically withdrawn. After 8 h of reaction, the product mixture was filtered for removing enzyme and kept for purification.

2.2.3 Purification of esterified products

The esterified product was purified in a molecular distillation unit (Model MS-300, SIBATA Scientific Co. Ltd., Japan). It was a falling film type apparatus and was provided with a rotating wiper that continuously rubbed the falling film on the evaporating surface. The temperature of the reaction product was maintained at 145±2°C and 12 pascal pressure to remove the residual fatty acids and volatile impurities. The amount of DAG in the final products was determined by standard column chromatographic method.

2.2.4 Determination of fatty acid composition by gas liquid chromatographic (GLC) method

Fatty acid composition was determined by gas liquid chromatographic method after converting the fatty acids into methyl esters. The HP-5890A GLC was connected with an

HP-3390A data integrator. The GLC was fitted with a glass column (1.83 m X 3.173 mm i.d.) packed with 10% DEGS supported on Chromosorb-WHP (100/200 mesh) of HP make. The oven temperature was programmed from 100°C to 190°C at 5° / min. The injector and the detector block temperature were maintained at 230°C and 240°C respectively. IOLAR-2 nitrogen was used as the carrier gas with flow rate 30 ml/min. The fatty acid esters peak were identified and calibrated with standard methyl esters. Data were represented an average of three determinations.

2.2.5 Determination of tocopherols by colorimetric method (Emerie-Engel method)

Total tocopherol content was measured according to the standard IUPAC method of Emerie – Engel [25].

2.2.6 Determination of sterols and squalene by HPLC method

The HPLC instrument (Waters, USA) was provided with Binary HPLC pump 1525 and Waters Dual Absorbance UV detector 2487 and Refractive Index detector 2414. The column (4.6 X 155 mm) used was Novapak bonded C18 having micro particulate silica of particle size of about 5 µm. The isocratic flow rate was 0.5ml/min. The whole system was supported by Breeze 2000 software. For the determination of sterols approximately 1.0 mg of unsaponifiable matter was dissolved in HPLC grade hexane and was filtered through a Millipore filter. 10 µl of the solution was injected and the materials were detected according to the retention time and quantified with reference to the standard sample. The mobile phase used was consisted of HPLC grade hexane, acetonitrile and isopropyl alcohol in the ratio of 75:15:10 v/v. The UV detector was used at 210 nm for squalene and 230 nm for sterols.

3. Results and discussion

Table 1 shows the compositions of fatty acids, unsaponifiable matters and neutral glycerides present in RBOFAD. It contains 75.19% free fatty acids and 9.7% neutral glycerides of which 5.2% TAG, 3.1% DAG and 1.4% MAG. RBOFAD contains higher amount (14.23%) of unsaponifiable matters of which sterols, tocopherols and squalene are present at 13.3, 37.4 and 49.3% respectively. Before enzymatic esterification process, RBOFAD was thoroughly bleached to remove peroxides. Bleached RBOFAD was esterified with glycerol in different proportions using non-specific NS 40013 lipase. Here three sets of experiments namely P-I, P-II and P-III were done to study the enzymatic esterification reaction. FAD and glycerol concentration were maintained at 2:1 and 2:1.25 and 2:1.50 for P-I, P-II and P-III respectively.

Table 1: Analytical characteristics of RBOFAD

Characteristics	Amount (% w/w)
Free fatty acid	75.19±2.03
i) Palmitic acid	28.47±0.44
ii) Stearic acid	2.05±0.003
iii) Oleic acid	37.24±1.24
iv) Linoleic acid	33.2±0.78
Neutral glycerides	9.7±0.4
i) Monoacylglycerol	1.4±0.007

ii) Diacylglycerol	3.1±0.01
iii) Triacylglycerol	5.2±0.009
Unsaponifiable matters	14.23±0.16
i) Sterols	13.3 ±0.33
ii) Tocopherols	37.4±0.74
iii) Squalene	49.3± 0.77

Values are represented as mean ± S.D. n=3

Figure 1 demonstrates that the rate of enzymatic esterification reaction in each case for 8 h and it shows that increasing concentration of glycerol decreases the concentration of FFA during the course of reaction. After 8 h of reaction, the product mixture contains 5.4, 5.1 and 4.8% FFA in products I, II and III respectively. The esterified products were then purified in molecular distillation apparatus at 145±2°C and 12 Pascal pressure to remove the residual fatty acids and volatile impurities. The composition of the molecular distilled esterified products is shown in Table 2.

From Table 2 it can be seen that the products, P-I, P-II and P-III contained 25.59, 7.27 and 11.11% TAG, 47.54, 72.86 and 61.16% DAG and 13.24, 6.99 and 15.36% MAG respectively. Also P-I, P-II and P-III contained 13.63, 12.88 and 12.37% unsaponifiable matters respectively. Increasing amount of TAG and MAG are formed in the bio esterification reaction mainly due to the random nature of the lipase. It also reveals from Table 2 that among the three products, P-II contained maximum amount of DAG (72.86%) than the other products. So it can be said that the ratio of RBOFAD and glycerol concentration 2:1.25 is the ideal ratio for getting the higher conversion DAG and the product P-II can be considered as DAG rich product with a good amount of unsaponifiable matters.

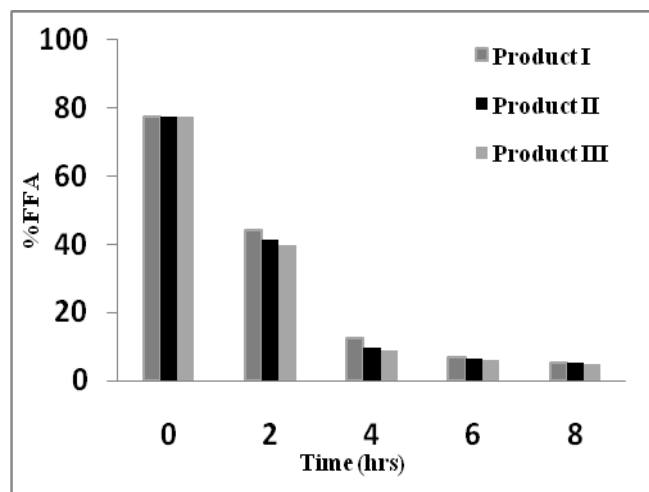


Figure 1: Enzymatic esterification of RBOFAD with glycerol in different proportions.

Table 2: Composition of DAG rich products (%w/w)

Product	TAG	DAG	MAG	Unsap. matters
P-I	25.59±0.37	47.54±0.34	13.24±0.13	13.63±0.24
P-II	7.27±0.04	72.86±0.58	6.99±0.05	12.88±0.22
P-III	11.11±0.11	61.16±0.21	15.36±0.09	12.37±0.13

Values are represented as mean ± S.D. n=3

Experimental studies also shows that increasing concentration of glycerol enhances the rate of formation of DAG but higher amount of glycerol further enhances the formation of TAG and MAG than DAG. From Table 2, we can see that when the ratio of RBOFAD to glycerol changes from 2:1.25 to 2:1.50 (enhancement of glycerol) in P-III, conversion of TAG increases from 7.27 to 11.11% and conversion of MAG increases from 13.24 to 15.36% but conversion of DAG decreases from 72.86 to 61.16%. Thus enhancement of concentration of glycerol is not permitted in this experimental study for the higher conversion of DAG. So, DAG rich product P-II with sufficient amount of unsaponifiable matters can be regarded a product of commercial importance.

P-I, P-II and P-III contain reasonable amounts of unsaponifiable matters which consisted tocopherols, sterols and squalene. Table 3 shows the composition of the unsaponifiable matters of DAG rich products where the desirable product P-II contains 29.31% tocopherols, 13.86% sterols and 51.93% squalene which is sufficiently quite good. Other products P-I and P-III contain nearly same amount of tocopherols, sterols and squalene. So P-II along with important micronutrients may be useful for functional food applications.

The qualities of DAG rich products are evaluated on the basis of acid value, peroxide value, anisidine value and colour. Table 4 shows the features of the DAG rich products P-I, P-II and P-III. It has been observed from Table 4 that all the products namely P-I, P-II and P-III confront good qualities with regard to above parameters. So considering maximum conversion of DAG from RBOFAD with micronutrients and on the basis of quality parameters of finished product, product P-II can be considered for further use as structured lipid or functional food ingredients.

Table 3 Composition of unsaponifiable matters of DAG rich products (%w/w)

Product	Tocopherols (Total)	Sterols (Total)	Squalene	Others
P-I	30.74± 0.33	13.11±0.16	52.46±0.37	3.69±0.12
P-II	29.31±0.32	13.86±0.17	51.93±0.31	4.9±0.14
P-III	28.78±0.17	14.63± 0.09	52.49±0.29	4.1±0.13

Values are represented as mean ± S.D. n=3

Table 4 Quality parameters of DAG rich products

Product	Acid value	Peroxide value(meq/kg)	Anisidine value	Colour (Lovibond 1" cell)
P-I	<0.1	<1	0.4±0.01	2.2Y+1.2R
P-II	<0.1	<1	0.6±0.01	1.9Y+0.9R
P-III	<0.1	<1	0.3±0.01	1.8Y+1.3R

Values are represented as mean ± S.D. n=3

4. Conclusion

DAG rich product can be produced from the relatively inferior grade raw material RBOFAD with the help of microbial lipase technology. The product contains considerable amount of micronutrients like sterols, tocopherols and squalene. The product is useful for

functional food applications, specialty products and has commercial importance. Further experimental study is needed for the product with much higher DAG along with the micronutrients.

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References

- [1] Z. Meng, W. X. Geng, J. W. Li, Z. Q. Yang, J. Jiang, X. G. Wang and Y. F. Liu, "Enzymatically Catalyzed Synthesis of Anti-blooming Agent 1,3-Dibehenoyl-2-oleoyl Glycerol in a Solvent-Free System: Optimization by Response Surface Methodology", *Journal of agricultural and Food Chemistry*, 61 (45), pp. 10798–10806, 2013.
- [2] Y. Cao, S. Qi, Y. Zhang, X. Wang, B. Yang and Y. Wang, "Synthesis of Structured Lipids by Lipase-Catalyzed Esterification of Triacetin with Camellia Oil Methyl Esters and Preliminary Evaluation of their Plasma Lipid-Lowering Effect in Mice.", *Molecules*, 18, pp. 3733-3744, 2013.
- [3] J. H. Kim, K. T. Lee, M. K. Lee, S. M. Jeon and M. S. Choi, "Diacylglycerol-Enriched Structured Lipids Containing CLA and Capric Acid Alter Body Fat Mass and Lipid Metabolism in Rats", *Ann. Nutr. Metab.*, 50, pp. 219–228, 2006
- [4] R. Rosu, M. Yasui, Y. Iwasaki, T. Yamane, "Enzymatic synthesis of symmetrical 1,3-diacylglycerols by direct esterification of glycerol in solvent-free system", *JAACS*, 76 (7), pp. 839-843, 1999.
- [5] N. Ota, S. Soga, T. Hase, I. Tokimitsu and T. Murase, "Dietary Diacylglycerol Induces the Regression of Atherosclerosis in Rabbits", *The Journal of Nutrition*, 137, pp. 1194–1199, 2007.
- [6] T. Nagao, H. Watanabe, N. Goto, K. Onizawa, H. Taguchi, N. Matsuo, T. Yasukawa, R. Tsushima, H. Shimasaki and H. Itakura, "Dietary diacylglycerol suppresses accumulation of body fat compared to triacylglycerol in men in a double blind controlled trial", *J. Nutr.*, 130, pp. 792-797, 2000.
- [7] X. H. Meng, D. Y. Zou, Z. P. Shi, Z. Y. Duan and Z. G. Mao, "Dietary diacylglycerol prevents high fat diet-induced lipid accumulation in rat liver and abdominal adipose tissue" *Lipids*, 39, pp. 37-41, 2004.
- [8] M. Murata, T. Ide and K. Hara, "Reciprocal response to dietary diacylglycerol of hepatic enzymes of fatty acid synthesis and oxidation in the rat", *Br. J. Nutr.*, 77, pp. 107-121, 1997.
- [9] K. Yamamoto, H. Asakawa, K. Tokunaga, H. Watanabe, N. Matsuo, I. Tokimitsu and N. Yagi, "Long-term ingestion of dietary diacylglycerol lowers serum triacylglycerol in type II diabetic patients with hypertriglyceridemia", *J. Nutr.*, 131, pp. 3204-3207, 2001.
- [10] H. Yanai, H. Yoshida, Y. Hirowatari and N. Tada, "Therapeutic Application of Diacylglycerol Oil for Obesity: Serotonin Hypothesis" *Functional Foods in Health and Disease*, 2(1), pp.1-10, 2012.
- [11] H. Taguchi, H. Watanabe, K. Onizawa, T. Nagao, N. Gotoh, T. Yasukawa, R. Tsushima, H. Shimasaki and H. Itakura, "Double-blind controlled study on the effects of dietary diacylglycerol on post-prandial serum and chylomicron triacylglycerol responses in healthy humans", *J. Am. Coll. Nutr.*, 19 (6), pp. 789-796, 2000.
- [12] N. Tada, H. Watanabe, N. Matsuo, I. Tokimitsu and M. Okazaki, "Dynamics of post-prandial remnant-like lipoprotein particles in serum after loading of diacylglycerols", *Clin. Chim. Acta.*, 311 (2), pp. 109-117, 2001.
- [13] S. Yasuhito and N. Saito, "Diacylglycerol kinase as a possible therapeutic target for neuronal diseases", *Journal of Biomedical Science*, 21(28), pp. 1-8, 2014.
- [14] K. C. Maki, M. H. Davidson, R. Tsushima, N. Matsuo, I. Tokimitsu, D. M. Umporowicz, M. R. Dicklin, G. S. Foster, K. A. Ingram, B. D. Anderson, S. D. Frost, and M. Bell, "Consumption of diacylglycerol oil as part of a reduced-energy diet enhances loss of body weight and fat compared with a triacylglycerol control oil", *Am. J. Clin. Nutr.*, 76, pp. 1230-1236, 2002.
- [15] T. Murase, M. Aoki, T. Wakisaka, T. Hase and I. Tokimitsu, "Anti-obesity effect of dietary diacylglycerol in C57BL/6J mice: dietary diacylglycerol stimulates intestinal lipid metabolism", *J. Lipid Res.*, 43, pp. 1312-1319, 2002.
- [16] A. L. Ronco and E. D. Stéfani, "Squalene: a multi-task link in the crossroads of cancer and aging", *Functional Foods in Health and Disease*, 3(12), pp. 462-476, 2013.
- [17] P. S. Granados, J. L. Quiles, C. L. Ramirez-Tortosa, J. Ochoa-Herrera, P. Pérez-López, M. Pulido-Moran and M. C. Ramírez-Tortosa, "Squalene ameliorates atherosclerotic lesions through the reduction of CD36 scavenger receptor expression in macrophages", *Mol Nutr Food Res*, 56(5), pp. 733-740, 2012.
- [18] C. V. Rao, H. L. Newmark and B. S. Reddy, "Chemopreventive effect of squalene on colon cancer", *Carcinogenesis*, 19, pp. 287-290, 1998.
- [19] N. Zhong, Z. Gui, L. Xu, J. Huang, K. Hu, Y. Gao, X. Zhang, Z. Xu, J. Su, and B. Li, "Solvent-free enzymatic synthesis of 1, 3-Diacylglycerols by direct esterification of glycerol with saturated fatty acids" *Lipids in Health and Disease*, 12 (65), pp. 1-7, 2013.
- [20] C. E. Martinez, C. E., J. C. Vinay, R. Brieva, C. G. Hill (Jr.) and H. S. Garcia, "Preparation of Mono- and Diacylglycerols by enzymatic esterification of glycerol with conjugated linoleic acid in hexane" *Applied Biochemistry and Biotechnology*, 125(1), pp. 63-75, 2005.
- [21] Z. Song, Y. Liu, Q. Jin, L. Li, X. Wang, J. Huang, and R. Liu, "Lipase-catalysed preparation of diacylglycerol-enriched oil from high-acid rice bran oil in solvent free system" *Applied Biochemistry and Biotechnology*, 168 (2), pp. 364-74, 2012.
- [22] S. Santisawadi, S. Chaiseri, N. Jinda and U. Klinkesorn, "Process optimization of using response surface design for Diacylglycerol synthesis from palm fatty acid distillate by enzymatic esterification", *Songklanakarini J. Sci. Technol.*, 35 (1), pp. 23-32, 2013.

- [23] S. K. Lo, B. S. Baharin, C. P. Tan and O. M. Lai, "Enzyme catalyzed production and chemical composition of Diacylglycerol from corn oil deodorizer distillate", 18 (3), pp. 265-278, 2005.
- [24] S. Nandi, S. Gangopadhyay and S. Ghosh, "Lipase catalyzed synthesis neutral glycerides rich in micronutrients form rice bran oil fatty acid distillate", Journal of Oleo Science, 57(11), pp. 599-603, 2008.
- [25] C. Paquot and A. Hautfenne, Standard methods for the analysis of oils, fats and derivatives. 7th revised and enlarged edition, Blackwell scientific publications, pp. 143-148 (mono-, di- & triglycerides); pp. 174-178 (total tocopherols).

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